

Protective effects of troxerutin on maternal high-fat diet-induced impairments of spatial memory and apelin in the male offspring

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ARTICLE INFO

Article type:

Original article

Article history:

Received: Dec 17, 2017

Accepted: Mar 15, 2018

Keywords:

Apelin
Maternal high-fat diet
Memory
Morris water maze
Troxerutin

ABSTRACT

Objective(s): Maternal high-fat diet (HFD) is linked with metabolic and cognitive deficits in offspring. Neuroprotective effects of troxerutin, a natural bioflavonoid, have been reported recently. This study aimed to investigate the effects of troxerutin on spatial memory and serum and hippocampal apelin levels in the male offspring of HFD fed mothers.

Materials and Methods: Three-week-old female Wistar rats (n= 40) received HFD or control diet (CD) for 8 weeks. After mating, pregnant animals were divided into two subgroups according to the troxerutin (TRO) supplementation: CD, CD+TRO, HFD, and HFD+TRO. HFD continued to the end of lactation in HFD and HFD+TRO groups. TRO was gavaged (150 mg/kg/day) during pregnancy. After weaning, the male offspring were fed a normal diet until 12 weeks of age. Spatial memory was evaluated in the Morris water maze (MWM) on postnatal day (PND) 90. Total apelin concentration was measured in the serum of maternal rats before mating and after lactation and also in the serum and hippocampus of their male offspring.

Results: Both traveled distance ($P<0.05$) and time spent ($P<0.05$) in the target quadrant were significantly decreased in the offspring of HFD-fed dams, which were reversed by TRO treatment. Moreover, TRO significantly ($P<0.05$) decreased serum apelin levels in dams. Furthermore, TRO treatment in dams significantly ($P<0.05$) increased serum and hippocampal levels of apelin in their offspring.

Conclusion: These results indicated that TRO treatment during pregnancy improved maternal HFD-induced spatial memory impairments of the offspring possibly through modulation of serum and hippocampal apelin levels.

► Please cite this article as:

Diba R, Mohaddes G, Mirzaie Babil F, Farajdokht F, Bayandor P, Hosseindoost M, Mehri K, Zavvari Oskuye Z, Babri Sh. Protective effects of troxerutin on maternal high-fat diet-induced impairments of spatial memory and apelin in the male offspring. Iran J Basic Med Sci 2018; 21:682-687. doi: 10.22038/IJBMS.2018.28170.6901.

Introduction

Maternal nutrition during pregnancy and lactation influence the health of mother and child (1). Maternal high-fat diet (HFD) is linked to the high risk of developing metabolic disorders including chronic liver disease, diabetes, high blood pressure, and obesity in offspring (2, 3). Maternal diet can also alter offspring behaviors and cognitive abilities by modifying the intrauterine environment and maternal behaviors (4, 5).

Offspring of obese mothers consuming HFD are vulnerable to mental and behavioral disorders including depression, anxiety, attention deficit hyperactivity disorder, and autism (5, 6). A possible mechanism of increased risk for behavioral disorders is alterations in neural pathways development involved in behavioral regulation (3). It has also been reported that intake of HFD during pregnancy is associated with learning impairments and memory loss in the adult offspring (6, 7).

Exact mechanisms explaining the link between HFD during pregnancy and memory disorders in infants are not known. Recent findings indicate factors such as

the type of nutrient intake (sugar and fat), hormones (insulin and leptin), and inflammatory cytokines affect fetal brain development (3). Development of neurotransmitter systems is also influenced by the levels of circulating cytokines (8, 9). Cytokines also have direct detrimental effects on hippocampal circuitry and cognition (10).

Apelin, as an adipokine, is secreted by adipocytes. It was initially isolated in 1998 from bovine stomach and defined as an endogenous ligand for an orphan G-protein coupled receptor, APJ (11). Apelin and its receptors are broadly distributed in the peripheral tissues and central nervous system (CNS) particularly in the hippocampus and amygdala, brain regions involved in learning and memory, and hypothalamus (11-13). Expression of apelin is influenced by nutritional status and decreased by fasting and recovered by refeeding (14). Apelin is considered as an anti-obesity peptide which increases the sensitivity of various tissues especially skeletal muscles to insulin, enhances energy consumption, and reduces body fat mass (15, 16). Moreover, apelin has

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important roles in the regulation of immune response (17), hemodynamic and body fluid homeostasis, angiogenesis, oxidative stress-linked atherosclerosis, and brain signaling (18). It is also reported that apelin is involved in the regulation of memory formation (19, 20), and its active form protects the nervous system from different diseases such as Parkinson's disease, Alzheimer's disease, and Huntington's disease (21).

Troxerutin, a natural bioflavonoid rutin, is present in tea, coffee, cereal grains, and different types of fruits and vegetables (22). Previous studies revealed that it has multiple biological properties including anti-neoplastic, antioxidant, and anti-inflammatory effects (22-25). Our previous studies demonstrated that troxerutin reduces oxidative stress markers, improves spatial learning and memory, and ameliorates synaptic plasticity of dentate gyrus neurons in an amyloid-beta induced rat model of Alzheimer's disease (26-28).

To better understand the impact of maternal HFD on offspring behavior, the effect of troxerutin treatment on HFD-induced spatial learning and memory impairments, as well as serum and hippocampal apelin levels in offspring was evaluated.

Materials and Methods

Animals and treatments

The present study was carried out at Tabriz University of Medical Sciences, Tabriz, Iran from March 2016 to Jun 2017. All procedures and experiments were carried out in accordance with the regulations of Tabriz University of Medical Sciences Ethical Committee for the Protection of Animals in Research. Forty virgin female Wistar rats (3 weeks old) were obtained from the colony of Tabriz University of Medical Sciences. Animals were kept in standard conditions, three per cage in a temperature (22 °C–25 °C) and humidity (40–60%) controlled room with a 12 hr light/12 hr dark cycle. Food and water were provided *ad libitum* except during the behavioral tests.

Animals were randomly divided into two groups: HFD group, which was fed a high-fat diet (52.0% lipids, 27.1% protein, 20.9% carbohydrate; 4.07 kcal/g) and CD group, which was fed a standard rat chow diet (14.7% lipids, 33.0% protein; 52.2 % carbohydrate; 2.75 Kcal/g) purchased from Behparvar CO, Tehran, Iran. Both groups were fed for 8 weeks before mating. All twenty female animals in each group were housed with adult males overnight for mating, and pregnancy was verified by examining vaginal smears for the presence

of vaginal plugs.

Then pregnant animals of each group were divided into two subgroups as follows: CD, CD+TRO, HFD, and HFD+TRO. Animals in the CD and HFD groups received saline as the vehicle of troxerutin; however, rats in the CD+TRO and HFD+TRO groups were treated with troxerutin (150 mg/kg/day, oral gavage) (29) during the pregnancy. HFD continued to the end of lactation in the HFD and HFD+TRO groups. Troxerutin (TRO) was acquired from Merck (Germany) and all other chemicals and reagents were obtained from commercial sources of the highest quality available. At the end of lactation, male offspring were separated from their mothers and kept in separate cages (4 per cage) in their respective groups. All offspring were fed a normal diet until postnatal day (PND) 90. The protocol of the study is shown in Figure 1.

Morris water maze test

Animals were transferred to the behavioral test room 30 min before the test. Spatial memory was tested on PND 90 between 10 a.m. and 2 p.m using the MWM apparatus, which was a black circular pool with a diameter of 130 cm and height of 80 cm, filled with water (23 ± 2 °C) to a depth of 60 cm. The maze was divided into four equal quadrants, and the release points in the quadrants were designated as N, E, S, and W. A hidden Plexiglas platform (10 × 10 cm) was placed in the center of the NW quadrant submerged 1.5 cm below the water surface. Fixed positioned distinctive visual cues were placed at several locations around the maze for facilitating spatial orientation of the animal and kept unchanged throughout the experiments.

On the first day, the training session was performed in two subsequent blocks (four trials in each block and 5 min intervals between the blocks) with four different starting points. The task required animals to swim to the hidden platform guided by the visual cues. The time taken to reach the platform and the traveled distance were measured. Twenty four hours after the last session, rats were subjected to a probe trial which consisted of a 60 sec free-swimming period without a platform. The traveled distance and time spent by animals in search of the platform in the target quadrant were measured. In order to assess the effect of animals' sensory and motor coordination or the animals' motivation, the capability of rats to escape to a visible platform was tested 5 min

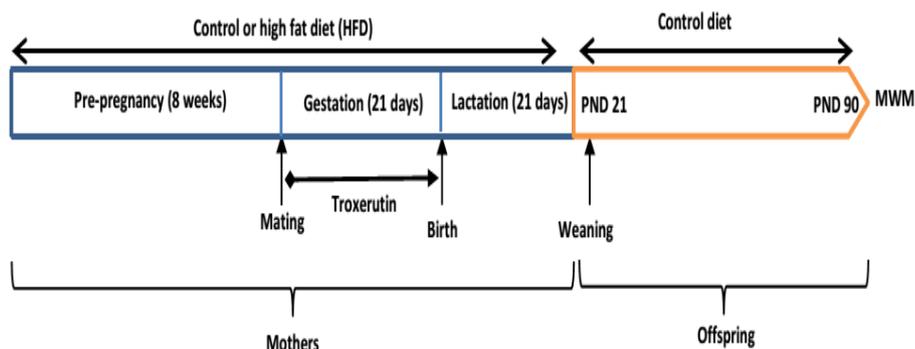


Figure 1. The protocol of the study. PND: postnatal day; MVM: Morris water maze

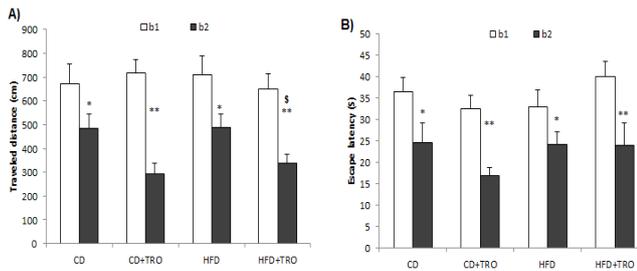


Figure 2. The effect of maternal high-fat diet (HFD) and troloxerutin (TRO) treatment on traveled distance (A) and escape latency (B) to find the hidden platform in the offspring groups. Each block represents the average of four consecutive trials. Paired sample *t*-test; * $P < 0.05$ and ** $P < 0.01$ indicate the difference between block 1 (b1) and block 2 (b2) in each group. \$ $P < 0.05$ indicates the difference between HFD+TRO and HFD groups in block 2. Data are expressed as mean \pm SEM ($n = 7$). [CD: control diet; TRO: troloxerutin; HFD: high fat diet]

after probe tests (30).

ELISA assays

At the end of behavioral testing, animals were deeply anesthetized by intraperitoneal injection of ketamine (60 mg/kg) and xylazine (5 mg/kg). Blood samples were collected from the heart and then centrifuged at 4000 rpm for 10 min. Serum samples were separated and kept at -80°C until analysis. After decapitation, the brain was rapidly excised and the hippocampus immediately dissected on an ice-cold dissection board. Isolated hippocampal tissues were kept at -80°C for later measurements. In order to measure total apelin levels, hippocampal samples were homogenized in 1.15% KCl solution, then homogenates were centrifuged (5000 rpm for 10 min at 4°C) and the supernatants were removed. The total apelin levels in the serum and hippocampal samples were determined using rat-specific apelin (nonspecific sensitivity for other factors) enzyme-linked immunosorbent assay (ELISA) kit (EASTBIOPHARM, China) in accordance with the manufacturer's assay protocol.

Statistical analysis

All results are expressed as mean \pm SEM for seven animals and analyses were performed using SPSS statistical software version 16. The paired Student's *t*-test was used in each group for comparison between blocks 1 and 2, and comparing total serum apelin levels of animals before pregnancy and after lactation in

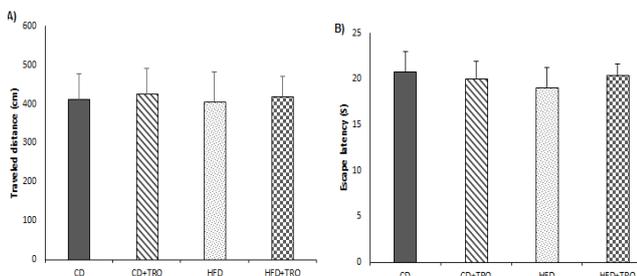


Figure 4. The effect of maternal HFD and troloxerutin on the traveled distance (A) and time spent in the target quadrant (B) in the visible platform test in the offspring groups. Data are expressed as mean \pm SEM ($n = 7$). There were no significant differences among the four groups. [CD: control diet; TRO: troloxerutin; HFD: high fat diet]

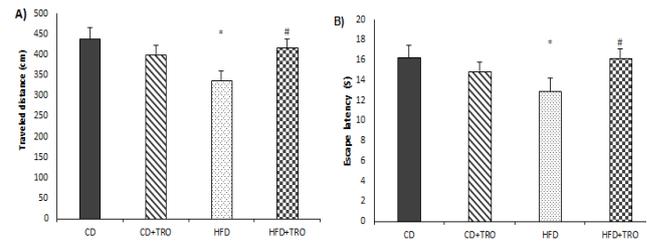


Figure 3. The effect of maternal HFD and troloxerutin on the traveled distance (A) and time spent in the target quadrant (B) of the offspring groups in the probe test. Data are expressed as mean \pm SEM ($n = 7$). One-way ANOVA followed by LSD post hoc test; * $P < 0.05$ vs. CD group, # $P < 0.05$ vs. HFD group. [CD: control diet; TRO: troloxerutin; HFD: high fat diet]

mothers groups. For other comparisons in the offspring groups, one-way ANOVA followed by LSD post hoc test was used. In all cases, *P*-values less than 0.05 were considered statistically significant.

Results

Effects of HFD and troloxerutin on cognitive performance of offspring in the Morris water maze

Our results showed that during the training sessions, the performance of all groups improved as indicated by shorter traveled distance and escape latency. The results of paired *t*-test for traveled distance revealed significant differences between two blocks ($t = 8.24$, $df = 27$) in the offspring of CD ($P < 0.05$), CD+TRO ($P < 0.01$), HFD ($P < 0.05$), and HFD+TRO ($P < 0.01$) groups (Figure 2A). Moreover, the results of escape latency showed significant differences between two blocks ($t = 7.61$, $df = 27$) in the offspring of CD ($P < 0.05$), CD+TRO ($P < 0.01$), HFD ($P < 0.05$), and HFD+TRO ($P < 0.01$) groups (Figure 2B).

Furthermore, the results of one-way ANOVA in block 2 showed that TRO treatment during gestation significantly ($P < 0.05$) decreased traveled distance in the HFD+TRO group as compared to the HFD group.

One-way analysis of ANOVA in probe test showed that traveled distance ($F_{(3,24)} = 2.98$, $P < 0.05$) and time spent in target quadrant ($F_{(3,24)} = 3.92$, $P < 0.05$) to find the hidden platform (Figures 3 A and B, respectively) in the HFD

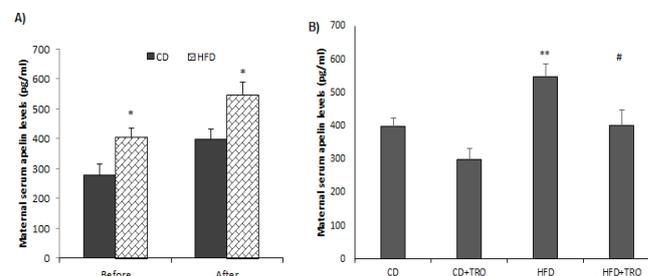


Figure 5. (A) Total serum maternal apelin levels before and after pregnancy. Paired sample *t*-test; * $P < 0.05$ indicates the difference in the CD groups and HFD groups before and after pregnancy. (B) The effect of HFD and troloxerutin on serum total apelin levels of dams at the end of lactation. One-way ANOVA followed by LSD post hoc test; ** $P < 0.01$ vs. CD group; # $P < 0.05$ vs. HFD group. Data are expressed as mean \pm SEM ($n = 7$). [CD: control diet; TRO: troloxerutin; HFD: high fat diet]

offspring were significantly lower than the CD group, indicating memory impairment in this group. However, daily treatments with TRO significantly ($P<0.05$) increased traveled distance and time spent in the target quadrant in the HFD+TRO group compared to the HFD received group, indicating memory improvement in this group.

Furthermore, there were no significant differences in the traveled distance and escape latency to find the visible platform among all the offspring groups. These results suggest that HFD had no effect on sensorimotor and motivational aspects, indicating impairment in spatial memory is due to cognitive deficits (Figures 4A and B).

Effects of HFD and troloxerutin on maternal total serum apelin levels

To determine the effects of HFD on maternal endogenous apelin levels, serum concentration of apelin was determined before pregnancy and after pregnancy (Figure 5A), and at the end of lactation (Figure 5B). Our findings demonstrated that serum levels of total apelin in both CD and HFD dams after pregnancy were significantly ($P<0.05$) higher than before pregnancy. Moreover, one-way ANOVA analysis showed that serum apelin levels in the CD group were significantly ($P<0.05$) lower than in the HFD group at the end of lactation. Daily TRO administration (150 mg/kg) during pregnancy could significantly decrease total serum apelin levels in the CD+TRO ($P<0.05$) and HFD+TRO ($P<0.05$) groups in comparison with CD and HFD groups, respectively (Figure 5B).

Effects of HFD and troloxerutin on serum and hippocampal total apelin levels of the offspring

According to data analysis, HFD offspring groups showed significantly lower serum ($F_{(3, 24)} = 2.25$, $P<0.05$) and hippocampal ($F_{(3, 24)} = 2.44$, $P<0.05$) total apelin levels in comparison with the corresponding CD offspring group (Figures 6A and B). However, daily TRO treatments significantly ($P<0.05$) increased serum and hippocampal total apelin levels in the HFD+TRO offspring group as compared to the HFD group.

Discussion

In the present study, it was demonstrated that maternal HFD prior to breeding and throughout

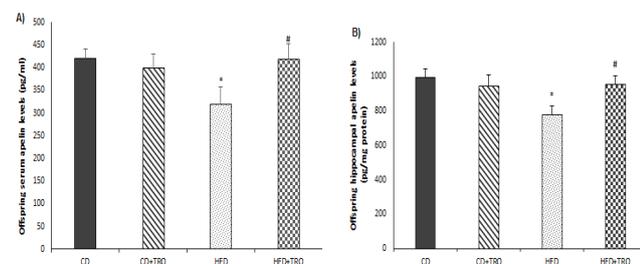


Figure 6. The effect of maternal HFD and troloxerutin on serum (A) and hippocampal (B) total apelin levels of offspring groups (PND=90). One-way ANOVA followed by LSD post hoc test; * $P<0.05$ vs. CD group, # $P<0.05$ vs. HFD group. Data are expressed as mean \pm SEM (n=7). [CD: control diet; TRO: troloxerutin; HFD: high fat diet]

gestation and lactation could have deleterious effects on spatial memory of the offspring. Moreover, HFD decreased serum and hippocampal apelin levels in the offspring. However, chronic treatment with TRO during pregnancy and lactation improved spatial memory and increased apelin levels in the offspring of HFD fed mothers.

Previous studies indicated that HFD prior to breeding in female rats led to obesity and hyperlipidemia in newborn pups, which was associated with oxidative stress and reduced hippocampal neurogenesis (31, 32). Moreover, HFD stimulates stress signaling cascades which can damage the neurodevelopment of offspring (33). Several experimental studies have also demonstrated that chronic HFD is associated with cognitive decline (32, 34). White *et al.* have shown that maternal and offspring HFD consumption increases IL-6, a pro-inflammatory cytokine, in the cortex (32). In the present study, maternal HFD impaired the spatial memory of offspring in the MWM task. Similarly, several studies reported impaired spatial memory in the male offspring of obese mothers in the Barnes maze and MWM tests (4, 32, 35, 36). In another study, young adult male offspring exposed to maternal HFD showed a decreased exploration in the novel object recognition test (37). In contrast to our results, Bilbo and Tsang (38) showed that adult male and female rats born to dams fed HFD performed better on the MWM test compared to the control animals. Different impacts of HFD on memory might be due to different experimental animals or HFD models, age of animals at assessment, and cognitive test paradigms.

In the present study, oral administration of TRO (150 mg/kg) for 21 days in the HFD pregnant animals effectively protected their offspring against HFD-induced impairment of learning and memory. This finding is consistent with recent studies which demonstrated that TRO attenuates cognitive impairment and oxidative stress induced by administration of amyloid- β or D-galactose in the hippocampus of rats treated with high cholesterol diet (25, 27, 39). Moreover, a recent study has reported that TRO promotes learning and memory in streptozotocin-induced type 1 diabetes by inhibiting oxidative stress response (40).

Our results also revealed that HFD fed mothers exhibited high levels of apelin in the serum after pregnancy and at the end of lactation as compared to the CD group. It has been demonstrated that nutritional status and insulin-dependent mechanisms of apelin secretion may also have effects on apelin concentration (14). Exposure to high levels of fatty acids can reduce the sensitivity of cells to insulin and induce hyperinsulinemia, which in turn increases apelin secretion from the adipose tissue (41). Zhang *et al.* also demonstrated that HFD feeding increased fasting blood glucose and serum insulin levels in mice (42). Therefore, increased serum apelin levels in the HFD dams could be explained by this mechanism. On the other hand, we found that administration of TRO during gestation decreased apelin levels in the mother's serum. In support of our finding, evidence shows that TRO has a restoring effect on adipokine protein expression in the liver of HFD-treated rats (43). Recently, a study also reported that TRO inhibits inflammatory cytokines release and

declines adiponectin levels in the HFD-received mice. Furthermore, TRO treatment reduces serum insulin levels in the HFD-treated mice (42). It has also been reported that TRO can prevent obesity by improving insulin signaling pathway, and it returns blood glucose, fatty acids, and cholesterol levels to the normal levels (44). Therefore, it is likely that TRO through these mechanisms reduced maternal apelin levels.

Here, we also showed that offspring's apelin levels in the HFD group decreased as a result of maternal HFD consumption which was increased in the HFD+TRO group. Researchers (45) reported that in utero HFD exposure changes adipocytokine promoter epigenetic and leads to abnormal adipocytokine levels in the offspring. Therefore, it is likely that maternal HFD modifies post-translational processes and cellular mechanisms involved in the apelin release. Moreover, maternal HFD augments hypothalamus-pituitary axis (HPA) activity and increases corticosterone levels in their offspring (46). Previous studies also showed that glucocorticoids suppress apelin gene expression in adipocytes in a dose-dependent manner which reduces apelin production (47, 48). Therefore, it seems that high cortisol levels of the HFD-fed offspring group have suppressed the apelin production in the HFD offspring, and TRO has increased apelin levels in the offspring through suppression of cortisol.

Apelin and its receptors are widely distributed in the brain regions involved in learning and memory functions, demonstrating that apelin may participate in the regulation of memory processes (12, 13, 49). In this study, low serum and hippocampal apelin levels were allied with spatial memory impairment in the HFD offspring suggesting that low apelin levels are probable mediators of HFD-induced cognitive disruption. A study (20) showed that intracerebroventricular (ICV) administration of apelin-13 blocks short-term memory formation and long-term memory consolidation in the novel object recognition task. Nevertheless, researchers (50) reported that administration of apelin-13 (2 µg) facilitates learning and memory consolidation in the passive avoidance learning paradigm. Likewise, a recent study (51) reported that central injection of apelin improved memory in the novel object recognition test in the stressed rats.

Furthermore, elevation of the serum and hippocampal apelin levels was accompanied with improved memory of the HFD+TRO offspring group. Anti-inflammatory and antioxidant properties of TRO have also been proven in several studies (22-24). Therefore, it seems that chronic TRO treatment improves spatial memory through increasing the apelin levels and decreasing the pro-inflammatory cytokines in the hippocampus.

Conclusion

TRO reversed cognitive deficits in the adult male offspring of HFD fed mothers. This effect may be partly mediated by modification of apelin levels. Further studies are needed to elucidate the exact mechanism of TRO action in this regard.

Acknowledgment

The results presented in this paper were part of

a student thesis which was supported by the Drug Applied Research Center of Tabriz University of Medical Sciences, Tabriz, Iran (grant No. 33/94). The authors would like to thank Dr Saeed Sadigh-Eteghad for his helpful assistance in the laboratory.

References

- Mecacci F, Biagioni S, Ottanelli S, Mello G. Nutrition in pregnancy and lactation: how a healthy infant is born. *J Pediatr Neonat Individual Med* 2015; 4:e040236.
- Barker DJ. Fetal nutrition and cardiovascular disease in later life. *Br Med Bull* 1997; 53:96-108.
- Sullivan EL, Nousen EK, Chamlou KA. Maternal high fat diet consumption during the perinatal period programs offspring behavior. *Physiol Behav* 2014; 123:236-242.
- Tozuka Y, Kumon M, Wada E, Onodera M, Mochizuki H, Wada K. Maternal obesity impairs hippocampal BDNF production and spatial learning performance in young mouse offspring. *Neurochem Int* 2010; 57:235-247.
- Arnold SE, Lucki I, Brookshire BR, Carlson GC, Browne CA, Kazi H, et al. High fat diet produces brain insulin resistance, synaptodendritic abnormalities and altered behavior in mice. *Neurobiol Dis* 2014; 67:79-87.
- Boitard C, Cavaroc A, Sauvart J, Aubert A, Castanon N, Layé S, et al. Impairment of hippocampal-dependent memory induced by juvenile high-fat diet intake is associated with enhanced hippocampal inflammation in rats. *Brain Behav Immun* 2014; 40:9-17.
- Lu J, Wu D-m, Zheng Y-l, Hu B, Cheng W, Zhang Z-f, et al. Ursolic acid improves high fat diet-induced cognitive impairments by blocking endoplasmic reticulum stress and IκB kinase β/nuclear factor-κB-mediated inflammatory pathways in mice. *Brain Behav Immun* 2011; 25:1658-1667.
- Miller AH, Haroon E, Raison CL, Felger JC. Cytokine Targets in the Brain: Impact on Neurotransmitters and Neurocircuits. *Depress Anxiety* 2013; 30:297-306.
- Das U. Is obesity an inflammatory condition? *Nutrition* 2001; 17:953-966.
- Freeman LR, Haley-Zitlin V, Rosenberger DS, Granholm A-C. Damaging effects of a high-fat diet to the brain and cognition: A review of proposed mechanisms. *Nutr Neurosci* 2014; 17:241-251.
- Tatemoto K, Hosoya M, Habata Y, Fujii R, Kakegawa T, Zou M-X, et al. Isolation and characterization of a novel endogenous peptide ligand for the human APJ receptor. *Biochem Biophys Res Commun* 1998; 251:471-476.
- Lee DK, Cheng R, Nguyen T, Fan T, Kariyawasam AP, Liu Y, et al. Characterization of apelin, the ligand for the APJ receptor. *J Neurochem* 2000; 74:34-41.
- Medhurst AD, Jennings CA, Robbins MJ, Davis RP, Ellis C, Winborn KY, et al. Pharmacological and immunohistochemical characterization of the APJ receptor and its endogenous ligand apelin. *J Neurochem* 2003; 84:1162-1172.
- Boucher J, Masri B, Daviaud D, Gesta S, Guigné C, Mazzucotelli A, et al. Apelin, a newly identified adipokine up-regulated by insulin and obesity. *Endocrinol* 2005; 146:1764-1771.
- Bertrand C, Valet P, Castan-Laurell I. Apelin and energy metabolism. *Front Physiol* 2015; 6:115.
- Castan-Laurell I, Dray C, Knauf C, Kunduzova O, Valet P. Apelin, a promising target for type 2 diabetes treatment? *Trends Endocrinol Metab* 2012; 23:234-241.
- Falcão-Pires I, Leite-Moreira AF. Apelin: a novel neurohumoral modulator of the cardiovascular system. Pathophysiologic importance and potential use as a therapeutic target. *Revista Portuguesa de Cardiologia* 2005;

- 24:1263-1276.
18. Galanth C, Hus-Citharel A, Li B, Llorens-Cortes C. Apelin in the control of body fluid homeostasis and cardiovascular functions. *Curr Pharm Des* 2012; 18:789-798.
19. Han L, Luo H, Huang F, Tian S, Qin X. Apelin-13 Impaired Acquisition but Not Consolidation or Expression of Contextual Fear in Rats. *Neurochem Res* 2016; 41:2345-2351.
20. Han R-w, Xu H-j, Zhang R-s, Wang R. The role of apelin-13 in novel object recognition memory. *Peptides* 2014; 62:155-158.
21. Cheng B, Chen J, Bai B, Xin Q. Neuroprotection of apelin and its signaling pathway. *Peptides* 2012; 37:171-173.
22. Fan S-h, Zhang Z-f, Zheng Y-l, Lu J, Wu D-m, Shan Q, et al. Troloxerutin protects the mouse kidney from d-galactose-caused injury through anti-inflammation and anti-oxidation. *Int Immunopharmacol* 2009; 9:91-96.
23. Thomas NS, George K, Arivalagan S, Mani V, Siddique AI, Namasivayam N. The in vivo antineoplastic and therapeutic efficacy of troloxerutin on rat preneoplastic liver: biochemical, histological and cellular aspects. *Eur J Nutr* 2016.
24. Badalzadeh R, Layeghzadeh N, Alihemmati A, Mohammadi M. Beneficial effect of troloxerutin on diabetes-induced vascular damages in rat aorta: histopathological alterations and antioxidation mechanism. *Int J Endocrinol Metab Disord* 2015; 13:e25969.
25. Liu C-M, Ma J-Q, Lou Y. Chronic administration of troloxerutin protects mouse kidney against D-galactose-induced oxidative DNA damage. *Food Chem Toxicol* 2010; 48:2809-2817.
26. Babri S, Amani M, Mohaddes G, Alihemmati A, Ebrahimi H. Protective effects of troloxerutin on β -Amyloid (1-42)-induced impairments of spatial learning and memory in rats. *Neurophysiol* 2012; 44:387-393.
27. Babri S, Mohaddes G, Feizi I, Mohammadnia A, Niapour A, Alihemmati A, et al. Effect of troloxerutin on synaptic plasticity of hippocampal dentate gyrus neurons in a β -amyloid model of Alzheimer's disease: An electrophysiological study. *Eur J Pharmacol* 2014; 732:19-25.
28. Farajdokht F, Amani M, Bavil FM, Alihemmati A, Mohaddes G, Babri S. Troloxerutin protects hippocampal neurons against amyloid beta-induced oxidative stress and apoptosis. *EXCLI J* 2017; 16:1081-1089.
29. Lu J, Wu D-m, Hu B, Cheng W, Zheng Y-l, Zhang Z-f, et al. Chronic administration of troloxerutin protects mouse brain against d-galactose-induced impairment of cholinergic system. *Neurobiol Learn Mem* 2010; 93:157-164.
30. Tabari S-s, Babri S, Mirzaie F, Farajdokht F, Mohaddes G. Enduring amnesia induced by ICV scopolamine is reversed by sesame oil in male rats. *Acta Cirurgica Brasileira* 2016; 31:520-526.
31. Tozuka Y, Wada E, Wada K. Diet-induced obesity in female mice leads to peroxidized lipid accumulations and impairment of hippocampal neurogenesis during the early life of their offspring. *FASEB J* 2009; 23:1920-1934.
32. White CL, Pistell PJ, Purpera MN, Gupta S, Fernandez-Kim S-O, Hise TL, et al. Effects of high fat diet on Morris maze performance, oxidative stress, and inflammation in rats: contributions of maternal diet. *Neurobiol Dis* 2009; 35:3-13.
33. Niculescu MD, Lupu DS. High fat diet-induced maternal obesity alters fetal hippocampal development. *Int J Dev Neurosci* 2009; 27:627-633.
34. Greenwood CE, Winocur G. High-fat diets, insulin resistance and declining cognitive function. *Neurobiol Aging* 2005; 26:42-45.
35. Page KC, Jones EK, Anday EK. Maternal and postweaning high-fat diets disturb hippocampal gene expression, learning, and memory function. *Am J Physiol Regul Integr Comp Physiol* 2014; 306:R527-R537.
36. Lépinay AL, Larriou T, Joffre C, Acar N, Gárate I, Castanon N, et al. Perinatal high-fat diet increases hippocampal vulnerability to the adverse effects of subsequent high-fat feeding. *Psychoneuroendocrinol* 2015; 53:82-93.
37. Graf AE, Lallier SW, Waidyaratne G, Thompson MD, Tipple TE, Hester ME, et al. Maternal high fat diet exposure is associated with increased hepcidin levels, decreased myelination, and neurobehavioral changes in male offspring. *Brain Behav Immun* 2016; 58:369-378.
38. Bilbo SD, Tsang V. Enduring consequences of maternal obesity for brain inflammation and behavior of offspring. *FASEB J* 2010; 24:2104-2115.
39. Lu J, Wu D-m, Zheng Y-l, Hu B, Cheng W, Zhang Z-f, et al. Troloxerutin counteracts domoic acid-induced memory deficits in mice by inhibiting CCAAT/enhancer binding protein β -mediated inflammatory response and oxidative stress. *J Immunol* 2013; 190:3466-3479.
40. Zhang S, Li H, Zhang L, Li J, Wang R, Wang M. Effects of troloxerutin on cognitive deficits and glutamate cysteine ligase subunits in the hippocampus of streptozotocin-induced type 1 diabetes mellitus rats. *Brain Res* 2017; 1657:355-360.
41. Kabarhan S, Besler HT. Do fatty acids affect fetal programming? *Health Popul Nutr* 2015; 33:14-23.
42. Zhang Z, Wang X, Zheng G, Shan Q, Lu J, Fan S, et al. Troloxerutin attenuates enhancement of hepatic gluconeogenesis by inhibiting NOD activation-mediated inflammation in high-fat diet-treated mice. *Int J Mol Sci* 2017; 18:31.
43. Zhang Z-F, Fan S-H, Zheng Y-L, Lu J, Wu D-M, Shan Q, et al. Troloxerutin improves hepatic lipid homeostasis by restoring NAD⁺-depletion-mediated dysfunction of lipin 1 signaling in high-fat diet-treated mice. *Biochem Pharmacol* 2014; 91:74-86.
44. Vinothkumar R, Kumar RV, Sudha M, Viswanathan P, Balasubramanian T, Nalini N. Modulatory effect of troloxerutin on biotransforming enzymes and preneoplastic lesions induced by 1, 2-dimethylhydrazine in rat colon carcinogenesis. *Exp Mol Pathol* 2014; 96:15-26.
45. Masuyama H, Mitsui T, Eguchi T, Tamada S, Hiramatsu Y. The effects of paternal high-fat diet exposure on offspring metabolism with epigenetic changes in the mouse adiponectin and leptin gene promoters. *Am J Physiol Endocrinol Metab* 2016; 311:E236-E245.
46. D'Asti E, Long H, Tremblay-Mercier J, Grajzer M, Cunnane SC, Di Marzo V, et al. Maternal dietary fat determines metabolic profile and the magnitude of endocannabinoid inhibition of the stress response in neonatal rat offspring. *Endocrinol* 2010; 151:1685-1694.
47. Wei L, Hou X, Tatemoto K. Regulation of apelin mRNA expression by insulin and glucocorticoids in mouse 3T3-L1 adipocytes. *Regul Pept* 2005; 132:27-32.
48. Henley DE, Buchanan F, Gibson R, Douthwaite JA, Wood SA, Woltersdorf WW, et al. Plasma apelin levels in obstructive sleep apnea and the effect of continuous positive airway pressure therapy. *J Endocrinol* 2009; 203:181-188.
49. Hosoya M, Kawamata Y, Fukusumi S, Fujii R, Habata Y, Hinuma S, et al. Molecular and functional characteristics of APJ tissue distribution of mRNA and interaction with the endogenous ligand apelin. *J Biol Chem* 2000; 275:21061-21067.
50. Telegdy G, Adamik A, Jászberényi M. Involvement of neurotransmitters in the action of apelin-13 on passive avoidance learning in mice. *Peptides* 2013; 39:171-174.
51. Li E, Deng H, Wang B, Fu W, You Y, Tian S. Apelin-13 exerts antidepressant-like and recognition memory improving activities in stressed rats. *Eur Neuropsychopharmacol* 2016; 26:420-430.