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Vitamin B12 and estradiol benzoate improve memory retrieval through activation of the hippocampal AKT, BDNF, and CREB proteins in a rat model of multiple sclerosis

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ARTICLEINFO	A B S T R A C T
<i>Article type:</i> Original article	<i>Objective(s):</i> Multiple sclerosis (MS) causes extensive damage in the hippocampus. Vitamin B12 (vit B12) and estradiol benzoate (EB) have anti-inflammatory and re-myelination properties that make them proper in improvement of cognitive impairment. This study aimed to evaluate the effects of these compounds on learning and memory disturbances. <i>Materials and Methods:</i> 77 adult male rats were implanted with stainless steel guide cannula bilaterally
<i>Article history:</i> Received: Aug 24, 2020 Accepted: Dec 14, 2020	
<i>Keywords:</i> Estradiol benzoate Hippocampus Memory Multiple sclerosis Vitamin B12	into the hippocampal area. The animals received 3 µl intrahippocampal EtB 0.01% and were randomly divided into eleven groups (7 rats/group). The groups included control, peanut oil (sham1), distilled water (sham 2), vit B12 (0.25, 0.5, 1 mg/kg), EB (25 and 50 mg/kg), vit B12 (0.25 mg/kg) plus EB (25 mg/kg), vit B12 (0.5 mg/kg) plus EB (25 mg/kg), and vit B12 (1 mg/kg) plus EB (50 mg/kg). The control group received intrahippocampal saline (as solvent). The locomotor activity and learning and memory functions were evaluated by open-field and shuttle-box tests, respectively. AKT, CREB, and BDNF levels were analyzed by Western blotting.
	Results: This study has found significant deficit in passive avoidance learning, locomotor activity, as well as decrease in the levels of phosphorylated AKT, BDNF, and CREB in groups that received EtB. Vit B12 (1 mg/kg), EB (50 mg/kg), and their combination markedly improved these side effects.
	Conclusion: This study demonstrated that vit B12 and estradiol benzoate, especially in combination therapy, can be helpful in treatment of memory problems and MS-induced dysfunction through activation of the hippocampal AKT, BDNF, and CREB proteins.

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Introduction

Multiple sclerosis (MS) is a multifocal inflammatory and demyelinating disorder that grossly affects the central nervous system (CNS) resulting in progressive neurological impairment. MS causes extensive damage in the brain and spinal cord (1), including brain volume reduction, ventricular enlargement, and spinal cord atrophy (2). Approximately 2.5 million people around the world suffer from MS as the most debilitating neurological disease after trauma. MS is induced by various factors, such as immune disorders, viral infections, genetics, environmental factors (3), and metals such as iron (4). In fact, the immune system acts against the axonal myelin sheath and causes chronic disability (5).

Based on previous study, the most important symptom of MS is myelin destruction, especially the gray matter atrophy (6), which is associated with a patient's permanent disability (7). Oligodendrocyte precursor cells (OPCs), which are present in brain's white matter and around MS lesions, are responsible for axonal remyelination (8). Although endogenous re-myelination which may occur in MS patients, it is a relatively shortterm process, imperfect, and ineffective (8). It is also indicated that a decrease in activity of the antioxidant enzymes, such as glutathione peroxidase and catalase, play an important role in MS disease induction (9).

Animal models of MS are so important to evaluate the pathological mechanisms of MS and how they may be targeted for therapeutic intervention. Experimental autoimmune encephalomyelitis (EAE) is the most widely used animal model for MS and induced by injection of proteins, myelin peptides, and other substances, such as EtB, in mice, rats and monkeys (10). Direct injection of EtB into the brain is a simple and repeatable model that can offer the advantage of evaluating neuronal demyelination, re-myelination, and remodeling processes in the CNS (11).

The hippocampus is composed of several sub-regions with different histology and functional characteristics that make them susceptible to various diseases (12). One of the most important sites of injury in MS and EAE

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is hippocampus which plays a major role in memory formation and retrieval. Cognitive deficits, including impairment of information processing speed, attention, and learning and memory formation, are presented at more than 50% of MS patients (13).

Currently, disease-modifying therapies for MS are focusing on decreasing the inflammatory response and the severity of clinical symptoms. The main goal of MS treatment is to identify new drugs that enhance oligodendrocyte differentiation and myelin sheath formation as well as minimize axonal degeneration and inflammation, immune system activity, and the rate and degree of disease progression. With regard to immunomodulatory, anti-inflammatory, and neuroprotective effects of vit B12 and EB, they can be considered as a possible candidates for the treatment of MS (14).

Vitamin B12 (cobalamin) is a metal-organic compound the human body cannot synthesize and should be provided through diet, especially animal products, including meat and dairy products. The minimum human daily requirement for vit B12 is approximately 2.5 micrograms (15). Vit B12 absorption relies on two important factors, including intrinsic factor (aglycoprotein produced by the gastric parietal cells) and trans-cobalamin receptor (located in the mucosal cells of the ileum) (15). Vit B12, as a co-factor, plays a vital role in many body processes, such as hematopoiesis and myelin sheath production in the nervous system. Vit B12 deficiency induces inflammatory and neurodegenerative disorders, specially myelin sheath damage in the central and peripheral nervous systems (16).

Estriol, estradiol, and estrone are three main endogenous estrogens that exert their genomic and non-genomic effects through intracytoplasmic or nuclear receptors (ER α and β) and plasma membrane receptors (such as ER-X). The ER α and β have antiinflammatory and neuroprotective effects and are located in the subgranular zone (SGZ) of the hippocampal dentate gyrus (17). Estradiol (18) and estriol can influence hippocampal neurogenesis (19) and have neuroprotective effects on EAE via the ER α receptor. In addition, several studies have indicated that the effect of estrogens and estrogen receptor (ER) agonists on diverse signaling pathways, resulted in proliferation and differentiation of oligodendrocyte cells (20, 21). Estrogen treatment meaningfully diminishes the severity of EAE in numerous strains of mice and also decreases inflammatory immune response (19, 22) by inhibiting pro-inflammatory cytokines, chemokines, extracellular matrix proteins, antigens, and dendritic cell function (18, 22). Estrogen treatment also reduces white (23) and gray matter (24) inflammation, demyelination, and axonal destruction in the spinal cord of EAE (23).

Based on the aforementioned research background, this study was designed to investigate the effects of vit B12 and EB alone or in combination on memory function in a rat model of MS.

Materials and Methods

Chemicals

Vitamin B12 (cobalamin) (CAS # V2876), peanut oil (CAS # P2144), and estradiol benzoate (CAS # E8515) were purchased from Sigma-Aldrich (Germany). Ketamine hydrochloride and xylazine hydrochloride were provided by Merck (Rotexmeica, Germany).

Animals

In this experimental study, 77 adult male Wistar rats with an approximate weight of 230 to 250 g (two-month-old) were provided by the Animal Center, Department of Biology, Damghan Branch, Islamic Azad University, Damghan, Iran. All rats were maintained in standard conditions with a 12-hr light/dark cycle and at a temperature of 23 ± 1 °C, with free access to food and water. Each rat was used only once. All behavioral experiments were performed in the light cycle between 9:00 a.m. to 2:00 p.m. These conditions were constantly maintained during the experimental period and all animal experiment protocols were performed in accordance with Ethical Committee Acts of Damghan Branch, Islamic Azad University, Damghan, Iran (IR.DB, IAU.REC.1397.628).

Treatment

All rats were anesthetized with IP injection of ketamine and xylazine at dose of 50 and 5 mg/ kg, respectively. Anesthetized rats were placed on stereotaxic instrument in the skull-flat position for implantation of stainless steel guide cannula into the left and right hippocampal CA1 areas. After a recovery period of 5–7 days, 3 µl of 0.01% EtB in sterile 0.9% saline at the rate of 1 μ l/min was injected into the hippocampal CA1 area (25). Three days after injection, the rats were divided into 11 groups and each group consisted of 7 rats, including control, sham 1 (peanut oil), sham 2 (distilled water), groups 4-6 (vit B12 at doses of 0.25, 0.5, 1 mg/kg), groups 7 and 8 (EB at doses of 25 and 50 mg/kg), group 9 (vit B12 (0.25 mg/ kg) plus EB (25 mg/kg)), group 10 (vit B12 (0.5 mg/kg) plus EB (25 mg/kg)), and group 11 (vit B12 (1 mg/kg) plus EB (50 mg/kg)). The control group received intrahippocampal normal saline (as solvent). Distilled water (as vit B12 vehicle) and vit B12 were gavaged while peanut oil (as EB vehicle) and EB were administrated by intraperitoneal (IP) injection. Treatment duration was 5 days. At the end of the period, the behavioral tests, including open-field for locomotor activity and shuttle-box for learning and memory function, were evaluated. Then, the animals were anaesthetized via IP injection of ketamine and xylazine at doses of 50 and 5 mg/kg, respectively and the hippocampus tissues were removed and stored at -80 °C until protein extraction was performed. A schematic diagram showing the experimental study design is exhibited in Figure 1.

Stereotaxic procedure

The rats were anesthetized as mentioned above and then placed in a stereotaxic apparatus in a flat skull position (incisor bar \pm 3.3 mm and ear bars positioned symmetrically) (Stoelting Co., Illinois, USA). Subsequently, the scalp was shaved and sterilized and the skin incised on the mid line. In the next step, two holes were drilled in the skull and two stainless steel guide cannulas (22 gauge, 12 mm length, and 0.7 mm diameter) were implanted bilaterally, 1 mm above the dorsal hippocampus (CA-1) (coordinates: anteroposterior (AP) = -2 mm from the bregma point,

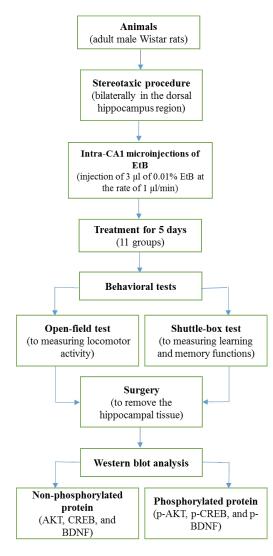


Figure 1. A schematic diagram of the experimental study design

media lateral (ML) = ± 1.6 mm from the sagittal suture on both sides, and dorsoventrally (DV) = -1.5 mm from the skull surface), according to rat brain atlas (Paxinos and Watson) (26). The position of guide cannulas and jewelers' screws, for more stability, were permanently fixed to the bone using acrylic dental cement. The stainless steel insect pins in the size of 27-gauge were entered into the guide cannula to reduce the incidence of occlusion. Finally, all rats were allowed 5–7 days for recovery (27).

Intra-MS/CA1 microinjections of drug

For drug microinjection, all rats were quietly restrained by hand, the stainless steel stylets removed from the guide cannula, and switched with dental 27-gauge injection needles (1 mm below the tip of the guide cannula). Next, the needle was connected to a 10-µl Hamilton micro syringes by a polyethylene tube (PE-20). Injection of 3 µl of 0.01% EtB in sterile 0.9% saline at the rate of 1 µl/min was bilaterally performed into the hippocampal CA1 region (25). During the injection period, the animal was freely allowed to move without any stress. The drugs were injected 5–7 days after surgery and carried out 5 min before behavioral evaluation (27).

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Passive avoidance training test

The passive avoidance test (PAT) is used to evaluate fear learning and memory in animal models of neurological disorders and subjects learning to avoid an unpleasant stimulus (such as a foot-shock). In this study, the PAT protocol was used according to Yousefi et al., 2012 (28) with little modification. Briefly, the PAT apparatus consisted of two compartments, including a light and a dark box (opaque resin, 20 cm × 20 cm × 30 cm) with a connecting open door (7 cm \times 9 cm). A parallel stainless steel grid at 2.5 mm diameter and 1 cm intervals was located on the floor of the dark box to generate foot shock (29). In the first step, habituation phase, rats were allowed to stay in the experimental room for 1 hr. In the next step, acquisition trial or training stage, animals were gently placed in the white box, the guillotine lid was opened, and the animals were allowed to enter the dark box where the single electric shock (50 Hz, 3 sec, and 1 mA intensity) (Borj Sanat Co, Tehran, Iran) is done on the animal's foot by an electrical generating device. Finally, the last step or the retrieval stage, was performed 24 hr after the training stage and during this phase the interval time between animal movements from light box into the dark box (stepthrough latency) was measured. The cut-off time of this step was 300 sec. All steps were manually implemented between 8:00 a.m. to 2:00 p.m.

Open-field test

The open-field test is carried out to measure behavioral responses, including locomotor activity, hyperactivity, and stereotypical and exploratory behaviors. Summary, each rat was placed in the center of the cage made of white wood (45 cm × 45 cm × 45 cm). The animal's movements were recorded using a digital camera for 5 min. The locomotor activity was evaluated by measuring the number of peripheral (those adjacent to the walls), central, and total square crossings; the three measures were referred to as peripheral (PL), central (CL), and total (TL) locomotion, respectively. At the end of each experiment, the open-field device was cleaned using a damp sponge and a dry paper towel (30).

Western blot analysis

The hippocampal region of the rat brain tissue was rapidly separated, frozen in liquid nitrogen, and kept at -80 °C. Protein analysis of protein kinase B (AKT), cAMP response element-binding protein (CREB), and brain-derived neurotrophic factor (BDNF), as well as their phosphorylation types were performed by Western blotting assays on protein extraction from hippocampus tissues. Approximately, 200 mg of each sample was homogenized in lysis buffer, including 50 mM Tris-HCL with pH 7.4, 2 mM EDTA, 2 mM EGTA, 10 mM NaF, 1 mM sodium orthovanadate (Na3VO4), 10 mM β -glycerol-phosphate, 0.2% W/V sodium deoxycholate, 1 mM phenyl methyl sulfonyl fluoride (PMSF), and protease inhibitor cocktail (1% protease and phosphatase inhibitor cocktail) (Sigma, P8340) by a Polytron homogenizer (POLYTRON-PT10-35, Kinematica, Switzerland) in ice and next centrifuged at 10,000 g for 10 min at 4° C (31, 32). Subsequently, supernatants were collected and the concentration of total protein, via the Bradford protein assay kit (Bio-Rad), was determined.

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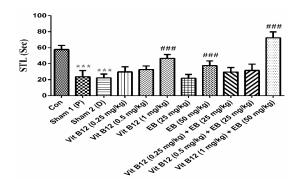


Figure 2. Effects of vit B12 (0.25, 0.5, and 1 mg/kg) and EB (25 and 50 mg/kg) alone or in combination therapy in passive avoidance test. Animals received EtB in sterile 0.9% saline at the rate of 1 µl/min into the hippocampal CA1 area. Three days after injection, the rats were randomly divided into 11 groups. Distilled water (as vit B12 vehicle) and vit B12 were gavaged and peanut oil (as EB vehicle) and EB were administrated by intraperitoneal injection. The control group received intra-hippocampal saline (as solvent). Treatment duration was 5 days. Data are shown as mean±SD, one-way ANOVA and Tukey–Kramer. ****P*<0.001 compared with the control group, and ****P*<0.001 compared with the sham groups

Con: Control; P: Peanut oil; D: Distilled water; Vit: Vitamin; EB: Estrogen benzoate; EtB: Ethidium bromide; STL: Step through latency (n=7)

Then, protein lysate aliquots were instantly denatured at 95 °C in sodium dodecyl sulfate (SDS)-Laemmli sample buffer, 2 mM DTT for 5 min, and stored at -80 °C. In summary, equal quantities of proteins (50 µg) were separated through 12% SDS-polyacrylamide gel (SDS-PAGE gel) and transferred to polyvinylidene difluoride (PVDF) membranes (Millipore). The PVDF membranes were blocked with 5% skim milk or %1 bovine serum albumin (BSA) in Tris-buffered saline plus 0.1% Tween 20 (TBST) for 2 hr at room temperature to prevent nonspecific protein binding. The membranes were incubated at 37 °C for 2 hr with primary rabbit monoclonal antibodies from Cell Signaling Technology (Beverly, MA, USA): anti-CREB (#4820), anti-BDNF (#3897), anti-AKT (#9272), anti-p-CREB (#9198), anti-p-BDNF (#47808), and anti-p-AKT (#9271) at a concentration of 1:1000 in TBST. After washing the membranes three

times with TBST buffer (3×5 min in TBST), they were exposed with the secondary antibodies (anti-rabbit IgG) conjugated with horseradish peroxidase enzyme (HRP) (cell signaling, #7074) 1:3000 in TBST for 90 min at room temperature. After washing three times again, PVDF membranes were separately exposed to enhanced chemiluminescence (ECL) reagent (Pierce ECL Western Blotting Substrate) and H₂O₂ for 3 min. The intensity of protein bands was analyzed by Alliance 4.7 Gel Doc (UK). The protein expression levels were normalized versus beta-actin (#4967) protein level.

Statistical analysis

All data are presented as mean±standard deviation (mean±SD) (n=7). The statistical analysis of mean values was done by one-way ANOVA followed by the Tukey-Kramer test using SPSS (ver. 16.0) software. P<0.05 was regarded as statistically significant.

Results

Effect of vitamin B12 and EB on passive avoidance learning and memory in EtB-induced multiple sclerosis

As shown in Figure 2, there was a significant difference in step through latency (STL) between the control and sham (lesion) groups (P<0.001). Vit B12 at a dose of 1 mg/kg and EB at a dose 50 mg/kg alone or in combination therapy markedly increased STL and promoted fear learning and memory in comparison with sham groups (F (10, 66) = 49.58 and P<0.001).

Effect of vitamin B12 and EB on locomotor activity in EtB-induced multiple sclerosis

According to the results obtained from locomotor activity, sham 1 and sham 2 groups had significantly reduced the peripheral (A), central (B), and total (C) locomotor activity in comparison with the control group (P<0.001). Vit B12 (1 mg/kg), EB (50 mg/kg), either alone or in combination significantly improved peripheral (A), central (B), and total (C) locomotor activity compared with the sham groups (P<0.05, F (10, 66) = 70.92 (A), 10.97 (B), 17.32 (C), and P<0.001).

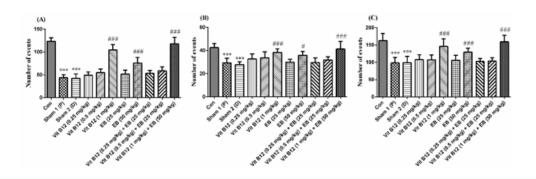


Figure 3. Effects of vit B12 (0.25, 0.5, and 1 mg/kg) and EB (25 and 50 mg/kg) alone or in combination therapy in peripheral (A), central (B), and total (C) locomotor activity. Animals received EtB in sterile 0.9% saline at the rate of 1 μ l/min into the hippocampal CA1 area. Three days after injection, the rats were randomly divided into 11 groups. Distilled water (as vit B12 vehicle) and vit B12 were gavaged and peanut oil (as EB vehicle) and EB were administrated by intraperitoneal injection. The control group received intra-hippocampal saline (as solvent). Treatment duration was 5 days. Data are shown as mean±SD, one-way ANOVA and Tukey–Kramer. ****P*<0.001 compared with the control group, and **P*<0.05 and ****P*<0.001 compared with the sham groups

Con: Control; P: Peanut oil; D: Distilled water; Vit: Vitamin; EB: Estrogen benzoate; EtB: Ethidium bromide (n=7)

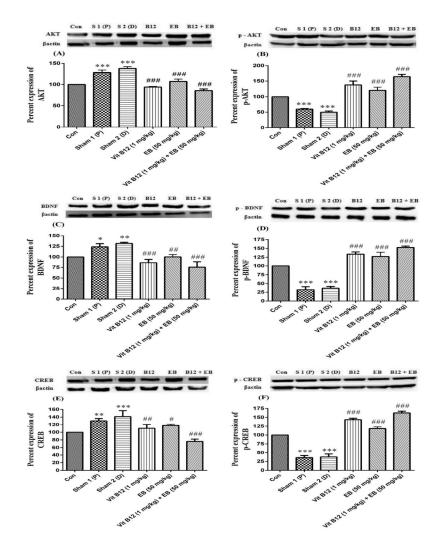


Figure 4. Effects of vit B12 (0.25, 0.5, and 1 mg/kg) and EB (25 and 50 mg/kg) alone or in combination therapy on the protein levels of AKT (A), p-AKT (B), BDNF (C), p-BDNF(D), CREB (E), and p-CREB (F). Animals received EtB in sterile 0.9% saline at the rate of 1 μ l/min into the hippocampal CA1 area. Three days after injection, the rats were randomly divided into 11 groups. Distilled water (as vit B12 vehicle) and vit B12 were gavaged and peanut oil (as EB vehicle) and EB were administrated by intraperitoneal injection. The control group received intra-hippocampal saline (as solvent). Treatment duration was 5 days. Data are shown as mean±SD, one-way ANOVA and Tukey–Kramer. **P*<0.05, ***P*<0.01, and ****P*<0.001 compared with the control group and **P*<0.05, ***P*<0.01, and ****P*<0.001 compared with the sham groups Con: Control; P: Peanut oil; D: Distilled water; Vit: Vitamin; EB: Estrogen benzoate; EtB: Ethidium bromide (n=7)

Western blot analysis

The effect of different doses of vit B12 (0.25, 0.5, and 1 mg/kg) and EB (25 and 50 mg/kg) on the rat hippocampus proteins is displayed in Figure 4. The results showed that the protein levels of non-phosphorylated AKT, BDNF, and CREB were significantly up-regulated, while the phosphorylated types of these proteins were down-regulated in the sham groups compared with the control group (P<0.05, P<0.01, and P<0.001). Vit B12 (1 mg/kg), EB (50 mg/kg), and their co-administration markedly decreased the non-phosphorylated AKT, BDNF, and CREB levels. However, the proteins levels of phosphorylated AKT, BDNF, and CREB levels. However, the sham groups (P<0.05, P<0.01, F (5, 12) = 111.8, 157.7, and 302.3, and P<0.001).

Discussion

The results of this study showed that vit B12 (1 mg/kg) and EB (50 mg/kg) either alone or in combination,

at high doses, had a promoting effect on memory task or increased STL as well as peripheral, central, and total locomotor activity compared with the sham groups. Regulation of neurite outgrowth has a key role in neuronal plasticity and neuronal regeneration from injuries. It was clarified that the neuroprotective effect of vit B12 (1 mg/kg) and EB (50 mg/kg), especially in combination, was attributed to its activation of phosphorylated Akt/CREB signaling pathways through increasing the expression of phosphorylated BDNF.

Although the precise mechanisms of demyelination have not been fully elucidated, several previous studies have shown that myelin destruction can happen via pro-inflammatory and neurotoxic factors produced by macrophages. For example, TNF- α and especially autoreactive CD4+ T cells can pass the blood-brain barrier, enter the central nervous system, identify the antigens of myelin sheaths, and start a chronic inflammatory cascade (33). The inflammatory environment in demyelinating MS lesions leads to the production of reactive oxygen species (ROS) and reactive nitrogen species (RNS), proinflammatory cytokines, specific demyelinating antibodies that contribute to disease development and progression (34). In fact, myelin destruction, astrocytic scar formation, and even axonal loss can be a major cause of long-term and permanent disability in patients with neurological disorders, such as Parkinson's and MS diseases (35).

The hippocampus is involved in learning, memory, and attention processes through GABA and cholinergic receptors (28). Any damage resulting from internal or external factors, such as inflammation, can lead to cellular and molecular changes in synaptic integrity, axonal transport, and glutamate homeostasis (36) and impair learning and memory function in EAE mice (37) and MS patients (38). The most important ingredients for myelination and maintenance of neuron axons are vit B12, folic acid, and iron and their deficiency in serum and CSF cause myelin destruction and MS (39).

In this study, we demonstrated that vit B12 reduced EtB-induced fear learning and memory, and locomotor activity impairments in a dose-dependent manner. As stated in previous studies, vit B12 has anti-inflammatory, neuroprotective, and immunomodulatory properties, such as a decrease in the activity of cytokines, like TNF- α (40, 41). In several studies, an important association has been reported between a decrease in the level of vit B12 and an increase in homocysteine concentration and demyelination (42). The vit B12 deficiency can result in damage to mitochondria, oxidative stress, apoptosis, cognitive dysfunction, and brain disorders, such as Alzheimer's and MS diseases (41).

Estrogen, especially estradiol, via ER- β inhibits mitochondrial dysfunction against oxidative stress and apoptosis of neurons and protects neurons from demyelination (43). Estriol and estradiol exert their anti-inflammatory effects through decreasing cytokines and chemokines (44) in the EAE model (45). There is a strong relationship between estrogens, hippocampal CA1 neurogenesis, and spatial learning and memory performances (46). Estradiol may have an effect on the dopaminergic system in the amygdala, prefrontal cortex, and nucleus accumbens (47). Dopamine is a central neurotransmitter that has an impact on effortbased decision-making (48).

According to previous studies, the results of this study showed that EB reduced EtB-induced fear learning and memory impairments and locomotor activity disorders at a dose-dependent manner. Although estradiol has a dual effect on learning and memory functions, some studies indicated that estrogen derivatives promoted learning and memory performances. It may be due to different methods, doses, time of exposure or even genomic and non-genomic effects (49). In one study estradiol administration into the dorsal hippocampus or prefrontal cortex facilitated or impaired the working memory, in the delayed win-shift test, depending on the dose and site of injection. The working memory was impaired 24 hr after estradiol injection into the hippocampus, but not the prefrontal cortex (49). Several studies have also expressed that a high level of estradiol impaired hippocampal-dependent spatial working/ reference memory function in radial arm maze, caudatedependent cued win-stay, and amygdala-dependent conditioned place preference (50), while low levels of estradiol facilitated working memory (50, 51).

We have also shown that vit B12 and EB alone or in combination therapy increased the phosphorylation of BDNF, CREB, and AKT proteins in a dose-dependent manner. BDNF, a small neuroprotective and antiapoptotic secretory protein belongs to a class of the neurotrophin family (52). This protein plays a key role in survival and function of specific neuronal populations, including cholinergic and dopaminergic neurons, astrocytes, oligodendrocytes, and microglia cells, as well as reduction in nervous system immune response (52). The CREB activity has also a critical role in neuronal plasticity and memory formation in the brain. The CREB deficiency increases neurological disorders, such as Alzheimer's disease (53, 54). In addition, Akt is an important signaling pathway oligodendrocyte progenitor cell proliferation, for differentiation, myelination, and memory formation (8, 20, 55).

Estrogens trigger the expression of BDNF via BDNF receptor (trkB) (52). Steroid hormones, especially estrogen, also have a huge impact on the function of brain cells, particularly the cells that produce BDNF and trkB, including the olfactory bulb, hippocampus, cortex, amygdala, septum dorsolateral area of the bed nucleus terminals, and the lateral habenular nucleus (52). Reduced estrogen and BDNF levels were reported in patients with Parkinson's disease and Alzheimer's disease. In addition, estrogen and BDNF affect the function of cholinergic, GABA ergic, and dopaminergic neurons (56). Several studies have shown that estrogen depletion leads to decreased BDNF protein expression in the hippocampus and cortex regions in Alzheimer's and Parkinson's diseases (52). Estradiol also up-regulates the CREB protein and prevents apoptotic pathway (53, 54, 57). Numerous studies have found that ER-β ligand, including DPN (2,3-bis(4hydroxyphenyl)-propionitrile) (20, 55, 58) and indazole chloride (8), reduced the clinical symptoms of the EAE disease. In addition, estrogen compounds exert their neuroprotective effect on progenitor's proliferation, oligodendrocyte differentiation, remyelination, and locomotor function through enhancement of BDNF activity, phosphorylation of ER-B, and activation of PI3K/Akt/mTOR signaling pathway. The main goals in treatment of neurodegenerative diseases like MS are reducing the disease symptoms, preventing oligodendrocytes loss, and improving oligodendrocyte progenitor's proliferation and differentiation. Another purpose is to identify an intervention strategy that stimulates endogenous remyelination and reduces disease progression. According to the findings of this study, it seems that vit B12 and EB have improved fear learning and memory, locomotor activity and increased phosphorylated Akt, BDNF and CREB proteins in rat hippocampus following EtB-induced demyelination in a dose dependent manner.

Conclusion

According to the results of this study, cobalamin (1 mg/kg) and estradiol benzoate (50 mg/kg), either alone or in combination therapy significantly increased fear learning and memory, whereas they

decreased the locomotor activities. Some portion of the effect of an exposure is mediated through upregulating phosphorylated AKT, BDNF, and CREB in rat hippocampus. It seems that co-administration of cobalamin and estradiol benzoate can be helpful in treatment of MS. However, the discovery of the exact mechanisms of hormone effects on cognitive functions, proteins, and genes requires more comprehensive research and since high doses of estradiol often cause memory degradation, low doses of estradiol are recommended.

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Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

1. Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. N Engl J Med 2000; 343:938-952.

2. Miller DH, Barkhof F, Frank JA, Parker GJ, Thompson AJ. Measurement of atrophy in multiple sclerosis: pathological basis, methodological aspects and clinical relevance. Brain 2002; 125:1676-1695.

3. de Villiers JN, Treurnicht FK, Warnich L, Carr J, van Rensburg SJ, Kotze MJ. Analysis of viral and genetic factors in South African patients with multiple sclerosis. Metab Brain Dis 2006; 21:163-169.

4. Rooney RN, Kotze MJ, de Villiers JN, Hillermann R, Cohen JA. Multiple sclerosis, porphyria-like symptoms, and a history of iron deficiency anemia in a family of Scottish descent. Am J Med Genet 1999; 86:194-196.

5. Steinman L. Autoimmune disease. Sci Am 1993; 269:106-114.

6. Wylezinska M, Cifelli A, Jezzard P, Palace J, Alecci M, Matthews PM. Thalamic neurodegeneration in relapsing-remitting multiple sclerosis. Neurology 2003; 60:1949-1954.

7. Ge Y, Grossman RI, Udupa JK, Wei L, Mannon LJ, Polansky M, *et al.* Brain atrophy in relapsing-remitting multiple sclerosis and secondary progressive multiple sclerosis: longitudinal quantitative analysis. Radiology 2000; 214:665-670.

8. Moore SM, Khalaj AJ, Kumar S, Winchester Z, Yoon J, Yoo T, *et al.* Multiple functional therapeutic effects of the estrogen receptor beta agonist indazole-Cl in a mouse model of multiple sclerosis. Proc Natl Acad Sci U S A 2014; 111:18061-18066.

9. Jensen GE, Clausen J. Glutathione peroxidase and reductase, glucose-6-phosphate dehydrogenase and catalase activities in multiple sclerosis. J Neurol Sci 1984; 63:45-53.

10. Takeuchi C, Yamagata K, Takemiya T. Variation in experimental autoimmune encephalomyelitis scores in a mouse model of multiple sclerosis. World J Neurol 2013; 3:56-61.

11. Shabnam Ghaffari HHN, Gholamreza Dehghan. Protective effect of short-term administration of ethanolic saffron extract on improvement of cognitive deficits and decrement of lipid peroxidation induced by ethidium bromide in experimental models of MS. Physiol Pharmacol 2013; 17:315-327.

12. Squire LR, Stark CE, Clark RE. The medial temporal lobe. Annu Rev Neurosci 2004; 27:279-306.

13. Amato MP, Portaccio E, Goretti B, Zipoli V, Hakiki B, Giannini M, *et al.* Cognitive impairment in early stages of multiple sclerosis. Neurol Sci 2010; 31:S211-214.

14. Gold SM, Voskuhl RR. Estrogen treatment in multiple sclerosis. J Neurol Sci 2009; 286:99-103.

15. Miller A, Korem M, Almog R, Galboiz Y. Vitamin B12, demyelination, remyelination and repair in multiple sclerosis. J Neurol Sci 2005; 233:93-97.

16. Reynolds EH, Bottiglieri T, Laundy M, Crellin RF, Kirker SG. VitaminB12metabolisminmultiplesclerosis.ArchNeurol1992; 49:649-652. doi:10.1001/archneur.1992.00530300089014.

17. Herrick SP, Waters EM, Drake CT, McEwen BS, Milner TA. Extranuclear estrogen receptor beta immunoreactivity is on doublecortin-containing cells in the adult and neonatal rat dentate gyrus. Brain Res 2006; 1121:46-58.

18. Polanczyk M, Zamora A, Subramanian S, Matejuk A, Hess DL, Blankenhorn EP, *et al.* The protective effect of 17betaestradiol on experimental autoimmune encephalomyelitis is mediated through estrogen receptor-alpha. Am J Pathol 2003; 163:1599-1605.

19. Liu HB, Loo KK, Palaszynski K, Ashouri J, Lubahn DB, Voskuhl RR. Estrogen receptor alpha mediates estrogen's immune protection in autoimmune disease. J Immunol 2003; 171:6936-6940.

20. Kumar S, Patel R, Moore S, Crawford DK, Suwanna N, Mangiardi M, *et al.* Estrogen receptor beta ligand therapy activates PI3K/Akt/mTOR signaling in oligodendrocytes and promotes remyelination in a mouse model of multiple sclerosis. Neurobiol Dis 2013; 56:131-144.

21. Yarger JG, Babine RE, Bittner M, Shanle E, Xu W, Hershberger P, *et al.* Structurally similar estradiol analogs uniquely alter the regulation of intracellular signaling pathways. J Mol Endocrinol 2013; 50:43-57.

22. Matejuk A, Hopke C, Vandenbark AA, Hurn PD, Offner H. Middle-age male mice have increased severity of experimental autoimmune encephalomyelitis and are unresponsive to testosterone therapy. J Immunol 2005; 174:2387-2395.

23. Morales LB, Loo KK, Liu HB, Peterson C, Tiwari-Woodruff S, Voskuhl RR. Treatment with an estrogen receptor alpha ligand is neuroprotective in experimental autoimmune encephalomyelitis. J Neurosci 2006; 26:6823-6833.

24. Tiwari-Woodruff S, Morales LBJ, Lee R, Voskuhl RR. Differential neuroprotective and antiinflammatory effects of estrogen receptor (ER) α and ER β ligand treatment. Proc Natl Acad Sci U S A 2007; 104:14813-14818.

25. Babri S, Mehrvash F, Mohaddes G, Hatami H, Mirzaie F. Effect of intrahippocampal administration of vitamin C and progesterone on learning in a model of multiple sclerosis in rats. Adv Pharm Bull 2015; 5:83-87.

26. Paxinos G, Watson C. The rat brain in stereotaxic coordinates. Sydney: Academic Press; 1998.

27. Naseri MH, Hesami-Tackallou S, Torabi-Nami M, Zarrindast MR, Nasehi M. Involvement of the CA1 GABAA receptors in MK-801-induced anxiolytic-like effects: an isobologram analysis. Behav Pharmacol 2014; 25:197-205.

28. Yousefi B, Nasehi M, Khakpai F, Zarrindast MR. Possible interaction of cholinergic and GABAergic systems between MS and CA1 upon memory acquisition in rats. Behav Brain Res 2012; 235:231-243.

29. Norozpour Y, Nasehi M, Sabouri-Khanghah V, Torabi-Nami M, Zarrindast MR. The effect of CA1 alpha2 adrenergic receptors on memory retention deficit induced by total sleep deprivation and the reversal of circadian rhythm in a rat model. Neurobiology of Learning & Memory 2016; 133:53-60. 30. Ghadiri A, Etemad L, Moshiri M, Moallem SA, Jafarian AH,



Hadizadeh F, *et al.* Exploring the effect of intravenous lipid emulsion in acute methamphetamine toxicity. Iran J Basic Med Sci 2017; 20:138-144.

31. Etemad L, Jafarian AH, Moallem SA. Pathogenesis of Pregabalin-Induced Limb Defects in Mouse Embryos. J Pharm Pharm Sci 2015; 18:882-889.

32. Valokola MG, Karimi G, Razavi BM, Kianfar M, Jafarian AH, Jaafari MR, *et al.* The protective activity of nanomicelle curcumin in bisphenol A-induced cardiotoxicity following subacute exposure in rats. Environ Toxicol 2019; 34:319-329. 33. McFarland HF, Martin R. Multiple sclerosis: a complicated

picture of autoimmunity. Nat Immunol 2007; 8:913-919.

34. Lucchinetti C, Bruck W, Parisi J, Scheithauer B, Rodriguez M, Lassmann H. Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. Ann Neurol 2000; 47:707-717.

35. Evangelou N, Esiri MM, Smith S, Palace J, Matthews PM. Quantitative pathological evidence for axonal loss in normal appearing white matter in multiple sclerosis. Ann Neurol 2000; 47:391-395.

36. Dutta R, Chang A, Doud MK, Kidd GJ, Ribaudo MV, Young EA, *et al.* Demyelination causes synaptic alterations in hippocampi from multiple sclerosis patients. Ann Neurol 2011; 69:445-454. 37. Ziehn MO, Avedisian AA, Tiwari-Woodruff S, Voskuhl RR. Hippocampal CA1 atrophy and synaptic loss during experimental autoimmune encephalomyelitis, EAE. Laboratory investigation 2010; 90:774-786.

38. Papadopoulos D, Dukes S, Patel R, Nicholas R, Vora A, Reynolds R. Substantial archaeocortical atrophy and neuronal loss in multiple sclerosis. Brain Pathol 2009; 19:238-253.

39. Selzer RR, Rosenblatt DS, Laxova R, Hogan K. Adverse effect of nitrous oxide in a child with 5,10-methylenetetrahydrofolate reductase deficiency. N Engl J Med 2003; 349:45-50.

40. Miller JW. Vitamin B12 deficiency, tumor necrosis factoralpha, and epidermal growth factor: a novel function for vitamin B12? Nutr Rev 2002; 60:142-144.

41. Mastronardi FG, Min W, Wang H, Winer S, Dosch M, Boggs JM, *et al.* Attenuation of experimental autoimmune encephalomyelitis and nonimmune demyelination by IFN-beta plus vitamin B12: treatment to modify notch-1/sonic hedgehog balance. J Immunol 2004; 172:6418-6426.

42. van Rensburg SJ, Kotze MJ, Hon D, Haug P, Kuyler J, Hendricks M, *et al.* Iron and the folate-vitamin B12-methylation pathway in multiple sclerosis. Metab Brain Dis 2006; 21:121-137.

43. Jover T, Tanaka H, Calderone A, Oguro K, Bennett MV, Etgen AM, *et al.* Estrogen protects against global ischemia-induced neuronal death and prevents activation of apoptotic signaling cascades in the hippocampal CA1. J Neurosci 2002; 22:2115-2124.

44. Palaszynski KM, Liu H, Loo KK, Voskuhl RR. Estriol treatment ameliorates disease in males with experimental autoimmune encephalomyelitis: implications for multiple

sclerosis. J Neuroimmunol 2004; 149:84-89.

45. Polanczyk MJ, Carson BD, Subramanian S, Afentoulis M, Vandenbark AA, Ziegler SF, *et al.* Cutting edge: estrogen drives expansion of the CD4+CD25+ regulatory T cell compartment. J Immunol 2004; 173:2227-2230.

46. Gould E, Beylin A, Tanapat P, Reeves A, Shors TJ. Learning enhances adult neurogenesis in the hippocampal formation. Nat Neurosci 1999; 2:260-265.

47. Shima N, Yamaguchi Y, Yuri K. Distribution of estrogen receptor beta mRNA-containing cells in ovariectomized and estrogen-treated female rat brain. Anat Sci Int 2003; 78:85-97. 48. Becker JB, Hu M. Sex differences in drug abuse. Front Neuroendocrinol 2008; 29:36-47.

49. Sinopoli KJ, Floresco SB, Galea LA. Systemic and local administration of estradiol into the prefrontal cortex or hippocampus differentially alters working memory. Neurobiology of Learning and Memory 2006; 86:293-304.

50. Galea LA, Wide JK, Paine TA, Holmes MM, Ormerod BK, Floresco SB. High levels of estradiol disrupt conditioned place preference learning, stimulus response learning and reference memory but have limited effects on working memory. Behav Brain Res 2001; 126:115-126.

51. Holmes MM, Wide JK, Galea LA. Low levels of estradiol facilitate, whereas high levels of estradiol impair, working memory performance on the radial arm maze. Behav Neurosci 2002; 116:928-934.

52. Sohrabji F, Lewis DK. Estrogen-BDNF interactions: implications for neurodegenerative diseases. Front Neuroendocrinol 2006; 27:404-414.

53. Azcoitia I, Arevalo MA, De Nicola AF, Garcia-Segura LM. Neuroprotective actions of estradiol revisited. Trends Endocrinol Metab 2011; 22:467-473.

54. Yang LC, Zhang QG, Zhou CF, Yang F, Zhang YD, Wang RM, *et al.* Extranuclear estrogen receptors mediate the neuroprotective effects of estrogen in the rat hippocampus. PLoS One 2010; 5:e9851.

55. Crawford DK, Mangiardi M, Song B, Patel R, Du S, Sofroniew MV, *et al.* Oestrogen receptor beta ligand: a novel treatment to enhance endogenous functional remyelination. Brain 2010; 133:2999-3016.

56. Blurton-Jones M, Kuan P, Tuszynski M. Anatomical evidence for transsynaptic influences of estrogen on brainderived neurotrophic factor expression. J Comp Neurol 2004; 468:347-360.

57. Jover-Mengual T, Zukin RS, Etgen AM. MAPK signaling is critical to estradiol protection of CA1 neurons in global ischemia. Endocrinology 2007; 148:1131-1143.

58. Tiwari-Woodruff S, Morales LB, Lee R, Voskuhl RR. Differential neuroprotective and antiinflammatory effects of estrogen receptor (ER)alpha and ERbeta ligand treatment. Proc Natl Acad Sci U S A 2007; 104:14813-14818.