

Nitric Oxide Functions; an Emphasis on its Diversity in Infectious Diseases

*¹Hossein Nahrevanian, ¹Marzyeh Amini

Abstract

Nitric oxide is a short-lived mediator, which can be induced in a variety of cell types and produces many physiologic and metabolic changes in target cells. It is important in many biological functions and generated from L-arginine by the enzyme nitric oxide synthase. Nitric oxide conveys a variety of messages between cells, including signals for vasorelaxation, neurotransmission and cytotoxicity. In macrophages, nitric oxide synthase activity appears slowly after exposure of the cells to cytokines and bacterial products, is sustained, and functions independently of calcium and calmodulin. The cytokine- inducible nitric oxide synthase (iNOS) is activated by several immunological stimuli, leading to the production of large quantities of nitric oxide which can be cytotoxic. To date, there have been conflicting reports concerning the clinical significance of nitric oxide in infections. Some authors have proposed that nitric oxide contributes to the development of severe and complicated cases, while others have argued that nitric oxide has a protective role. The aim of this review is to evaluate the functions of nitric oxide production toward oxidative stress induced by infections or inflammations. It is indicated that NO is an important, but possibly not essential contributor in the control of acute phase of infections and it is only part of an immunopathological chain against pathogens. The anti-microbial function does not relate only to nitric oxide action or its related molecules, a combination of nitric oxide and immune factors is required to resolve pathogenic micro-organisms. Consequently, the NO theory in infectious diseases may lead to the novel ideas for therapy and prevention.

Keywords: Infection, Infectious disease, Nitric oxide, Nitric oxide synthase, Reactive nitrogen intermediate

1- Department of Parasitology, Pasteur Institute of Iran, Tehran 13164, Iran

* Corresponding Author: Tel/Fax: +98-21-66968855, email: mobcghn@yahoo.co.uk

Introduction

The nitric oxide (NO) molecule consists of two atoms of oxygen and nitrogen, bound by a double bond. NO has a specific partial molecular polarity with a negative and positive charge on the oxygen and nitrogen atoms respectively (1). NO is a product of L-arginine conversion to L-citrulline by nitric oxide synthase (NOS) enzyme in the presence of nicotine amide adenine dinucleotide phosphate (NADPH) as a co-factor. NO is a reactive free radical, and in the presence of oxygen, is oxidised to a variety of nitrogen oxides (1). NO is known to react rapidly with oxyhemoglobin (*Oxy-Hb*) to give nitrate and *met-Hb* (2).

In addition, NO is a short-lived biological mediator produced by many cell types to induce many functions (3). It has recently been found to be a potent immuno-modulator, which has alternative roles during inflammation, infection and transplant rejection (4). Both oxygen and NO are vital for life processes, but too much of either can damage cells. It is suggested the attachment of NO to proteins enable them to activate gene(s) directly, but the body needs to keep NO in equilibrium by turning on and off expression of NOS gene(s) (5). NO also, has an extraordinary physiological role with an ability to diffuse freely through cell membranes to induce unrelated phenomena among different type of cells offering a new perspective on cell-cell communication (6). NO is produced in the neutrophils, lymphocytes and macrophages as part of the cytotoxic function of these cells (7). Macrophages, neutrophils, Kupffer cells and hepatocytes are stimulated to produce reactive nitrogen intermediate (RNI) *via* TNF- α and IFN- γ pathways (8).

NO acts as both a pro-inflammatory and an anti-inflammatory agent. The mechanisms that underline these effects remain poorly defined (9). Chronic inflammation is known to be associated with increased levels of both NO and reactive oxygen intermediate (ROI) including O₂ and H₂O₂ (10). It is now recognised that NO is not only involved in acute, but also, in chronic inflammation e.g. formation of a cellular granuloma (11). In some

chronic inflammatory states, target cells may be exposed to high amounts of NO as much as 10⁴ molecules NO per cell (10).

Although immunity to infections is complex and not properly understood, a number of different effector mechanisms in addition to NO have been implicated (12). It is suggested that a cascade of reactions leading to NO production are involved in infection processes (13). There are some contradictory reports about the role of NO and related molecules in infectious diseases. Some researchers propose NO is involved in the development of severe diseases, whereas, others argue a protective role for NO (14). Due to its contradictory actions, it is still an open question as to whether NO effects are protective or damaging (15).

We have previously published the detailed applications of detection assays for NO and its up/down stream molecules, NO-modulators, time courses and the associated changes in NO and its metabolite concentrations of host infected by some intracellular parasitic infections (16-19). These will provide a considerable help for this review to be discussed and clarified on NO functions with an emphasis on its diversity in infectious diseases.

Cytotoxicity of NO

Mononuclear cells and several other mammalian cells produce NO after stimulation with cytokines, bacterial endotoxin or antigens of infectious agents (11). Endothelial and Kupffer cells can be rich sources of cytotoxic levels of NO (20) and elevated levels of RNI and ROI can cause tissue damage (21). Peroxynitrite (ONOO⁻) and hydroxyl radical (OH[•]) species oxidise lipids, thiol groups and damage cell membranes (22). In spite of its toxic role during the formation of ONOO⁻ in phagocytosis and inflammation, NO has also, many non-toxic functions (23). Activated macrophages have an important role in the antimicrobial and antitumoural activity of NO (24). Generated NO has cytotoxic properties against tumour cells, intracellular bacteria, protozoa, extra-cellular fungi and helminths (25, 26).

Microbicidal and tumouricidal activities of NO

The antibacterial, antiviral and antifungal actions of NO *in vitro* in many culture media and *in vivo* in animal and human models are fully explained (27-29). NO exerts its protective function not only through direct antibacterial action, but also by preventing apoptosis and thereby contributing to antimicrobial defence during salmonellosis (30). Patients with brucellosis had significantly elevated serum levels of NO compared to healthy controls (31). In addition, NO is produced during *Pseudomonas aeruginosa* keratitis and may protect tissues from damage (32) or iNOS expressions were well correlated with *Helicobacter pylori* density, acute and chronic inflammation of gastric mucosa (33). In human immunodeficiency virus (HIV) negative and HIV co-infected tuberculosis (TB) patients, low levels of exhaled NO were observed which could be a risk factor in acquiring TB and the relative importance of NO in human TB (34). Moreover, as an antifungal, NO plays an important role in platelet-activating factor (PAF)-induced protection against systemic candidiasis caused by *Candida albicans* (35). However, there are some reports indicating no protective effects or unchanged NO levels during some infections e.g. chronic hepatitis C (36).

The possible key role of NO in the killing of infectious agents and cancer cells should be considered during control of infection and tumour cell growth (4). However, there are substantial literatures on NO suggested a potent antimicrobial role for NO. Both NO and NO₂ inhibit growth, respiration and active transport of fungi and bacteria (26). Some pathogens including *Herpes simplex*, *Cryptococcus neoformans* are reported to be inhibited by RNI (37), therefore, NO formation may be one of the principle mechanisms for decreasing infection (38).

Antiparasitic effects of NO, RNI and ROI

A role for antiparasitic effects of NO and RNI *in vivo* and *in vitro* have been demonstrated against a number of parasites including *Plasmodium spp.*, *Leishmania spp.*,

Toxoplasma gondii, *Schistosoma spp.* and *Trypanosoma brucei* (2, 15, 23, 37-41). Antiparasitic activity of NO has been illustrated with two most important malaria and leishmania parasites (42). NO reported to destroy some intracellular parasites such as *L. major* (43). Moreover, ROI produced by macrophages and granulocytes are responsible for phagocytosis of *Try. cruzi*, *Toxo. gondii*, *Leishmania spp.* and *Plasmodium spp.* (23). There is evidence that the killing of intracellular parasites (*L. major*, *Toxo. gondii*) and extracellular parasites (*Schistosoma spp.*) by activated macrophages through IFN- γ , correlates with the release of NO and RNI (23, 26). NO production has been shown to act directly as a leishmanicidal factor of parasitised macrophages. It seems that NO is not only necessary, but is also sufficient to account for the entire antiparasitic activity (42). The roles of NO and superoxide have been also described in the killing of enteropathogenic protozoa e.g. *Giardia lamblia* (44). The role of NO on human *mansoni* schistosomiasis points to a possible regulatory role of NO in the development of granulomas (11). Furthermore, the formation of NO, RNI and ONOO⁻ has been reported for other parasites including *L. amazonensis* (45), *Opisthorchis viverrini* (46), *Clonorchis sinensis* (38) and *L. mexicana* (47). Increased NO synthesis might have a protective rather than pathological role in the majority of parasitic infections including malaria (39) and leishmania (19). Therefore, the involvement of NO and its up/downstream metabolites in parasitic infections is in the agenda and under debate; however it is required more investigations.

NO and C-reactive protein (CRP)

In addition to NO, CRP is a major acute phase protein present in normal serum, which increases significantly after most forms of tissue injuries and infections as a non-specific innate defence mechanism of the host. CRP as a protein is mainly regulated at the transcriptional level, induced by cytokines (48). It is a marker of inflammatory reactions and cytokine activation (49), which is

produced very early after infection (50). CRP is reported to be a critical element during a majority of infections (51). The data have been revealed a correlation of CRP and NO in some infections. This may clarify the co-involvement of CRP and NO as two major immune elements during infection (52); however, it is not justified, whether the CRP / NO production is beneficial or detrimental to the host. Notwithstanding the conflicting publications, the role of CRP (53) and NO (54, 55) in the immune responses to infections remains uncertain. It is suggested that NO alone or in accompany with CRP and other chemokines is involved in protective or pathogenic responses of human infections (52).

Antimicrobial effects of NO by DNA damage

NO can directly attack many molecules including DNA, but the reaction rate is very slow. It also inactivates ribonucleotide reductase, a key enzyme in the biosynthesis of DNA (24). NO can inhibit several intracellular enzymes or change cellular gene transcription machinery e.g. inhibition of DNA binding to the transcription factor (56). NO has also been shown to cause G:C→A:T transitions, DNA strand breaks (57), induction of oxidative DNA damage in activated macrophages and inhibition of enzyme(s) involved in DNA repair (58). There are some studies indicating the effect of NO on DNA damage (59), activation of poly ADP-ribose polymerase (PARP) (60), inhibition or reduction of DNA repair enzymes in DNA synthesis (61), inhibition of DNA-methyl transferase (57), inactivation of ribonucleotide reductase (62) and promotion of nitrosative deamination of DNA bases (10). One of the DNA modifications induced by exposure to ONOO⁻, NO and O₂ is the formation of 8-nitroguanine and 8-oxoguanine as well as DNA single strand breakage (63). Mutagenesis and carcinogenesis are promoted by alkylation of specific sites in DNA or bases (G to O6-methylguanine), or on the ring-nitrogen positions in A, G, C and T. DNA repair proteins are also inhibited by NO-derived agents such as N₂O₃ *in vitro* and *in vivo* (10).

Antimicrobial effects of NO on mitochondrial respiration

Another intracellular target for NO is mitochondria. It is known that activated macrophages inhibit mitochondrial respiration. NO changes the ion currents through the mitochondrial membrane leading to the release of Ca²⁺ into the cytosol (57). Two distinct effects of NO reported for mitochondria are: *i*) accelerated onset of swelling Ca²⁺ loaded transition, and *ii*) changing the permeability of mitochondria (64). It is suggested that mitochondria are inactivating the NO iron-dependent enzyme system, controlling mitochondrial respiration and chemical energy (65).

Antimicrobial effects of NO by apoptosis

NO may contribute to the development of apoptosis, an endogenous process of programmed cell death (24). NO appears to cause both apoptotic and carcinogenic effects. It may inhibit induction of the tumour-suppressor protein (P53) as a guardian of the genome (59). There are some studies reporting apoptosis and necrosis induced by NO, in human chondrocytes (65), rat neurones (66) and rat Islet cells (67). Pro-apoptotic effects are caused by high amounts of NO produced by inducible NOS (iNOS), but anti-apoptotic effects can occur due to the continuous activity of endothelial NOS (eNOS) (68). Consequently, NO causes apoptotic and necrotic cell death in susceptible cells, depending on cell type and the time of exposure (59).

Pathological effects of NO by neurotransmission and vascular tone in host

One of the important features described for NO is neurotransmission. In both the brain and peripheral nervous system, NO is implicated in neurotoxicity associated with stroke and neurodegenerative diseases (4). NO is a neuromodulator in its own right and may impair consciousness by increasing acetylcholine release and inhibiting N-methyl-D-aspartate (NMDA) activity in the brain. The most important type of NOS enzyme to be involved in neurotransmission is suggested as

neuronal form of NOS (nNOS) (69). NO and O₂ in endothelium modulates vascular tone as a vasodilator substance (14). In addition, NO modulates basal coronary artery tone and it is responsible for the flow-mediated vasodilatation (70).

Other functions of NO in infections

NO may react with proteins and nucleic acids by binding to haem groups, guanylate cyclase (GC), Hb and cytochrome-C oxidase. Theoretically, NO may react with nucleophilic centres like sulphur, nitrogen, oxygen and aromatic carbons. It seems SH groups are prime targets for NO [30]. NO and O₂ rapidly react to produce ONOO⁻, which is a potent oxidant of proteins, lipids and DNA (38). There are some examples of damage of cell membrane integrity, apoptosis, changes in cell cycle and DNA strand breaks, which were induced by NO and its metabolites (71).

Conclusive remarks

Taken together, the data provided by researchers, highlight the fact that NO and / or its related molecules have many functions and are involved in a large number of inflammations, infectious diseases, and biomedical concepts, but the involvement is not independent of other immune events. It is indicated that NO is an important, but possibly not essential contributor in the control of acute phase of infection. Although, the protective immune responses against micro-organisms is multifactorial and the final effector molecules that mediate organism death are not known, NOS, NO and RNI have been significantly

implicated (72-74). It is concluded that NO is only part of an immunopathological chain against infection and the antimicrobial function does not relate only to NO action, so, a combination of NO and other immune factors is required to resolve pathogens. Therefore, the involvement of NO and its up / downstream molecules as an immuno-protective target in infections is highly under debate; and it is required more investigations to be resolved (75-77).

An association of some key cytokines appears to be essential for NOS gene regulation. Perhaps, NO comes from several cellular sources, further study in defining these sources will be important for the understanding of cell-mediated defence mechanism(s) in infectious diseases. However, the involvement of NO in infected host is conflicting, the complex relationship between symptoms, genetic polymorphisms and NO production in populations require more studies to address their immunomodulatory roles in viral, bacterial, parasitic and fungal infections (78, 79).

Although, the knowledge about cytotoxic effects of NO is steadily increasing, we are still at the beginning of understanding as to how, why, when and where cells are affected by NO. Consequently, the NO theory in infectious diseases may lead to the novel ideas for therapy and prevention (80).

Acknowledgment

I would like to express my gratitude to Dr. Mohsen Abolhassani from the Department of Immunology, Pasteur Institute of Iran for his constructive comments.

References

1. Lancaster JRJ. A tutorial on the diffusibility and reactivity of free nitric oxide. *Nitric Oxide* 1997; 1:18-30.
2. Nahrevanian H, Dascombe MJ. The role of nitric oxide and its up/downstream molecules in malaria: cytotoxic or preventive? *Southeast Asian J Trop Med Public Health* 2003; 4:4-50.
3. Nussler AK, Di Silvio M, Billiar TR, Hoffman RA, Geller DA, Selby R, *et al.* Stimulation of the nitric oxide synthase pathway in human hepatocytes by cytokines and endotoxin. *J Exp Med* 1992; 176:261-264.
4. Ellis G, Adatia I, Yazdanpanah M, Makela SK. Nitrite and nitrate analyses: a clinical biochemistry perspective. *Clin Biochem* 1998; 31:195-220.
5. Hede George K. Nitric Oxide: From pollutant to biochemical celebrity. *Duke University Research Magazine*, University of Utah Dar es Salaam Tanzania. 2000. Available at: <http://www.dukenews.duke.edu/dr97/nitric.htm>.
6. Clark IA, Rockett KA, Cowden WB. Possible central role of nitric oxide in conditions clinically similar to cerebral malaria. *Lancet* 1992; 340:894-896.

7. Burdon MG, Butler AR, Renton LM. A study of antibacterial activity of nitric oxide donor compounds. In: Moncada S, Stamler J, Gross S, Higgs EA. The biology of nitric oxide. Ed. London: Portland Press Ltd; 1996. p. 170.
8. Motard A, Landau I, Nussler A, Grau G, Baccam D, Mazier D, Targett GA. The role of reactive nitrogen intermediates in modulation of gametocyte infectivity of rodent malaria parasites. *Parasite Immunol* 1993; 15:21-26.
9. Granger DL, Kubes P. Nitric oxide as anti-inflammatory agent. In: Packer L, editor. Nitric oxide, Part B; physiological and pathological processes. 269th ed. Academic Press; 1983. p. 435-441.
10. Jourdeuil D, Kang D, Grisham MB. Interactions between superoxide and nitric oxide: Implications in DNA damage and mutagenesis. *Front Biosci* 1997; 2:189-196.
11. Oliveira DM, Silva-Teixeira DN, Carmo SA, Goes AM. Role of nitric oxide on human *Schistosomiasis mansoni*: upregulation of *in vitro* granuloma formation by N omega-nitro-L-arginine methyl ester. *Nitric Oxide* 1998; 2:57-65.
12. Good MF, Doolan DL. Immune effector mechanisms in malaria. *Curr Opin Immunol* 1999; 11: 412-419.
13. Hommel M. Immunology of malaria. In: WHO. Health co-operation papers, Quaderni di cooperazione sanitaria. World Health Organization 1996. p. 53-70.
14. Chiwakata CB, Hemmer CJ, Dietrich M. High levels of inducible nitric oxide synthase mRNA are associated with increased monocyte counts in blood and have a beneficial role in *Plasmodium falciparum* malaria. *Infect Immun* 2000; 68:394-399.
15. Nahrevanian H. Nitric oxide involvement during malaria infection; Immunological concepts, mechanisms and complexities; A novel review. *J Trop Med Parasitol* 2004; 27:93-101.
16. Nahrevanian H, Dascombe MJ. Nitric oxide and reactive nitrogen intermediates in lethal and nonlethal strains of murine malaria. *Parasite Immunol* 2001; 23:491-501.
17. Nahrevanian H, Dascombe MJ. Expression of inducible nitric oxide synthase (iNOS) mRNA in target organs of lethal and non-lethal strains of murine malaria. *Parasite Immunol* 2002; 24:471-478.
18. Dascombe MJ, Nahrevanian H. Pharmacological assessment of the role of nitric oxide in mice infected with lethal and non-lethal species of malaria. *Parasite Immunol* 2003; 25:149-159.
19. Nahrevanian H, Farahmand M, Aghighi Z, Assmar M, Amirkhani A. Pharmacological evaluation of anti-leishmanial activity by *in vivo* nitric oxide modulation in Balb/c mice infected with *Leishmania major* MRHO/IR/75/ER; An Iranian strain of cutaneous leishmaniasis. *Exp Parasitol* 2007; 116:233-240.
20. Klotz FW, Scheller LF, Seguin MC, Kumar N, Marletta MA, Green SJ, *et al*. Co-localization of inducible-nitric oxide synthase and *Plasmodium berghei* in hepatocytes from rats immunized with irradiated sporozoites. *J Immunol* 1995; 154:3391-3395.
21. Zuber M, Miesel R. Elevated levels of reactive nitrogen intermediates in serum of patients with auto-immune and inflammatory rheumatic disease. In: Moncada S, Feelisch M, Busse R, Higgs EA, editors. The biology of nitric oxide; physiopathological and clinical aspects. London and Chapel Hill, London, UK: Portland Press; 1994. p. 503-505.
22. Munzel T, Heitzer T, Harrison DG. The physiology and pathophysiology of the nitric oxide/superoxide system. *Herz* 1997; 22:158-172.
23. Roitt IM, Brostoff J, Male D. Immunology. 5th ed. London: Mosby Publication; 1998. p. 132-314.
24. Beckman JS. Biochemistry of nitric oxide and peroxynitrite. In: Kubes P. Springer RG. Ed. Nitric oxide, A modulator of cell-cell interactions in the micro-circulations. USA: Landes Co; 1995. p. 1-17.
25. Ferreira SH. Direct blockade of inflammatory hyperalgesia by nitric oxide analgesics. In: Moncada S, Feelisch M, Busse R, Higgs EA. editors. The biology of nitric oxide; physiopathological and clinical aspects. London and Chapel Hill, London, UK: Portland Press ; 1994. p. 324-326.
26. Mellouk S, Green SJ, Nacy CA, Hoffman SL. IFN-gamma inhibits development of *Plasmodium berghei* exoerythrocytic stages in hepatocytes by an L-arginine-dependent effector mechanism. *J Immunol* 1991; 146: 3971-3976.
27. Vazquez-Torres A, Stevanin T, Jones-Carson J, Castor M, Read RC, Fang FC. Analysis of nitric oxide-dependent antimicrobial actions in macrophages and mice. *Methods Enzymol* 2008; 437:521-538.
28. Poole RK. Nitric oxide and nitrosative stress tolerance in bacteria. *Biochem Soc Trans* 2005; 33:176-180.
29. Hussain S, Malik M, Shi L, Gennaro ML, Drlica K. *In vitro* model of mycobacterial growth arrest using nitric oxide with limited air. *Antimicrob Agents Chemother* 2009; 53:157-161.
30. Alam MS, Zaki MH, Sawa T, Islam S, Ahmed KA, Fujii S, *et al*. Nitric oxide produced in Peyer's patches exhibits antiapoptotic activity contributing to an antimicrobial effect in murine salmonellosis. *Microbiol Immunol* 2008; 52:197-208.
31. Refik M, Mehmet N, Durmaz R, Ersoy Y. Cytokine profile and nitric oxide levels in sera from patients with brucellosis. *Braz J Med Biol Res* 2004; 37:1659-1663.
32. Wang W, Xue M, Willcox M, Thakur A. Role of nitric oxide in *Pseudomonas aeruginosa* keratitis caused by distinct bacterial phenotypes. *Eye Contact Lens* 2008; 34:195-197.

Nitric Oxide Functions in Infections

33. Kim SS, Sung YJ, Park MK, Lim CH, Yang HJ, Kim TH, *et al.* The change of cyclooxygenase-2 and inducible nitric oxide synthase in the gastric mucosa one year after eradication of *Helicobacter pylori*. *Korean J Gastroenterol* 2008; 52:286-292.
34. Idh J, Westman A, Elias D, Moges F, Getachew A, Gelaw A, *et al.* Nitric oxide production in the exhaled air of patients with pulmonary tuberculosis in relation to HIV co-infection. *BMC Infect Dis* 2008 24; 8:146.
35. Kim HA, Kim SH, Ko HM, Choi JH, Kim KJ, Oh SH, *et al.* Nitric oxide plays a key role in the platelet-activating factor-induced enhancement of resistance against systemic candidiasis. *Immunology* 2008; 124:428-435.
36. Lluch P, Cortina B, Vila JM, Segarra G, Mauricio MD, Del Olmo JA, *et al.* Unchanged plasma levels of dimethylarginines and nitric oxide in chronic hepatitis C. *Scand J Gastroenterol* 2008; 25:1-5.
37. Rockett KA, Awburn MM, Cowden WB, Clark IA. Killing of *Plasmodium falciparum in vitro* by nitric oxide derivatives. *Infect Immun* 1991; 59:3280-3283.
38. Modlin R, Rickinson A. Immunity to infection. *Curr Opin Immunol* 2000; 12: 387-389.
39. Nahrevanian H. Immune effector mechanisms of nitric oxide pathway in malaria: Cytotoxicity versus cytoprotection. *Braz J Infect Dis* 2006; 10:283-292.
40. Nahrevanian H, Gholizadeh J, Farahmand M, Assmar M, Sharifi K, Ayatollahi Mousavi SA, *et al.* Nitric oxide induction as a novel immunoepidemiological target in malaria-infected patients from endemic areas of the Islamic Republic of Iran. *Scand J Clin Lab Invest* 2006; 66:201-210.
41. Clark IA, Al-Yaman FM, Cowden WB, Rockett KA. Does malarial tolerance, through nitric oxide, explain the low incidence of autoimmune disease in tropical Africa? *Lancet* 1996; 348:1492-1494.
42. Liew FY. Regulation of nitric oxide synthase in macrophages. In: Moncada S, Stamler J, Gross S, Higgs EA, editors. *The biology of nitric oxide: Enzymology, biochemistry and immunology*. London, UK: Portland Press; 1992. p. 223-229.
43. Tizard IR. Effector T-Cell functions. In: Tizard IR, editor. *Immunology, An introduction*. Chapt. 15, 3rd ed. USA: Saunders College Publishing; 1992. p. 258-273.
44. Fernandes PD, Assreuy J. Role of nitric oxide and superoxide in *Giardia lamblia* killing. *Braz J Med Biol Res* 1997; 30: 93-99.
45. Augusto O, Linares E, Giorgio S. Possible roles of nitric oxide and peroxynitrite in murine leishmaniasis. *Braz J Med Biol Res* 1996; 29: 853-862.
46. Haswell-Elkins M, Satarug S, Sithithaworn P, Mairiang E, Mairiang P, Elkins D. Nitrate excretion and parasite-specific T lymphocyte responses of humans infected with the liver fluke, *Opisthorchis viverrini*. In: Moncada S, Marletta MA, Hibbs JRJB, Higgs A, editors. *The biology of nitric oxide: Physiological and clinical aspects*. London, UK: Portland Press; 1992. p. 380.
47. Mannick EE, Oliver PD, Sadowska-Krowicka H, Miller MJS. Inhibition of inducible nitric oxide synthase reverse endotoxin tolerance in cultured macrophages. In: Moncada S, Stamler J, Gross S, Higgs EA, editors. *The biology of nitric oxide*. London: Portland Press Ltd; 1996. p.144.
48. Ablj HC, Meinders AE. C-reactive protein: history and revival. *Eur J Intern Med* 2002; 13: 412-422.
49. Jakobsen PH, McCay V, N'Jie R, Olaleye BO, D'Alessandro U, Zhang G-H, *et al.* Decreased antitoxic activities among children with clinical episodes of malaria. *Infect Immun* 1998; 66:1654-1659.
50. McCarty MF. AMPK activation may suppress hepatic production of C-reactive protein by stimulating nitric oxide synthase. *Med Hypotheses* 2004; 63: 328-333.
51. Kremsner PG, Winkler S, Wildling E, Prada J, Bienzle U, Graninger W, *et al.* High plasma levels of nitrogen oxides are associated with severe disease and correlate with rapid parasitological and clinical cure in *Plasmodium falciparum*. *Trans R Soc Trop Med Hyg* 1996; 90: 44-47.
52. Nahrevanian H, Gholizadeh J, Farahmand M, Assmar M. Patterns of co-association of C-reactive protein and nitric oxide in malaria in endemic areas of Iran. *Mem Inst swaldo Cruz* 2008;103:39-44.
53. Gyan B, Kurtzhals JAL, Akanmori BD, Ofori M, Goka BQ, Hviid L, *et al.* Elevated levels of nitric oxide and low levels of haptoglobin are associated with severe anaemia in African children. *Acta Trop* 2002; 83:133-140.
54. Clark IA, Awburn MM, Whitten RO, Harper CG, Liomba NG, Molyneux ME, *et al.* Tissue distribution of migration inhibitory factor and inducible nitric oxide synthase in falciparum malaria and sepsis in African children. *Malar J* 2003; 2:1-17.
55. Cramer JP, Mockenhaupt FP, Ehrhardt S, Burkhardt J, Otchwemah RN, Dietz E, *et al.* iNOS promoter variants and severe malaria in Ghanaian children. *Trop Med Int Health* 2004; 9:1074-1080.
56. Vodovotz Y. Control of nitric oxide production by transforming growth factor-beta1: mechanistic insights and potential relevance to human disease. *Nitric Oxide* 1997; 1:3-17.
57. Wink DA, Laval J. The Fpg protein, a DNA repair enzyme, is inhibited by the biomediator nitric oxide *in vitro* and *in vivo*. *Carcinogenesis* 1994; 15: 2125-29.
58. Kroncke KD, Fehsel K, Kolb-Bachofen V. Nitric oxide: cytotoxicity versus cytoprotection: how, why, when, and where? *Nitric Oxide* 1997; 1:107-120.
59. Nguyen T, Brunson D, Crespi CL, Penman BW, Wishnok JS, Tannenbaum SR. DNA damage and mutation in human cells exposed to nitric oxide *in vitro*. *Proc Natl Acad Sci USA* 1992; 89:3030-3034.

60. Zhang J, Dawson VL, Dawson TM, Snyder SH. Nitric oxide activation of poly (ADP-ribose) synthetase in neurotoxicity. *Science* 1994; 263: 687-689.
61. Kwon NS, Stuehr DJ, Nathan CF. Inhibition of tumor cell ribonucleotide reductase by macrophage-derived nitric oxide. *J Exp Med* 1991; 174:761-767.
62. Rockett KA, Awburn MM, Rockett EJ, Cowden WB, Clark IA. Possible role of nitric oxide in malarial immunosuppression. *Parasite Immunol* 1994; 16: 243-249.
63. Szabo C, Ohshima H. DNA damage induced by peroxynitrite: subsequent biological effects. *Nitric Oxide* 1997; 1:373-385.
64. Balakirev MY, Khramtsov VV, Zimmer G. Modulation of the mitochondrial permeability transition by nitric oxide. *Eur J Biochem* 1997; 246: 710-718.
65. Blanco FJ, Ochs RL, Schwarz H, Lotz M. Chondrocyte apoptosis induced by nitric oxide. *Am J Pathol* 1995; 146:75-85.
66. Bonfoco E, Krainc D, Ankarcrona M, Nicotera P, Lipton SA. Apoptosis and necrosis: two distinct events induced, respectively, by mild and intense insults with N-methyl-D-aspartate or nitric oxide/superoxide in cortical cell cultures. *Proc Natl Acad Sci USA* 1995; 92:7162-7166.
67. Kroncke KD, Kolb-Bachofen V, Berschick B, Burkart V, Kolb H. Activated macrophages kill pancreatic syngeneic islet cells *via* arginine-dependent nitric oxide generation. *Biochem Biophys Res Commun* 1991; 175:752-758.
68. Dimmeler S, Zeiher AM. Nitric oxide and apoptosis: another paradigm for the double-edged role of nitric oxide. *Nitric Oxide* 1997; 1:275-281.
69. White NJ. Malaria pathophysiology. In: Sherman IW, editor. *Malaria, parasite biology, pathogenesis and protection*. Washington DC, Am Soc Microbiol: ASM Press; 1998. p. 371-385.
70. Shiode N, Morishima N, Nakayama K, Yamagata T, Matsuura H, Kajiyama G. Flow-mediated vasodilation of human epicardial coronary arteries: effect of inhibition of nitric oxide synthesis. *J Am Coll Cardiol* 1996; 27:304-310.
71. Burney S, Tamir S, Gal A, Tannenbaum SR. A mechanistic analysis of nitric oxide-induced cellular toxicity. *Nitric Oxide* 1997;1:130-144.
72. Nahrevanian H, Dascombe MJ. Reactive nitrogen intermediate (RNI) levels inside and outside *Plasmodium* infected red blood cells in murine malaria. *J Trop Med Parasitol* 2003; 26:13-19.
73. Nahrevanian H, Dascombe MJ. Simultaneous increases in immune competent cells and nitric oxide in the spleen during *Plasmodium berghei* infection in mice. *J Microbiol Immunol Infect* 2006; 39:11-17.
74. Nahrevanian H, Gholizadeh SJ, Farahmand M, Aghighi Z, Assmar M, Abolhassani M. Reactive nitrogen intermediate production and tolerance variability in different mouse strains after *in vivo* treatment with lipopolysaccharide from *Salmonella abortus equi*. *J Microbiol Immunol Infect* 2005; 38:164-168.
75. Nahrevanian H. Direct monitoring of *in situ* and *in vitro* nitric oxide release in the brain during malaria infection with *Plasmodium berghei* N/13/1A. *J Trop Med Parasitol* 2004; 27:1-6.
76. Nahrevanian H. Nitric oxide and reactive nitrogen intermediates are released by immune response during malaria infection. *J Commun Dis* 2004; 3:23-26.
77. Nahrevanian H. The penetration of *Plasmodium* into red blood cell is a protective mechanism of malaria parasite against high levels of accumulated nitric oxide in blood circulation. *J Parasit Dis* 2004; 28:83-89.
78. Gyan B, Troye-Blomberg M, Perlmann P, Bjorkman A. Human monocytes cultured with and without interferon-gamma inhibits *Plasmodium falciparum* parasite growth *in vitro* *via* secretion of reactive nitrogen intermediates. *Parasite Immunol* 1994; 16: 371-375.
79. Singh R, Manjunatha U, Boshoff HI, Ha YH, Niyomrattanakit P, Ledwidge R, Dowd CS, Lee IY, Kim P, Zhang L, Kang S, Keller TH, Jiricek J, Barry CE 3rd. PA-824 kills nonreplicating *Mycobacterium tuberculosis* by intracellular NO release. *Science* 2008; 322:1337-1338.
80. Playfair JH, Taverne J, Bate CA, de Souza JB. The malaria vaccine: anti-parasite or anti-disease? *Immunol Today* 1990; 11:25-27.