

Review of the traditional uses, phytochemistry, pharmacology and toxicology of giant fennel (*Ferula communis* L. subsp. *communis*)

Maryam Akaberi¹, Milad Iranshahy¹, Mehrdad Iranshahi^{1*}

¹ Biotechnology Research Center, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

ARTICLE INFO

Article type:
Review article

Article history:
Received: Jul 25, 2015
Accepted: Nov 12, 2015

Keywords:
Anticoagulant
Ferula communis L.
Sesquiterpene coumarins
Sesquiterpenes
Toxicity

ABSTRACT

Ferula communis L., subsp. *communis*, namely giant fennel, has extensively been used in traditional medicine for a wide range of ailments. Fresh plant materials, crude extracts and isolated components of *F. communis* showed a wide spectrum of *in vitro* and *in vivo* pharmacological properties including antidiabetic, antimicrobial, antiproliferative, and cytotoxic activities. The present paper, reviews the traditional uses, botany, phytochemistry, pharmacology and toxicology of *F. communis* in order to reveal its therapeutic potential and future research opportunities. A bibliographic literature search was conducted in different scientific databases and search engines including Scopus, Cochrane Library, Embase, Google Scholar, Pubmed, SciFinder, and Web of science. Phytochemical studies have led to the isolation of different compounds such as sesquiterpenes from *F. communis*. This plant has two different chemotypes, the poisonous and non-poisonous chemotypes. Each chemotype is endowed with various constituents and different activities. The poisonous chemotype exhibits anticoagulant and cytotoxic activities with sesquiterpene coumarins as major constituents, while the non-poisonous one exhibits estrogenic and cytotoxic effects with daucane sesquiterpene esters as the main compounds. In addition, although various pharmacological properties have been reported for *F. communis*, antimicrobial activities of the plant have been investigated in most studies. Studies revealed that *F. communis* exhibits different biological activities, and contains various bioactive compounds. Although, antibacterial and cytotoxic activities are the two main pharmacological effects of this plant, further studies should focus on the mechanisms underlying these actions, as well as on those biological activities that have been reported traditionally.

► Please cite this article as:

Akaberi M, Iranshahy M, Iranshahi M. Review of the traditional uses, phytochemistry, pharmacology and toxicology of giant fennel (*Ferula communis* L. subsp. *communis*). Iran J Basic Med Sci 2015; 18:1050-1062.

Introduction

The genus *Ferula* (Apiaceae) with about 170 species, mostly occurs throughout central Asia, the Mediterranean region and Northern Africa. *Ferula* spp. has a long history of medicinal use and their pharmacological effects are well documented in both the human and animal studies. A large number of compounds have been identified in this genus. These include sesquiterpenes (1, 2), sesquiterpene coumarins (3-5), sesquiterpene lactones (6) and sulfur-containing compounds (7-10).

Ferula communis L. subsp. *communis* (Giant fennel) is a latex-containing perennial plant, 1-2.5 m high, odoriferous, with dense roots. Its cylindrical peduncle is green, striated, with slimy exudate. The branches, 8-10 cm long, are alternate (inferior) or opposite (superior). The leaves are glabrous, with a large sheath. The inflorescence is attached on the terminal part of the peduncle. The plant, despite the

name, is not a type of fennel proper belonging to the other genus *Foeniculum*. The name of the phenolic compound ferulic acid, which can be isolated from giant fennel, is derived from the Latin name of the plant.

Two chemotypes of *F. communis* L. have been characterized with different biological effects. Poisonous chemotype contain mainly prenyl coumarins such as ferulenol (1) and related compounds that are responsible for ferulosis (a lethal haemorrhagic disorder which mainly affects goats, sheep, cattle, and horses) and its toxicity. The other chemotype is non-poisonous and contains daucane esters (11, 12). Genetically, these two chemotypes are different and distributed in different geographic regions (13). Non-poisonous plant is used traditionally as a hormonal plant because of its phytoestrogens content, while the poisonous one may cause different intoxication and even death (14).

*Corresponding author: Mehrdad Iranshahi. Biotechnology Research Center, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran. Tel: +98-51-38823255; email: Iranshahim@mums.ac.ir

In the present paper, we aimed to review the traditional uses, pharmacology, phytochemistry and toxicology of this ancient medicinal plant. All related available information on *F. communis* was collected via electronic search (using Pubmed, SciFinder, Scirus, Embase, Google Scholar, Scopus, Cochrane Library and Web of Science) without time limit.

Traditional uses

F. communis has traditionally been used as anti-hysteria and for the treatment of dysentery (15, 16). The rhizomes of this plant which is known as Al-kalakh in Saudi Arabia are used locally as a traditional remedy for the treatment of skin infections, while the roasted flower buds are used against fever and dysentery (17).

Traditional uses of non-poisonous *F. communis* as phytohormone are attributed to ferutin (21), an aromatic ester of a daucane alcohol. It has been reported that this plant acts as a possible source of phytoestrogens of the daucane type. In Morocco, *F. communis* has traditionally been used as a hypoglycemic medicinal plant (18) but its use has been restricted due to its toxicity.

In traditional medicinal books such as Dioscorides, numerous beneficial applications have been reported for this plant. For example, administration of mashed fresh kernel is useful for expulsion of oral bloody humors, and for the treatment of stomachache with diarrhea. It is also recommended for the treatment of snakebite. Preparation of the crushed plants in the form of wick seal bleeding of the bleeding organ. Administration of plant seeds improves cramps, and if rubbing the combination of mashed seeds with oil on the skin, causes perspiration (19).

Phytochemistry

Out of all compounds identified in *F. communis*, sesquiterpene and their derivatives (11, 20, 21), sesquiterpene coumarins (22) and prenylated sesquiterpene coumarins are found abundantly (11, 23, 24).

Investigations on *F. communis* have shown that each of the two identified chemotypes has their own major phytochemicals: one chemotype contains sesquiterpene daucane esters, and the other one is dominated by prenylated coumarins (20, 21).

To consider the chemical differences between two chemotypes of *F. communis*, Arnoldi *et al* (13), studied daucanes and coumarins of two chemotypes using HPLC-DAD-UV, HPLC-ESI-MS and HPLC-APCI-MS analyses. The chromatogram of non-poisonous chemotype showed two main peaks attributed to ferutin (21) and the prenylated coumarin ferulenol (1) as well as 10 minor peaks contributing to daucane esters including lapiferin (43) and jaeskeanadiol benzoate (ferutinol benzoate) (66).

The chromatogram of poisonous chemotype was totally different from the non-poisonous one. Five main peaks were detected possessing a 4-hydroxycoumarin structure. This chemotype was reported to contain the toxic coumarins ferulenol (1) and its ω -hydroxy derivatives (25). Daucane derivatives could not be detected in the 'poisonous' chemotype of *F. communis* (13).

Not only have phytochemical studies been established on various chemotypes of *F. communis*, but several studies also have investigated the major compounds in different plant parts. Studying the fruits of *F. communis* showed that prenylated coumarins were absent, whereas sesquiterpenoid esters, such as the phenylpropane laserin (79) (26) and the daucane ester of anisate (45) were present (20, 26). The fruits from the plants containing prenylated coumarins in their roots and leaves gave in fact a series of 1-oxojaeskeanadiol esters. The major constituent of the fruits was a crystalline mixture of the 5-(O)-angeloyl (46) and isovaleroyl (47) esters of 1-oxojaeskeanadiol. Also, the 5(O)-anisoyl ester of 1-oxojaeskeanadiol (48), was obtained from the more polar fractions (26).

Sesquiterpene coumarins (prenyated coumarins)

Ferula is a genus rich in coumarins, particularly sesquiterpene coumarins (27-29). To date, many sesquiterpene coumarins have been identified from this genus (29). In Figure 1, the chemical structures of sesquiterpene coumarins of this species that have been reported to date, are depicted. Ferulenol (1) is the most abundant and the first coumarin isolated from this species (30). Biological activities of ferulenol have been investigated in several studies (31). In 1986, Valle *et al* isolated two 4-hydroxycoumarin derivatives bearing a farnesyl [ferulenol (1)] and a 12-hydroxy farnesyl residue (2) at C-3 from the latex of *F. communis* (20). In a separate study, these two compounds were isolated from the root sap of *F. communis* var. *genuina* with haemorrhagic activity and toxicity (25). Derivatives of prenylated coumarins ferulenol and ferprenin (3) (a pyrano coumarin) (24) with functional groups hydroxyl, acetoxyl, aldehydic carbonyl at the end of the prenyl residue, have also been isolated from the poisonous chemotype of *F. communis* (11).

Some other sesquiterpene coumarins reported from *F. communis* include samarcandin (22), two new cyclic farnesylcoumarins (4,5), isoferprenin (6) (32), 2-nor-1,2-secoferulenol (33), feselol (7) and colladonin (8) (21, 34).

Further investigation on aerial parts of *F. communis* grown in Saudi Arabia led to the identification of three compounds namely umbelliferone (15), umbelliprenin (16), and farnesiferol A (17) (35). The mentioned compounds have been reported from other *Ferula* Species (36, 37), however, the scientific name of plant used in that study needs to be confirmed by

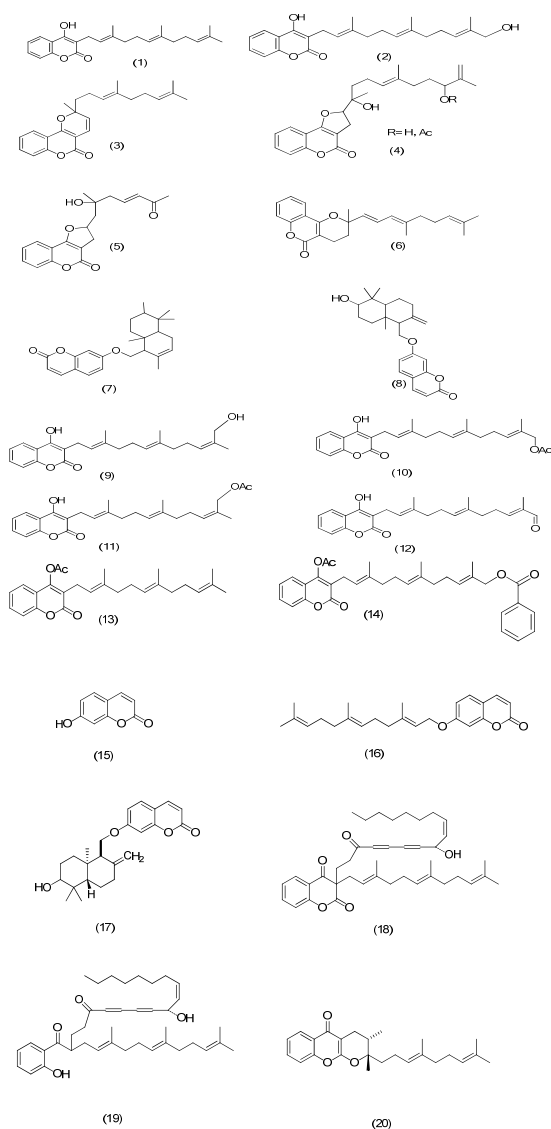


Figure 1. Sesquiterpene coumarins isolated from *F. communis*

further investigation. To our knowledge, *F. communis* is a Mediterranean species of *Ferula* and there is no another document that confirms the presence of *F. communis* in Saudi Arabia.

Daucane sesquiterpene esters

A large number of daucane sesquiterpene esters have been isolated from *F. communis* with a 1,5-*trans*-fused daucane ring system. For instance, Minski *et al* isolated a number of (21-37) daucane sesquiterpenes from the benzene extract of the dried roots of *F. communis* subsp. *communis* (21). Their further investigation on this plant led to the identification of the daucane ester 14-*p*-anisoyloxy-

dauc-4,8-diene (69), fercomin (50), and fercolide (51) (daucane- γ -lactone) which is the first daucane- γ -lactone identified (21, 38). Daucane esters including compounds 22, 52-54 were also reported from this species (20). Daucane esters (55-61) have been reported which are related to jaeschkeanadiol (67) (39). The major constituent of the fruits was a crystalline mixture of the 5-(*O*)-angeloyl (62) and isovaleroyl (63) esters of 1-oxojaeskeanadiol (39). In addition, 5-(*O*)-anisoyl ester of 1-oxojaeskeanadiol (64), was identified from more polar fractions (39). Compound 63 had previously been isolated as the oil from a plant of the Asteraceae family (17).

Investigation of *F. communis* rhizomes, using a bioautography-guided isolation technique, has led to the isolation and characterization of three antibacterial sesquiterpenes, namely, the daucane ester 14-(*O*-hydroxycinnamoyloxy)-dauc-4,8-diene (65), 2*a*-acetyl-6*a*- (benzoyl) jaeschkeanadiol (66) and 2*a*-acetyl-6*a*-(*p*-anisoyl)-jaeschkeanadiol (67) (40).

In a recent work, three daucane sesquiterpenes including acetoxyferutinin, oxojaeskeanadiol anisate (48) and ferutidin (22), together with ferulenol, have also been identified from the aerial parts of *F. communis* (31).

Volatile constituents of *F. communis*

A number of studies have reported the chemical composition of *Ferula* essential oils (41, 42). In a study on *F. communis* leaf oil, forty seven components were identified. The main constituents were myrcene (53.5%) and aristolene (8.5%) (43), however, the oil contained a high content of monoterpenes. In addition, two main sesquiterpenes including, aristolene (8.5%) and (*E,E*)-farnesol (4.3%), were also present in the volatile oil (43). Chemical composition of essential oils from different parts of the plant showed that the content of aristolene reached 19.0% in leaf oil, while it was absent in flower oil. In contrast, the chemical composition of samples obtained from peduncle was different and dominated by sesquiterpenes. The sesquiterpenes aristolene, β -gurjunene, α - and β -selinene were the major components of the essential oil of peduncle. It is notable that the yields of peduncle oil were at least two times lower than those of leaf and flower oils (43).

The study of the chemical composition of the oil from flower-heads using a carbon dioxide supercritical extraction (SPE) (44) showed that the main constituents in the oil were α -gurjunene, β -gurjunene, α -selinene and β -selinene. The oil obtained by SPE had a different appearance with that of obtained by a conventional hydrodistillation method. Indeed, the hydrodistilled oil due to the

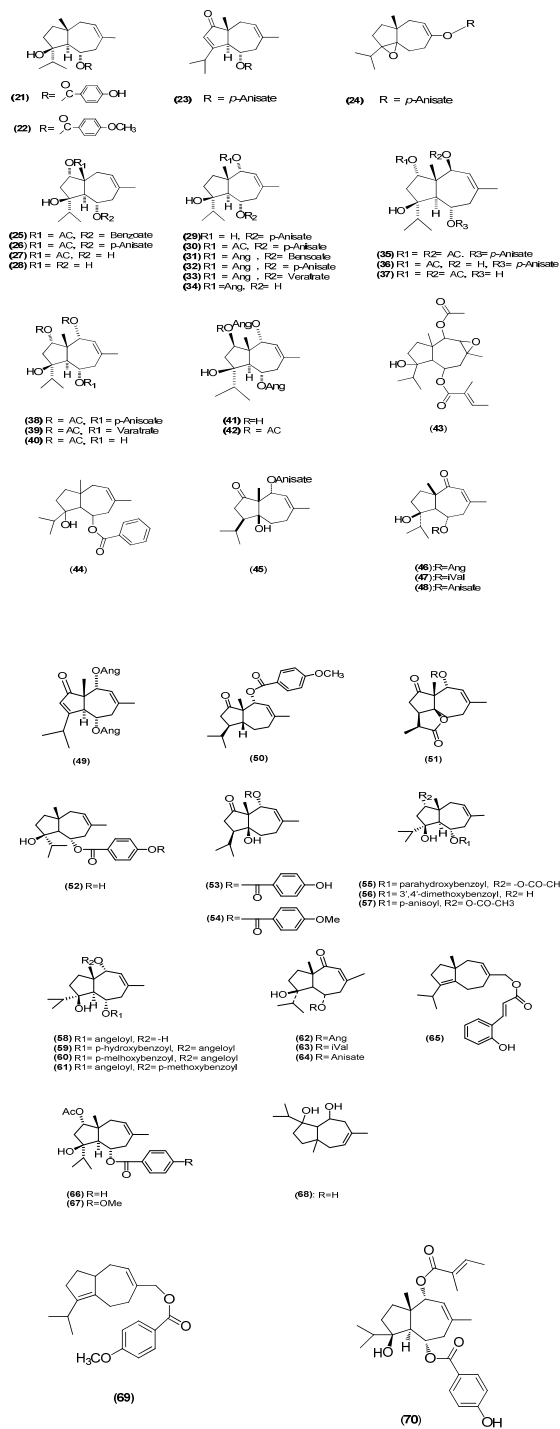


Figure 2. Daucane esters isolated from *Ferula communis*

presence of chamazulene had a blue color while the SPE oil showed a pale yellow color (44). In the SPE oil, there is no thermal degradation of matricin and its conversion to the blue compound chamazulene.

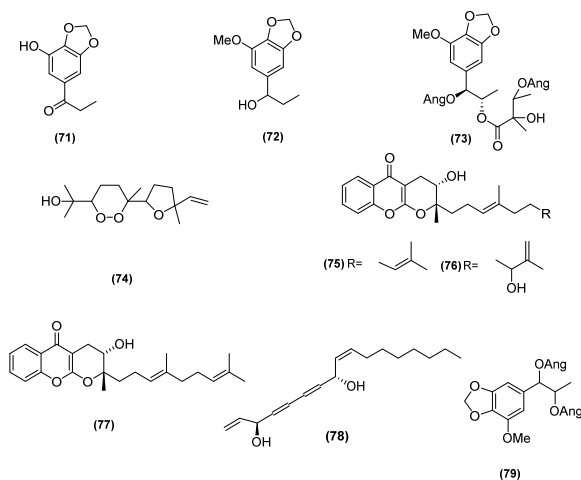


Figure 3. Miscellaneous compounds isolated from *Ferula communis*

Recently, ninety three compounds have also been identified in the total essential oil of Greek *F. communis* subsp. *communis* using a hydrodistillation method. Sesquiterpenes were the most dominant class of compounds in the oil of leaves and inflorescences, while the oil of infructescence was rich in monoterpenes, particularly α -pinene (35.2-40.6%) as the major component (45).

Miscellaneous compounds

Other compounds such as 3,4-methylenedioxy-5-hydroxy-propiofenone (71) (21), phenylpropanoids 1-hydroxy-1-(3'-methoxy-4',5'-methylenedioxy) phenylpropane (72) and 2-epihel-manticine (73) (21) and elemicin (46) were identified from *F. communis* (22). An unusual minor component fercoperol (74) which is a cyclic-endoperoxynerylidol derivative has been also reported (47).

Chromones are the other class of compounds which have been isolated from *F. communis*. For example, an investigation on *F. communis* grown in turkey caused the identification of two cyclic farnesyl-chromone derivatives (75, 76) (22). The chromone sesquiterpene ferchromone (77) (40) was also obtained from the plant (21).

Acetylenes including ferulinolone [3-(1,2-dihydrofalcarinolonyl)-ferulenol] (18) and decarboxyferulinolone [2-nor-3-(1,2-dihydrofalcarinolonyl)-ferulenol] (19) are the other class of metabolites identified from the roots of *F. communis* (48).

Biological effects

Most of *in vitro* studies on *F. communis* and its constituents have been focused on their antimicrobial properties. In 1998, Al-Yahya *et al*, examined anti-bacterial activity of constituents extracted from the rhizome of *F. communis* (40)

including 14-(O-hydroxycinnamoyloxy)-dauc-4,8-diene (**65**), the sesquiterpene coumarin ferulenol (**1**), and the chromane sesquiterpene ferchromone (**77**). The petroleum ether extract of *F. communis* and its constituents showed anti-bacterial effects. Their findings revealed that the sesquiterpene 14-(O-hydroxycinnamoyloxy)-dauc-4,8-diene possessed strong activity against *Staphylococcus aureus*, *Bacillus subtilis*, *Streptococcus durans* and *Enterococcus faecalis* comparable to those of streptomycin sulfate. On the other hand, ferulenol exhibited potent activity against four *Mycobacterium* strains, including *M. intracellulare*, *M. xenopei*, *M. chelonae* and *M. smegmatis*. The compound 14-(O-hydroxycinnamoyloxy)-dauc-4,8-diene exhibited significant activity against Gram-positive bacteria, while ferchromone was found to be less active (40).

Unasho *et al* also studied the *in vitro* antibacterial activities of *F. communis* root extract against clinical isolates of *Streptococcus pyogenes* and *Streptococcus pneumoniae* (49). But the fractions of the plant were found to have weak antibacterial effects to the mentioned bacteria.

The antibacterial activities of three daucane sesquiterpenes including acetoxyferutinol, oxojaeskeanadiol anisate (**48**), ferutidin (**22**) were examined against *Botryotinia fuckeliana*, *Penicillium digitatum*, *Penicillium expansum*, *Monilinia laxa*, *Monilinia fructigena* and *Aspergillus* spp. *F. communis* root extract only affected colony growth of *Botryotinia fuckeliana*, *Monilinia laxa* and *Monilinia fructigena* and no inhibitory activity was observed on conidial germination under applied experimental concentrations. *F. communis* aerial part extract showed no activity on colony growth and was not tested on *Conidia* (31).

Fercoperol (**74**) which is a cyclic-endoperoxynerylidol derivative has been also reported to have amoebicidal activity. Anti-malarial effects of fercoperol are comparable with other standard anti-malarial compounds (47).

Ferulenol and its acetate exhibited antibacterial activity against *Mycobacterium intracellulare*, *Mycobacterium xenopei*, *Mycobacterium chelonae* and *Mycobacterium smegmatis*, with MIC values much lower (1.25–5.0 µg/ml) than those of the positive controls streptomycin sulfate, isonicotinic acid hydrazide (INH) and amikacin sulfate (40, 50). In addition, ferulenol showed synergistic action with INH towards the same strains of *Mycobacterium*.

Plant defensive effects of *Ferula* extracts have been examined against insect herbivores (such as *Spodoptera littoralis* and *Myzus persicae*) and the compounds responsible for these activities were identified (51) by bio-guided fractionation method. The nematocidal (*Meloidogyne javanica*) and phytotoxic (*Lactuca sativa* and *Lolium perenne*) activity of these extracts and compounds have been

also investigated (51). Bioassay-guided fractionation led to the identification of ferulenol as the active anti-feedant compound.

Toxicological properties

Consumption of *F. communis* L. has been found to be associated with bleeding (ferulosis) in animals and humans (52). Studies showed that prenylated 4-hydroxycoumarins play an important role in pro-haemorrhagic effects of *F. communis* (11, 12, 14, 23, 53).

It has been known for a long time that the consumption of *F. communis* L. can cause in livestock an often lethal disease known as ferulosis. Cases of human poisoning from ingestion of *F. communis* have also been reported. Poisoning from *F. communis* causes symptoms in cattle similar to those described for the intoxication by fermented sweet clover, and it has been suggested that the plant contains antithrombic coumarin derivatives (53-55).

Biological tests established that, among compounds isolated from *F. communis*, only prenylated coumarins are toxic (56). Ferulenol (20) and ferprenin (24), the prenylated coumarins isolated from toxic variety of giant fennel, exhibited haemorrhagic effects *in vivo*. Extracts lacking these two compounds still remained toxic to experimental animals. This toxicity was assigned to more polar coumarins which were derivatives of ferulenol and ferprenin (24, 56). Other studies also confirm that only plants containing prenylated coumarins can elicit the toxic symptoms of ferulosis, whereas those containing daucane esters are not toxic (56).

In the root extract of prenylated coumarin-containing plants, based on the ratio of ferulenol to its ω -oxygenated derivatives, two patterns of constituents are detected; one in which ferulenol is the major constituent and one which consists almost exclusively of its more polar ω -oxygenated derivatives (11). But a very similar pattern of constituents is also found in leaves.

The toxic and the non-toxic varieties of *F. communis* differ not only in the presence of prenylated coumarins or sesquiterpene esters, but also in other classes of metabolites including volatile terpenoids and phenylpropanoids (11).

Alzweiri *et al* proposed acetylated ferulenol-oxyferulenol as a marker for fresh *Ferula* toxicity by using a metabolomic approach (57). Their results showed that chloroform extract of fresh *F. communis* had significantly different constituents from those detected in dried plants. One of the main different peaks in HPLC analysis was attributed to the mentioned compound.

Studying the mitochondrial effects of ferulenol revealed that it inhibited oxidative phosphorylation (58). Ferulenol has a dual effect on mitochondrial functions depending on concentration. At low concentrations, ferulenol inhibited ATP synthesis, while

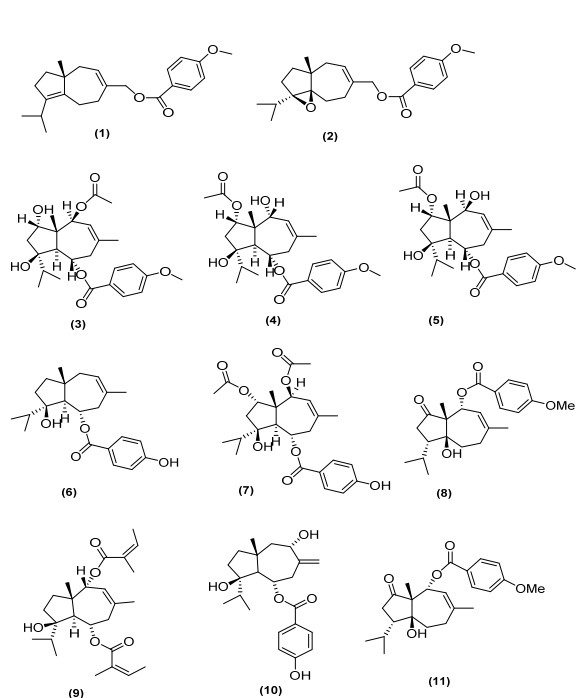


Figure 4. Cytotoxic sesquiterpenes isolated from *Ferula communis*

it did not have any effect on mitochondrial respiration. In contrast, at higher concentrations, ferulenol inhibited oxygen consumption. This data showed that ferulenol, and *F. communis* chemotype containing ferulenol, might exert their toxicity via other mechanisms including mitochondrial dysfunction (58).

Monti *et al* investigated the mechanisms by which *F. communis* caused severe bleeding and characterized anti-coagulant properties of prenylated coumarin ferulenol (59). Ferulenol did not exert its anti-coagulant activity directly. This compound exhibited hepatocyte cytotoxicity, and at non-cytotoxic concentrations (<100 nM), could reduce factor X biosynthesis. Structure activity studies showed that the presence of the prenyl residue in ferulenol derivatives is essential for ferulenol activity (59). Animals treated with ferulenol indicated a sharp decrease in activity for several coagulation factors (60).

Cytotoxic and antiproliferative activity

Because of the importance of cytotoxic effects of *F. communis*, we reviewed this activity and its related mechanisms separately in this section. A large number of studies have been performed for the characterization of its cytotoxic compounds. Some investigations suggested that the cytotoxic effect of *F. communis* could be related to ferulenol. For example, Bocca *et al*, investigated cytotoxic activity of ferulenol, and proposed that ferulenol stimulated tubulin polymerization *in vitro*, and inhibited the binding of

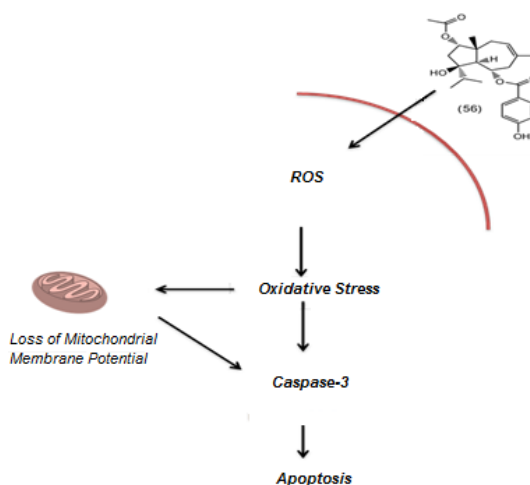


Figure 5. Daucane esters are able to induce apoptosis in cancer cells through interaction with mitochondrial membrane

radiolabeled colchicine to tubulin (61). It rearranged cellular microtubule network into short fibers, and altered nuclear morphology. Remarkably, ferulenol exerted a dose-dependent cytotoxic activity against various human tumor cell lines (61). Determination of the acute LD₅₀s of ferulenol showed that it had a higher LD₅₀ compared to warfarin and thus had a favorable toxicity profile. Hypoprothrombinemia with internal and external hemorrhages caused by the administration of ferulenol in mice was similar to symptoms of ferulosis and anti-vitamin K poisonings. Male mice were more sensitive to intoxication by ferulenol than females (12).

Antiproliferative effects of daucane esters (Figure 5) including certain jaesekanadiol p-hydroxy- and p-methoxybenzoates from *F. communis* on human colon cancer cell lines were investigated (62). Daucane esters from *F. communis* inhibited the growth of WiDr, COLO320-HSR and LS174 colon cancer cells in a dose-dependent manner. ID₅₀ calculated after 72 hr of treatment, revealed that the anti-proliferative capacity of the compounds was in the following descending order: ferutinol (21) > 2 α -OH-ferutinol > ferutinol (22) > siol anisate (54) > lapiferin (43) > jaesekanadiol (68). *Ferula* daucane esters do not act as non-specific toxins randomly impairing the cellular metabolic machinery, but rather act by interaction with specific metabolic pathways (62).

The study on the anti-proliferative mechanism of ferutinol shows that it acts as a calcium ionophore in the T-leukemia cell line Jurkat and this event leads to a collapse of the mitochondrial membrane potential with consequent generation of reactive oxygen species (ROS) that ultimately lead to apoptosis (63).

Dall'Acqua *et al* investigated anti-proliferative activity of constituents from *F. communis* on different cell lines such as HeLa, A549, HL-60, Jurkat, K562, RS

Table 1. Biological activities reported from the bioactive compounds of *Ferula communis*

Compound number	Compound name	Biological activities	Ref.
57 and other daucane esters		Apoptosis inducer through ROS production in leukaemic cells	(56)
1	Ferulenol and ω -hydroxyferulenol	Pro-haemorrhagic activity	(15)
1, 3	ω -oxygenated derivatives of ferulenol and ferprenin	Antithrombotic activity	(46)
66	Falcarindiol	Anti-platelet activity	(79)
1, 65, 77	Daucane ester 14-(O-hydroxycinnamoyloxy)- dauc-4,8-diene Ferulenol Ferchromone	Antibacterial activity	(30)
1	4-hydroxycoumarins Ferulenol	Anticoagulant activity Antimycobacterial activity, inhibitory activity on succinate ubiquinone reductase, cytotoxic	(49, 80) (40, 52, 81, 82)
21, 22	Ferutinin , 2 α -OH-Ferutidin , Ferutidin	Antiproliferative activity	(53)
21	Ferutinin	Anti-inflammatory	(82)
21	Ferutinin	Antifungal activity, Apoptosis inducer, Anti-cancer, bone formation, Calcium ionophoretic	(54, 59, 63, 65, 70, 74, 78)

4; 11 and SEM (64). Compounds (**1-11**, Figure 4) were identified and their anti-proliferative activities have been studied. Compounds (**5**), (**7**), and (**9**) exhibited cytotoxic effects against all the considered cell lines. Compound (**7**) was the most active against Jurkat while (**9**) showed the highest activity against all the other tested cell lines. All the most active compounds possessed ring fusion with trans geometry and the 6-(a)-ester linked group (*p*-methoxybenzoyl or angeloyl), while those with *cis* geometry (**1** and **11**) or 4- and 5-epoxy group (**2**) had poor activity. Also the double bond in the heptatomic ring appeared critical (64). The 10- β -hydroxy function or the 10- β -acetoxy substituent in these compounds is essential for the cytotoxic effects. Notably, for the two isomers (**4**) and (**5**) the IC₅₀ is lower or at least the same (for HL-60) in all the cell lines for the compound bearing the 10 β -hydroxy group (**5**). Data suggest that apoptosis may be the major cause of cell death (64).

Dall'Acqua *et al*, in continuation of their research on anti-proliferative agents from plants, found natural daucane esters isolated from *F. communis* and *Ferulago campestris* induced apoptosis in G1 phase in leukaemic cells through ROS production (65). Among all isolated compounds, only one of them showed more activity against leukemic cell lines (compound **76**). By using structure-activity relationship studies, they realized that there is a relation between cytotoxicity and *trans* geometry of the ring fusion.

Compound **76** also induces alterations of several intracellular antioxidant enzymes (65). This com-

pound promotes oxidative stress in SEM cells via reducing antioxidant enzymes such as superoxide dismutase and other related enzymes.

Discussion

As discussed in previous sections, the most bioactive compounds were highlighted in the mind of readers; however, in this section we have a brief overlook on bioactive compounds of giant fennel. Ferulenol (**1**), a coumarin derivative, is the major constituent of the poisonous chemotype *F. communis*. Ferulenol was isolated for the first time from *F. communis* and is the universal precursor of farnesylated 4-hydroxycoumarins (30). As shown in Table 1, ferulenol has diverse pharmacological effects such as antimicrobial, anticoagulant, antifeedant, and antiproliferative activities. Recently, Gliszczyn' ska *et al*, have reviewed sesquiterpene coumarins including ferulenol as lead compounds in drug discovery (66).

Ferutinin is a natural phytoestrogen with ionophoretic properties. The beneficial effects of non-poisonous chemotype of *F. communis* is attributed to phytoestrogens including ferutinin which was isolated for the first time from *Ferula tenuisecta* Eug. Kor. (67). It has been established that ferutinin has ionophoretic properties, i.e. it increases the permeability of thymocytes, mitochondria, sarcoplasmic reticulum, liposomes and bilayer lipid membranes (BLM) for cations, especially Ca²⁺ (68). It is more selective for divalent cations versus monovalent cations (69). Ferutinin is typically considered as non-toxic compound that is not able to

Table 2. Pharmacological/biological activities reported from *F. communis* in detail

Activity	Dosage form/type of extract	Concentrations/dosages	Tested living system/organ/cell	Active compound(s)	Result(s)	Ref.
Antibacterial	Petroleum ether extract	1.25-5.0 mg/ml	<i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i> , <i>Streptococcus durans</i> , <i>Enterococcus faecalis</i> , <i>Mycobacterium intracellulare</i> , <i>Myc. xenopei</i> , <i>Myc. Chelonei</i> , <i>Myc. Smegmatis</i>	Ester 14-(<i>o</i> -hydroxycinnamoyloxy)-dauc-4,8-diene, ferulenol, ferchromone	Ester 14-(<i>O</i> -hydroxycinnamoyloxy)-dauc-4,8-diene exhibited strong activity against <i>Staph. aureus</i> , <i>Bacillus subtilis</i> , <i>Strep. durans</i> and <i>Entro. faecalis</i> . Ferulenol showed strong antibacterial activity against <i>S. aureus</i> , <i>B. subtilis</i> , <i>S. durans</i> and <i>E. faecalis</i> and four <i>Mycobacterium</i> strains. Ferchromone exhibited potent activity against <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> , <i>Streptococcus durans</i> , <i>Enterococcus faecalis</i> .	(40)
Antibacterial	80% methanol crude extracts and hydro alcoholic solvent fractionates	500 mg/ml to 1000 mg/ml	Clinical isolates of <i>Strep. pyogenes</i> and <i>Strep. pneumoniae</i>		Eighty percent ethanol solubilized fraction was found to have antibacterial effects to all assayed bacteria while aqueous solubilized fractions did not exhibit any effect	(49)
Antifungal	<i>n</i> -hexane	ED ₅₀ almost > 400	The colonies and conidia of <i>Botryotinia fuckeliana</i> , <i>Penicillium digitatum</i> , <i>Pen. expansum</i> , <i>Monilinia laxa</i> , <i>Moni. fructigena</i> and <i>Aspergillus spp</i>	Acetoxyferutinol, oxojaeskeanadiol anisate, ferutidin, and ferulenol	Root extract presented a fungitoxic effect on the colony growth, but it was not able to inhibit the conidia germination. Aerial part extract showed less activity.	(51)
Amoebicidal				Fercoperol		
Antimycobacterial	Petroleum ether		<i>Mycobacterium intracellulare</i> , <i>M. smegmatis</i> , <i>M. xenopei</i> and <i>M. chelonei</i>	Ferulenol	Ferulenol exhibited synergism with isonicotinic acid hydrazide (INH). Its MIC decreased from 5.0 to 0.3 µg/ml.	(40, 50)
Anti-coagulant		<100 nM	BabyHamster Kidney (BHK) cells	A series of prenylated 4-hydroxycoumarins including ferulenol	They did not directly affect blood coagulation but showed hepatocyte cytotoxicity and, at non-cytotoxic concentrations (<100 nM), impaired factor X biosynthesis (40% reduction)	(59)

Aphid antifeedant	Hexane and ethanolic extracts		Herbivorous insects: (<i>Spodoptera littoralis</i> and <i>Myzus persicae</i>) Nematodes: (<i>Meloidogyne javanica</i>) Plants: (<i>Lactuca sativa</i> and <i>Lolium perenne</i>)	Ferulenol	The bioassay-guided search for aphid antifeedant compounds resulted in the isolation of ferulenol. <i>F. communis</i> root ethanolic exhibited a significant but moderate reduction in <i>Spodo. littoralis</i> larvae feeding. The hexane extracts displayed the most activity against <i>Myzus persicae</i> ethanolic extract showed the most phytotoxic activity.	(51)
Ionophoretic and apoptotic properties			Leukemia T-cell line, Jurkat	Ferutin	Ferutin induces Ca ²⁺ mobilization in Jurkat cells. It induces DCm disruption, ROS generation and apoptosis through a caspase-3 dependent pathway	(63)
The cytotoxic and apoptosis- inducing activities		24-46 µg/ml	Human breast (MCF7) and bladder (TCC) cancer cells as well as normal fibroblasts (HFF3)	Ferutin	Ferutin caused DNA damage and apoptosis which was significantly (p<0.001) higher in MCF7 and TCC than that of normal cells HFF3	(72)
Antiproliferative		0.8-60 µM	Human colon cancer cells	Daucane esters: ferutin, 2α-hydroxyferutin, Ferutin, Siolanisate, Lapiferin, Jaeskeanadiol	Daucane esters inhibited the growth of WiDr, COLO320-HSR and LS174 colon cancer cells in a dose-dependent manner. the antiproliferative capacity of the compounds was in descending order: ferutin > 2α-OH-ferutin > ferutin > siolanisate > lapiferin > jaeskeanadiol	(62)
Aphrodisiac		Acute (2.5 mg/kg) and subchronic (0.25 mg/kg/day for 10 days)	Male rats	Ferutin	Ferutin is able to stimulate sexual behavior after acute ingestion in sluggish/impotent animals, but exerts a negative influence on the sexual capacity of potent male rats	(84)
Aphrodisiac			Ovariectomized rats	Ferutin	Ferutin given alone markedly increased the intensity of the lordotic response but failed to significantly affect proceptivity. When administered in combination with estradiol, ferutin reduced the increase in receptivity and proceptivity due to estrogen effects, acting as an antiestrogen.	(86)
Proliferation and osteoblastic differentiation		10- 8 and 10 - 9 M	Human amniotic fluid and dental pulp stem cells	Ferutin	Ferutin treatment induced greater expression of the osteoblast phenotype markers osteocalcin (OCN), osteopontin (OPN), collagen I, RUNX-2 and osterix (OSX), increased calcium deposition and osteocalcin secretion in the culture medium compared to controls	(80)

exert anticoagulant activities like prenylated coumarins. Ferutinin, unlike the other Ca^{2+} ionophores which most likely increases $[\text{Ca}^{2+}]$ via the activation of platelet plasma membrane Ca^{2+} channels and the release of Ca^{2+} from intracellular stores, increases Ca^{2+} levels because of its Ca^{2+} -ionophoretic properties, and does not induce blood platelet aggregation. Ferutinin stimulates the expression of the active form of the GPIIb-IIIa complex and whole blood platelet aggregation only weakly and has no statistically significant effect on the binding of fibrinogen. In fact, ferutinin has inconsistent effects; it raises intra-platelet Ca^{2+} concentration but fails to have an effect on spontaneous blood platelet aggregation. This pattern of responses may be caused by the combination of ferutinin-related Ca^{2+} ionophoric effect and estrogenic properties (70).

Anti-proliferative, cytotoxic and apoptotic effects of ferutinin in different cell lines have been also investigated (62, 71-73). This compound induces calcium mobilization from external and internal stores and eventually triggers apoptosis through a caspase-3 dependent pathway. It caused morphological changes, DNA damage, significant regression in tumor size in cancerous cells, while it showed less toxic effects in non-tumoral cells (74). According to structure-activity studies, *p*-hydroxylation of the benzoyl moiety is crucial for the activity (71, 75) while the parent polyol (jaeschkeanadiol, 2α) was inactive. Homologation, vinylation, methylation of the *p*-hydroxyl substituent and the introduction of oxygen functions on the adjacent carbons were essential for its activity (75).

Studying the effect of ferutinin on human red blood cells revealed that it induced *in vitro* apoptosis through membrane permeabilization and calcium influx (76). Death promoting activities of ferutinin in a number of cancer cells by opening the mitochondrial permeability transition pores have been documented well. The induction of apoptosis in human red blood cells is known as eryptosis or erythroptosis. Ferutinin causes eryptosis/erythroptosis in human red blood cells and simultaneously increases caspase-3 activity and the cytosolic free Ca^{2+} ion level (76).

Phytoestrogens as alternative drugs can be used for the treatment of osteoporosis and many studies have been investigated the anti-osteoporosis effects of phytoestrogens (77). Since ferutinin has estrogenic activity, its beneficial effect on osteoporosis has been investigated. For instance, oral administration of ferutinin in recovering severe osteoporosis has been studied in ovariectomized rats (78, 79). It could increase the recovery of bone loss caused by severe estrogen deficiency. Thus ferutinin can be listed among the potential compounds for the treatment of postmenopausal osteoporosis. Expression of the osteoblast phenotype markers osteocalcin (OCN), osteopontin (OPN), collagen I, RUNX-2 and osterix

(OSX), increase in calcium deposition and osteocalcin secretion are proposed mechanisms for this activity of ferutinin (80). Anti-inflammatory (81), antibacterial (82, 83), aphrodisiac (84-86) and fungicidal (87) properties are other activities proposed for this compound.

Concluding remarks

F. communis subsp. *communis*, namely, giant fennel, showed versatile biological activities (Table 2) with various bioactive natural products. Two chemotypes of *F. communis* L. have been characterized with different chemical constituents. The toxic chemotype mainly produces prenylated coumarins such as ferulenol (1) that are responsible for a lethal haemorrhagic disorder called ferulosis. The non-toxic chemotype contains daucane type sesquiterpenoids such as ferutinin (21) and its pharmacological activities are attributed to these compounds. Antibacterial and cytotoxic activities are two areas that have been covered in previous studies. However, further studies should be focused on mechanisms behind the antibacterial and cytotoxic activities, and those biological activities that have been reported traditionally.

References

1. Iranshahi M, Amin GR, Jalalizadeh H, Shafiee A. New germacrane derivative from *Ferula persica* Willd. var. *latisecta* Chamberlain. *Pharm Biol* 2003; 41:431-433.
2. Iranshahi M, Ghiadi M, Sahebkar A, Rahimi A, Bassarello C, Piacente S, et al. Badrakemonin, a new eremophilane-type sesquiterpene from the roots of *Ferula badrakema* Kos.-Pol. *Iran J Pharm Res* 2009; 8:275-279.
3. Iranshahi M, Rezaee R, Sahebkar A, Bassarello C, Piacente S, Pizza C. Sesquiterpene coumarins from the fruits of *Ferula badrakema*. *Pharm Biol* 2009; 47:344-347.
4. Iranshahi M, Kalateghi F, Sahebkar A, Sardashti A, Schneider B. New sesquiterpene coumarins from the roots *Ferula flabelliloba*. *Pharm Biol* 2010; 48:217-220.
5. Iranshahi M, Masullo M, Asili A, Hamedzadeh A, Jahanbin B, Festa M, et al. Sesquiterpene coumarins from *Ferula gumosa*. *J Nat Prod* 2010; 73:1958-1962.
6. Kasaian J, Iranshahi M, Masullo M, Piacente S, Ebrahimi F, Iranshahi M. Sesquiterpene lactones from *Ferula oopoda* and their cytotoxic properties. *J Asian Nat Prod Res* 2014; 16:248-253.
7. Iranshahi M, Amin G, Amini M, Shafiee A. Sulfur containing derivatives from *Ferula persica* var. *latisecta*. *Phytochemistry* 2003; 63:965-966.
8. Iranshahi M, Amin G, Salehi Sourmaghi MH, Shafiee A, Hadjiakhoondi A. Sulphur containing compounds in the essential oil of *Ferula persica* Willd. var. *persica*. *Flavour Frag J* 2006; 21:260-261.
9. Iranshahi M, Hassanzadeh-Khayyat MH, Fazly Bazzaz BS, Sabeti Z, Enayati F. High content of polysulphides in the volatile oil of *Ferula latisecta* Rech. F. et Aell. fruits and antimicrobial activity of the oil. *J Essent Oil Res* 2008; 20:183-185.
10. Iranshahi M. A review of volatile sulfur-containing compounds from terrestrial plants: biosynthesis,

- distribution and analytical methods. *J Essent Oil Res* 2012; 24:393-434.
11. Appendino G, Tagliapietra S, Gariboldi P, Mario Nano G, Picci V. ω -Oxygenated prenylated coumarins from *Ferula communis*. *Phytochemistry* 1988; 27:3619-3624.
 12. Fraigui O, Lamnaouer D, Faouzi MYA. Acute toxicity of ferulenol, a 4-hydroxycoumarin isolated from *Ferula communis* L. *Vet Hum Toxicol* 2002; 44:5-7.
 13. Arnoldi L, Ballero M, Fuzzati N, Maxia A, Mercalli E, Pagni L. HPLC-DAD-MS identification of bioactive secondary metabolites from *Ferula communis* roots. *Fitoterapia* 2004; 75:342-354.
 14. Appendino G, Cravotto G, Sterner O, Ballero M. Oxygenated sesquiterpenoids from a nonpoisonous sardinian chemotype of giant fennel (*Ferula communis*). *J Nat Prod* 2001; 64:393-395.
 15. Gunther RT. *The Greek Herbal of Dioscorides*. New York: Hafner; 1959.
 16. Heywood VH. *The Biology and Chemistry of the Umbelliferae*. London: Academic Press; 1972.p.1247-1249.
 17. Collette S. *An Illustrated Guide to the Flowers of Saudi Arabia*. London: Scorpion Publishing Ltd; 1985.
 18. Bnouham M, Mekhfi H, Legssyer A, Ziyat A. Medicinal plants used in the treatment of diabetes in Morocco. *Int J Diabetes Metabol* 2002; 10:33-50.
 19. Dioscorides. *DE MATERIA MEDICA*. South Africa: AIBIDIS PRESS; 2000.p. 468.
 20. Valle MG, Appendino G, Nano GM, Picci V. Prenylated coumarins and sesquiterpenoids from *Ferula communis*. *Phytochemistry* 1986; 26:253-256.
 21. Miski M, Mabry TJ. Daucane esters from *Ferula communis* subsp. *communis*. *Phytochemistry* 1985; 24:1735-1741.
 22. Miski M, Jakupovic J. Cyclic farnesyl-coumarin and farnesyl-chromone derivatives from *Ferula communis* subsp. *communis*. *Phytochemistry* 1990; 29:1995-1998.
 23. Valle MG, Appendino G, Nano GM, Picci V. Prenylated coumarins and sesquiterpenoids from *Ferula communis*. *Phytochemistry* 1986; 26:253-256.
 24. Appendino G, Tagliapietra S, Nano GM, Picci V. Ferprenin, a prenylated coumarin from *Ferula communis*. *Phytochemistry* 1988; 27:944-946.
 25. Lamnaouer D, Bodo B, Martin MT, Molho D. Ferulenol and ω -hydroxyferulenol, toxic coumarins from *Ferula communis* var. *genuina*. *Phytochemistry* 1987; 26:1613-1615.
 26. Appendino G, Tagliapietra S, Paglino L, Nano GM, Monti D, Picci V. Sesquiterpenoid esters from the fruits of *Ferula communis*. *Phytochemistry* 1990; 29:1481-1484.
 27. Abd El-Razek MH, Ohta S, Hirata T. Terpenoid coumarins of the genus *Ferula*. *Heterocycles* 2003; 60:689-716.
 28. Iranshahi M, Amanolahi F, Schneider B. New sesquiterpene coumarin from the roots of *Ferula latisepta*. *Avicenna J Phytomed* 2012; 2:133-138.
 29. Nazari ZE, Iranshahi M. Biologically active sesquiterpene coumarins from *Ferula* species. *Phytother Res* 2011; 25:315-323.
 30. Carboni S, Malaguzzi V, Marsili A. Ferulenol a new coumarin derivative from *Ferula communis*. *Tetrahedron Lett* 1964; 5:2783-2786.
 31. Mamoci E, Cavoski I, Simeone V, Mondelli D, Al-Bitar L, Caboni P. Chemical composition and *in vitro* activity of plant extracts from *Ferula communis* and *Dittrichia viscosa* against postharvest fungi. *Molecules* 2011; 16:2609-2625.
 32. Lamnaouer D, Fraigui O, Martin MT, Gallard JF, Bodo B. Structure of isoferprenin, a 4-hydroxycoumarin derivative from *Ferula communis* var. *genuina*. *J Nat Prod* 1991; 54:576-578.
 33. Appendino G, Tagliapietra S, Cravotto G, Nano GM. Structure and synthesis of a prenylated acetophene from *Ferula communis*. *Gaz Chem Ital* 1989; 119:385±388.
 34. Pinar M, Rodriguez B. A new coumarin from *Ferula loscosii* and the correct structure of coladonin. *Phytochemistry* 1977; 16:1987-1989.
 35. Abu-Gabal NS, Edris FM, Abu Mustafa EA. Further investigation on *Ferula communis* grown in Saudi Arabia. *Egypt J Chem* 2008; 51:107-114.
 36. Iranshahi M, Amin G, Shafiee A. A New Coumarin from *Ferula persica*. *Pharm Biol* 2004; 42:440-442.
 37. Iranshahi M, Shahverdi AR, Mirjani R, Amin G, Shafiee A. Umbelliprenin from *Ferula persica* roots inhibits the red pigment production in *Serratia marcescens*. *Z Naturforsch C* 2004; 59:506-508.
 38. Miski M, Mabry TJ. Fercolide, a type of sesquiterpene lactone from *Ferula communis* subsp. *communis* and the correct structure of vaginatin. *Phytochemistry* 1986; 25:1673-1675.
 39. Lamnaouer D, Martin MT, Molho D, Bodo B. Isolation of daucane esters from *Ferula communis* var. *brevifolia*. *Phytochemistry* 1989; 28:2711-2716.
 40. Al-Yahya MA, Muhammad I, Mirza HH, El-Ferally FS. Antibacterial constituents from the rhizomes of *Ferula communis*. *Phytother Res* 1998; 12:335-339.
 41. Sahebkar A, Iranshahi M. Volatile constituents of the genus *Ferula* (Apiaceae): A review. *J Essent Oil Bear Pl* 2011; 14:504-531.
 42. Sahebkar A, Iranshahi M. Biological activities of essential oils from the genus *Ferula* (Apiaceae). *Asian Biomed* 2010; 4:835-847.
 43. Ferrari B, Tomi F, Casanova J. Composition and chemical variability of *Ferula communis* essential oil from Corsica. *Flavour Frag J* 2005; 20:180-185.
 44. Marongiu B, Piras A, Porcedda S. Comparative analysis of the oil and supercritical CO₂ extract of *Ferula communis* L. *J Essent Oil Res* 2005; 17:150-152.
 45. Manolakou S, Tzakou O, Yannitsaros A. Volatile constituents of *Ferula communis* L. subsp. *communis* growing spontaneously in Greece. *Record Nat Prod* 2013; 7:54-58.
 46. Maggi F, Lucarini D, Tirillini B, Sagratini G, Papa F, Vittori S. Chemical analysis of the essential oil of *Ferula glauca* L. (Apiaceae) growing in Marche (central Italy). *Biochem Syst Ecol* 2009; 37:432-441.
 47. Miski M, Mabry TJ, Bohlmann F. Fercoperol, an unusual cyclic-endoperoxynerylidol derivative from *Ferula communis* subsp. *communis*. *J Nat Prod* 1986; 49:916-918.
 48. De Pascual Teresa J, Villaseco MA, Hernandez JM. Complex acetylenes from the roots of *Ferula communis*. *Planta Med* 1986; 6:458-462.
 49. Unasho A, Geyid A, Melaku A, Debela A, Mekasha A, Girma S, et al. Investigation of antibacterial activities of *Albizia gummifera* and *Ferula communis* on *Streptococcus pneumoniae* and *Streptococcus pyogenes*. *Ethiop Med J* 2009; 47:25-32.

50. Mossa JS, El-Ferally FS, Muhammad I. Antimycobacterial constituents from *Juniperus procera*, *Ferula communis* and *Plumbago zeylanica* and their *in vitro* synergistic activity with isonicotinic acid hydrazide. *Phytother Res* 2004; 18:934-937.
51. Mamoci E, Cavoski I, Andres MF, Diaz CE, Gonzalez-Coloma A. Chemical characterization of the aphid antifeedant extracts from *Dittrichia viscosa* and *Ferula communis*. *Biochem Syst Ecol* 2012; 43:101-107.
52. Bruneton J. Toxic plants dangerous to human and animals. Paris: Lavoisier publishing Inc; 1999.
53. Rubiolo P, Matteodo M, Riccio G, Ballero M, Christen P, Fleury-Souverain S, et al. Analytical discrimination of poisonous and nonpoisonous chemotypes of giant fennel (*Ferula communis* L.) through their biologically active and volatile fractions. *J Agr Food Chem* 2006; 54:7556-7563.
54. Carta A. Ferulosis; isolation of the substance with hypoprothrombinizing action from the galbanum of *Ferula communis*. La ferulosi; l'isolamento della sostanza and azione ipoprothrombinemizzante dal galbano della *Ferula communis*. *Boll Soc Ital Biol Sper* 1951; 27:690-693.
55. Lamnaouer D. Anticoagulant activity of coumarins from *Ferula communis* L. *Therapie* 1999; 54:747-751.
56. Tagliapietra S, Aragno M, Ugazio G, Nano GM. Experimental studies on the toxicity of some compounds isolated from *Ferula communis* in the rat. *Res Commun Chem Pathol Pharmacol* 1989; 66:333-336.
57. Alzweiri M, Al-Shudeifat M, Al-Khalidi K, Al-Hiari Y, Affi FU. Acetylated ferulenol-oxy-ferulenol as a proposed marker for fresh *Ferula* toxicity: A metabolomic approach. *J Liq Chromatogr Relat Technol* 2015; 38:283-288.
58. Lahouel M, Zini R, Zellagui A, Rhouati S, Carrupt P-A, Morin D. Ferulenol specifically inhibits succinate ubiquinone reductase at the level of the ubiquinone cycle. *Biochem Biophys Res Commun* 2007; 355:252-257.
59. Monti M, Pinotti M, Appendino G, Dallochio F, Bellini T, Antognoni F, et al. Characterization of anti-coagulant properties of prenylated coumarin ferulenol. *Biochim Biophys Acta* 2007; 1770:1437-1440.
60. Tligui N, Ruth GR, Felice LJ. Plasma ferulenol concentration and activity of clotting factors in sheep with *Ferula communis* variety *brevifolia* intoxication. *Am J Vet Res* 1994; 55:1564-1569.
61. Bocca C, Gabriel L, Bozzo F, Miglietta A. Microtubule-interacting activity and cytotoxicity of the prenylated coumarin ferulenol. *Planta Med* 2002; 68:1135-1137.
62. Poli F, Appendino G, Sacchetti G, Ballero M, Maggiano N, Ranelletti FO. Antiproliferative effects of daucane esters from *Ferula communis* and *F. arrigonii* on human colon cancer cell lines. *Phytother Res* 2005; 19:152-157.
63. Macho A, Blanco-Molina M, Spagliardi P, Appendino G, Bremner P, Heinrich M, et al. Calcium ionophoretic and apoptotic effects of ferutin in the human Jurkat T-cell line. *Biochem Pharmacol* 2004; 68:875-883.
64. Dall'Acqua S, Linardi MA, Maggi F, Nicoletti M, Petitto V, Innocenti G, et al. Natural daucane sesquiterpenes with antiproliferative and proapoptotic activity against human tumor cells. *Bioorg Med Chem* 2011; 19:5876-5885.
65. Dall'Acqua S, Linardi MA, Bortolozzi R, Clauser M, Marzocchini S, Maggi F, et al. Natural daucane esters induces apoptosis in leukaemic cells through ROS production. *Phytochemistry* 2014; 108:147-156.
66. Gliszczynska ABrodelius PE. Sesquiterpene coumarins. *Phytochem Rev* 2012; 11:77-96.
67. Saidkhodzhaev AI, Nikonov GK. The structure of ferutin in. *Chem Nat Compd* 1975; 9:25-26.
68. Zamaraeva MV, Hagelgans AI, Abramov AY, Ternovsky VI, Merzlyak PG, Tashmukhamedov BA, et al. Ionophoretic properties of ferutin in. *Cell Calcium* 1997; 22:235-241.
69. Abramov AY, Zamaraeva MV, Hagelgans AI, Azimov RR, Krasilnikov OV. Influence of plant terpenoids on the permeability of mitochondria and lipid bilayers. *Biochim Biophys Acta* 2001; 1512:98-110.
70. Zamaraeva M, Charishnikova O, Saidkhodzhaev A, Isidorov V, Granosik M, Rózalski M, et al. Calcium mobilization by the plant estrogen ferutin in does not induce blood platelet aggregation. *Pharmacol Rep* 2010; 62:1117-1126.
71. Macho A, Blanco-Molina M, Spagliardi P, Appendino G, Bremner P, Heinrich M, et al. Calcium ionophoretic and apoptotic effects of ferutin in the human Jurkat T-cell line. *Biochem Pharmacol* 2004; 68:875-883.
72. Matin MM, Nakhaeizadeh H, Bahrami AR, Iranshahi M, Arghiani N, Rassouli FB. Ferutin in, an apoptosis inducing terpenoid from *Ferula ovina*. *Asian Pac J Cancer Prev* 2014; 15:2123-2128.
73. Ferretti M, Cavani F, Manni P, Carnevale G, Bertoni L, Zavatti M, et al. Ferutin in dose-dependent effects on uterus and mammary gland in ovariectomized rats. *Histol Histopathol* 2014; 29:1027-1037.
74. Arghiani N, Matin MM, Bahrami AR, Iranshahi M, Sazgarnia A, Rassouli FB. Investigating anticancer properties of the sesquiterpene ferutin in on colon carcinoma cells, *in vitro* and *in vivo*. *Life Sci* 2014; 109:87-94.
75. Appendino G, Spagliardi P, Cravotto G, Pocock V, Milligan S. Daucane Phytoestrogens: A Structure-Activity Study. *J Nat Prod* 2002; 65:1612-1615.
76. Gao M, Wong SY, Lau PM, Kong SK. Ferutin in induces *in vitro* eryptosis/erythroptosis in human erythrocytes through membrane permeabilization and calcium influx. *Chem Res Toxicol* 2013; 26:1218-1228.
77. Fu SW, Zeng GF, Zong SH, Zhang ZY, Zou B, Fang Y, et al. Systematic review and meta-analysis of the bone protective effect of phytoestrogens on osteoporosis in ovariectomized rats. *Nutr Res* 2014; 34:467-477.
78. Ferretti M, Bertoni L, Cavani F, Zavatti M, Resca E, Carnevale G, et al. Influence of ferutin in on bone metabolism in ovariectomized rats. II: Role in recovering osteoporosis. *J Anat* 2010; 217:48-56.
79. Cavani F, Ferretti M, Carnevale G, Bertoni L, Zavatti M, Palumbo C. Effects of different doses of ferutin in on bone formation/resorption in ovariectomized rats. *J Bone Miner Metab* 2012; 30:619-629.
80. Zavatti M, Resca E, Bertoni L, Maraldi T, Guida M, Carnevale G, et al. Ferutin in promotes proliferation and osteoblastic differentiation in human amniotic fluid and dental pulp stem cells. *Life Sci* 2013; 92:993-1003.
81. Geroushi A, Auzi AA, Elhwuegi AS, Elzawam F, Elsherif A, Nahar L, et al. Antiinflammatory

- sesquiterpenes from the root oil of *Ferula hermonis*. *Phytother Res* 2011; 25:774-777.
82. Abourashed EA, Galal AM, Shibl AM. Antimycobacterial activity of ferutinin alone and in combination with antitubercular drugs against a rapidly growing surrogate of *Mycobacterium tuberculosis*. *Nat Prod Res* 2011; 25:1142-1149.
83. Al-Ja'Fari AH, Vila R, Freixa B, Costa J, Cañigüeral S. Antifungal compounds from the rhizome and roots of *Ferula hermonis*. *Phytother Res* 2013; 27:911-915.
84. Zanolì P, Rivasi M, Zavatti M, Brusiani F, Vezzalini F, Baraldi M. Activity of single components of *Ferula hermonis* on male rat sexual behavior. *Int J Impot Res* 2005; 17:513-518.
85. Zavatti M, Montanari C, Zanolì P. Role of ferutinin in the impairment of female sexual function induced by *Ferula hermonis*. *Physiol Behav* 2006; 89:656-661.
86. Zanolì P, Zavatti M, Geminiani E, Corsi L, Baraldi M. The phytoestrogen ferutinin affects female sexual behavior modulating ER α expression in the hypothalamus. *Behav Brain Res* 2009; 199:283-287.
87. Al-Mughrabi KI, Aburjai TA. Fungitoxic activity of root extracts from *Ferula hermonis*. *Phytopathol Mediterr* 2003; 42:141-148.
88. Appendino G, Tagliapietra S, Nano GM, Picci V. An anti-platelet acetylene from the leaves of *Ferula communis*. *Fitoterapia* 1993; 64:179.
89. Lamnaouer D. Anticoagulant activity of the coumarins of *Ferula communis* L. Activite anticoagulante des coumarines de *Ferula communis* L. *Therapie* 1999; 54:747-751.
90. Appendino G, Mercalli E, Fuzzati N, Arnoldi L, Stavri M, Gibbons S, et al. Antimycobacterial coumarins from the Sardinian giant fennel (*Ferula communis*). *J Nat Prod* 2004; 67:2108-2110.
91. Lahouel M, Zini R, Zellagui A, Rhouati S, Carrupt PA, Morin D. Ferulenol specifically inhibits succinate ubiquinone reductase at the level of the ubiquinone cycle. *Biochem Biophys Res Commun* 2007; 355:252-257.