Iranian Journal of Basic Medical Sciences

ijbms.mums.ac.ir

Pathophysiologic mechanisms of obesity- and chronic inflammation-related genes in etiology of polycystic ovary syndrome

Zahra Shaaban¹, Arezoo Khoradmehr², Amir Amiri-Yekta³, Mohammad Reza Jafarzadeh Shirazi¹, Amin Tamadon^{4*}

¹ Department of Animal Science, College of Agriculture, Shiraz University, Shiraz, Iran

² Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

³ Reproductive Biomedicine Research Center, Royan Institute, Tehran, Iran

⁴ The Persian Gulf Marine Biotechnology Research Center, The Persian Gulf Biomedical Sciences Research Institute, Bushehr University of Medical Sciences, Bushehr, Iran

A B S T R A C T
Objective(s) : One of the common heterogeneous reproductive disorders in women of childbearing age is polycystic ovary syndrome (PCOS). It is characterized by lack of fertility due to anovulatory
cycles, hyperandrogenemia, polycystic ovaries, hyperinsulinemia, and obesity. Both reproductive anomalies and metabolic disorders are involved in PCOS pathology. Although the role of increased levels of androgens in initiation of PCOS is almost proven, mechanisms of PCOS pathophysiology are not clear. Here we discuss roles of altered metabolic conditions, obesity, and chronic inflammation in
PCOS pathophysiology. <i>Materials and Methods:</i> : In this review, we attempted to identify genes related to obesity and chronic inflammation aspects of PCOS and their physiological functions to explain the pathways that are regulated by these genes and can be a prominent function in PCOS predisposition. For this purpose, published articles and reviews dealing with genetic evaluation of PCOS in women in peer-reviewed journals in PubMed and Google Scholar databases were included in this review.
Results: Obesity and chronic inflammation are not prominent diagnostic features of PCOS, but they play an important role in exacerbating metabolic and hyperandrogenic states. <i>ADIPOQ, FTO TGF</i> β , and <i>DENND1A</i> as the main obesity- and chronic inflammation-related genes have roles in PCOS pathophysiology.
<i>Conclusion:</i> It seems that genes related to obesity pathology in genomic research association, are related to metabolic aspects and body mass index in PCOS patients. Genomes have roles in chronic inflammation, followed by obesity, in the pathogenesis of PCOS.

Shaaban Z, Khoradmehr A, Amiri-Yekta A, Jafarzadeh Shirazi MR, Tamadon A. Pathophysiologic mechanisms of obesity- and chronic inflammationrelated genes in etiology of polycystic ovary syndrome. Iran J Basic Med Sci 2019; 22:1378-1386. doi: 10.22038/IJBMS.2019.14029

Introduction

Polycystic ovary syndrome (PCOS), affects 5-15% of reproductive-aged women in the world. It is one of the most prevalent endocrine abnormalities that are characterized by biochemical hyperandrogenemia, chronic anovulation, and polycystic ovaries (1). Besides, PCOS relates to other symptoms among which insulin resistance, abdominal obesity, chronic inflammation, elevated risk of metabolic syndrome, type 2 diabetes, and cardiovascular diseases are more pronounced (1). The genetic basis of PCOS by twin-family based association studies was confirmed, and accordingly, heritability of PCOS is estimated 70% (2). The genome-wide association studies (GWAS) identified 15 susceptible single nucleotide polymorphisms (SNP) in 11 loci (LHCGR, FSHR, THADA, INSR, DENND1A, RAB5B, C9orf3, YAP1, SUMO1P1, TOX3, and HMGA2) in PCOS Chinese women (3, 4). These loci (LHCGR, FSHR, INSR, THADA, DENND1A, and YAP1) are likely more important because they were replicated in European population,

too (5-8). Undoubtedly, effects of environmental factors in interacting with genetic agents in creating important features, such as hyperandrogenism and insulin resistance of PCOS cannot be ignored (9).

IJ MS

The lifestyle interacts with genetics in the incidence of weight excess and obesity. For instance, sisters with irregular menses and hyperandrogenism were overweight in contrast with ones who have regular cycles and hyperandrogenism. Abdominal adiposity, excess weight, and obesity are usually common among PCOS patients. Obesity plays an underscored role in pathophysiology of insulin resistance and hyperandrogenism (10). For this reason, diet modification and exercise play an important role in improving PCOS reproductive and metabolic phenotypes (11, 12).

PCOS is also related to elevation of inflammatory indices such as increased levels of c- reactive protein, interleukins, and white blood cell count as well as oxidative stress and endothelial dysfunction, which are

^{*}Corresponding author: Amin Tamadon. The Persian Gulf Marine Biotechnology Research Center, The Persian Gulf Biomedical Sciences Research Institute, Bushehr University of Medical Sciences, Bushehr, Iran. Tel/Fax: +98-77-3332-8724; Email: amintamaddon@yahoo.com

components of low-grade chronic inflammation (13). Adipose tissue products can play a role in inflammation creation and its exacerbation. Interleukin 6 (IL-6) secreted from adipocytes, stimulates secretion of hepatic C-reactive protein (CRP) and both of them elevate in PCOS and obese patients (14). Abdominal obesity is associated with dysregulation of sex steroid levels in PCOS, such as androgen overproduction and reduction of sex hormone-binding globulin (SHBG). The problem is that obesity has a confounding effect on the exacerbating of PCOS main traits, hyperandrogenism, and insulin resistance (15).

Obesity influences the development or escalation of PCOS. Mother's obesity in late pregnancy predisposes her daughter to PCOS in adulthood, but effective pathophysiologic mechanisms are not clear (16). Also, the intrauterine androgen excess may create adiposity in offspring at adulthood. Abdominal obesity can be important in ovarian or adrenal hyperandrogenism in PCOS, although, the elevated levels of androgens can contribute to abdominal fat deposition (10). Apart from the key role of hyperandrogenism in PCOS pathophysiology, obesity, inflammation, and other metabolic factors have the main role in aggravating steroidogenic abnormalities. Furthermore, genetic predisposition underlies both primary steroidogenic disorders and other exacerbating factors (10). Obesity is related to low-grade chronic inflammation, which contributes to insulin resistance by adipocytokines functions such as tumor necrosis factor alpha (TNF α) and adiponectin (17, 18).

In this review, we attempted to identify genes related to obesity and chronic inflammation aspects of PCOS and their physiological functions to explain the pathways that are regulated by these genes and can be a prominent function in PCOS predisposition. Initially, we searched the major databases such as PubMed and Google Scholar based on gene, PCOS, obesity, inflammation, etiology, patient, and human keywords which were taken from the MeSH site. These genes are divided into two groups that are effective in obesity and chronic inflammation, respectively. Then, they are separated from each gene, their actions are described, and thus the involved physiologic pathways identified. Eventually, hypotheses associated with these findings are presented.

Even though the role of increased levels of androgens in the initiation of PCOS is almost documented and approved by most authors, mechanisms of PCOS pathophysiology are not clear. In fact, hyperandrogenism is the common loop of different hypotheses presented on PCOS etiology. These issues are explained in detail in a previous paper about the role of steroid and gonadotropin related genes in PCOS pathophysiology (19). But roles of altered metabolic conditions are prominent.

Obesity-related genes in etiology of polycystic ovary syndrome

Since the genetic nature of obesity is well known (20), environmental factors such as lifestyle, nutrition, and exercise also contribute to obesity. Obesity leads to insulin resistance, followed by other events that ultimately result in the occurrence of PCOS. Obesity can

result from the reduced lipolytic effect of insulin in PCOS, in turn, by increased serum inflammatory mediators such as TNF α and high-sensitivity CRP (hs-CRP), leads to beta cell dysfunction and insulin resistance of PCOS. Obesity can intensify the hyperandrogenic state in PCOS because abdominal obesity alters fat-soluble androgen clearance and deposition and also exacerbates hyperandrogenism by reduction of SHBG levels.

Obesity is associated with PCOS, and between 38–88% of PCOS patients are overweight or obese (21). Obesity is accompanied by other PCOS metabolic attributes, such as insulin resistance (22). Obesity is related to insulin resistance and compensatory hyperinsulinemia (23). Central adiposity and hyperandrogenemia by reduction of natural insulin sensitizer adipokines such as adiponectin lead to the development of insulin resistance in PCOS (24). Also, both obesity and insulin resistance elevate the risk of cardiovascular and type 2 diabetes mellitus diseases (25), which are considered metabolic features of PCOS. Body mass index (BMI) of women with PCOS is usually higher than normal women (26). Several growth factors and inflammatory factors were increased in obesity and could promote ovarian androgen overproduction (23). Thus, obesity can develop insulin resistance, androgen excess, and inflammation in PCOS women.

Dyslipidemia is prevalent in almost 70% of women with PCOS (27). In addition to obesity, dyslipidemia causes insulin resistance; in contrast, insulin resistance also affects lipid metabolism and serum lipid parameters, all of which are characteristics of PCOS (28). Dyslipidemia and metabolic syndrome have a pivotal role in PCOS development but not obesity and insulin resistance (25).

It seems that factors of genetic and lifestyle, both or alone lead to obesity in humans. Moreover, obesity has adverse effects on the physiological pathways in the body, which eventually result in PCOS metabolic parameters (Figure 1). Obesity is related to anovulation, loss of pregnancy, and occurrence of pregnancy complications (pre-eclampsia and gestational diabetes); as well as, delayed or failed responses to therapeutic strategies such as clomiphene citrate and gonadotropins. A five percent weight loss in women increased the rate of normal and spontaneous ovulation and pregnancy (25). Thus it seems the alteration of environmental factors such as exercise and diet is useful in the treatment of PCOS. But the effects of obesity-related genetic variants cannot be forgotten. Below, a number of these genes are described and are summarized in Table 1.

ADIPOQ

It is well known that insulin resistance is a prominent risk factor for obesity. The adipocyte cells are one of the target cells of insulin and secrete several adipokines such as adiponectin (34). About six polymorphisms of the adiponectin gene have been detected in various racial populations and with different environmental factors associated with metabolic abnormalities of PCOS. Among them, SNPs +45(T/G) and +276(G/T) are highly associated with obesity, insulin resistance, and type 2 diabetes mellitus in Asian populations (34). Also, in a family-based study of the Chinese Han population, two SNPs of the ADIPOQ gene are associated with PCOS (30).

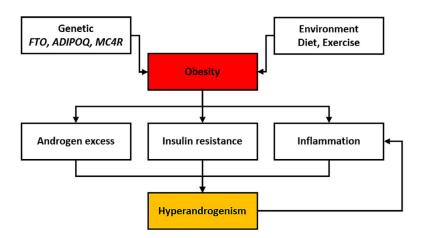


Figure 1. Causes of obesity and its adverse effects are expressed. Obesity mainly can occur due to genetic background and inappropriate lifestyle. Effect of abdominal obesity in exacerbating complications of PCOS is more than peripheral obesity. Abdominal obesity by alteration of metabolic clearance of androgens, increased levels of inflammatory mediators, and elevated beta cells mass in the pancreas is involved in hyperandrogenism, insulin resistance, and inflammation, respectively. The relationship between inflammation and hyperandrogenism is reciprocal, and androgens can create inflammation state

Generally, adiponectin is related to metabolic syndrome (insulin resistance, obesity, and dyslipidemia) in PCOS (42), which reflects the role of obesity in the mediation of adiponectin in PCOS. Because the genetic variants of the ADIPOQ gene in different racial populations were associated with PCOS (Table 1), this gene may be a risk factor for obesity and insulin resistance of PCOS.

FTO

Fat mass and obesity-associated (FTO) gene is a large gene that encodes 2-oxoglutarate-dependent nucleic acid demethylase and is mainly expressed in hypothalamic nuclei regulating feed intake. FTO has also been shown in other tissues such as liver, pancreas, muscles, and adipose tissue (43). Generally, the FTO

Table1. Candidate genes involved in etiology of polycystic ovary syndrome related to obesity and dyslipidemia

Gene	Genetic marker(s)	Type of study	Physiologic function	Studied population	Type of	References
					polymorphism	
ADIPOQ	T45G	Meta-analysis	Glucose regulation and	Different	G276T	(29)
	G276T		fatty acid oxidation	population		
ADIPOQ	rs2241766	Family-based	Glucose regulation and	Chinese Han	rs1501299	(30)
	rs1501299	analysis	fatty acid oxidation			
ADIPOQ	SNPs at position –11377 of	Case-control	Glucose regulation and	Japanese	ND	(31)
	the ADIPOQ gene		fatty acid oxidation			
ADIPOQ	45T/G	Meta-analysis	Glucose regulation and	Different	45T/G	(32)
	276G/T		fatty acid oxidation	population	276G/T	
ADIPOQ	C45G15G(T/G)	Case-control	Glucose regulation and	Chinese Han	+45G15G(T/G)	(33)
	C276(G/T)		fatty acid oxidation		+276(G/T)	
ADIPOQ	+45G15G(T/G)	Case-control	Glucose regulation and	Korean	+276(G/T)	(34)
	+276(G/T)		fatty acid oxidation			
FTO	rs9939609	Case-control	Energy homeostasis	British/Irish	rs9939609	(35)
			regulation			
FTO	Five SNPs	Family-based and	Energy homeostasis	White American	rs1421085	(36)
		case-control	regulation			
FTO	rs9939609	Case-control	Energy homeostasis	Australian	rs9939609	(37)
			regulation			
FTO	rs1421085	Case-control	Energy homeostasis	Korean	rs1421085	(38)
	rs17817449		regulation		rs17817449	
	rs8050136				rs8050136	
MC4R	rs17782313 (T/C)	Case-control	Regulator of melanocortin	Chinese	rs17782313	(39)
			neuronal pathways			
MC4R	rs12970134	Case-control	Regulator of melanocortin	Czech	rs12970134	(40)
			neuronal pathways			
ADIPOQ	rs2241766	Case-control	Metabolic features of PCOS	Iranian	rs2241766 "TT"	(41)
SREBP-2 LXRa	rs2228314	Case-control	Lipid metabolism	Chinese Han	rs2228314 G to C	(28)
	rs11039155				rs11039155 G to A	

ADIPOQ: Adiponectin; FTO: Fat mass, and obesity-associated; MC4R: Melanocortin 4 receptor; SREBP-2: Sterol Regulatory Element Binding Protein-2; LXRa: Liver X Receptor a; ND: No data

Gene	Genetic marker(s)	Type of study	Physiologic function	Studied population	Type of polymorphism	Reference
DENND1A	rs2479106	Case control	Clathrin-mediated endocytosis	Caucasian	rs2479106 G	(61)
DENND1A	rs10818854	Meta-analysis	Clathrin-mediated endocytosis	Asian and	rs10818854	(62)
	rs2479106			European	rs10986105	
	rs10986105					
DENND1A	rs2479106	Meta-analysis	Clathrin-mediated endocytosis	Chinese Han	rs10818854	(63)
	rs10818854					
DENND1A	rs10818854	Retrospective case-	Clathrin-mediated endocytosis	Tunisian	0818854	(64)
	rs2479106	control study			rs10986105	
	rs10986105					
AQP8	rs7198838	case control	Water channel protein	Chinese Han	rs2287798	(65)
	rs1076973					
	rs1076974					
	rs2287797					
	rs2287798					
	rs2287796					
YAP1	rs11225138	Replication study	Transcriptional regulator	Chinese Han	rs11225161 (A/G)	(66)
	rs11225161					
	rs1122516					
TNF	rs1799964	Family association	Pro-inflammatory cytokine	Chinese Han	rs1799964	(67)
	rs1799724					
TGF-β1	rs4803457C/T	GWAS	Low-grade chronic	Chinese Han	rs4803457C/T	(68)
	rs11466313		inflammation			
	deletion/AGG					
	rs2217130C/T					
	rs1800469C/T					
	rs1800470C/T					
TNFα	-308 G/A	case-control	Inflammatory cytokines	Turkish	IL-6 promoter region	(60)
IL-6	-174 G/C				polymorphism	
IL-10	-1082 G/A					
TNFRSF1B	M196R (676 T \rightarrow G) variant in exon 6	GWAS	TNF signaling	Spanish Italian	M196R (676 T→G)	(69)
IL-6	-174 G/C	Case- control	Inflammatory cytokine	Indian	-174 G/C SNP	(70)
TNFα	(-308 G/A),	meta-analysis	Chronic low-grade	Different	No association	(71)
IL-6	(-174 G/C)		inflammation	populations		

Table 2. Candidate genes involved in etiology of polycystic ovary syndrome related to cell proliferation and signaling and chronic inflammation

DENND1A: DENN domain containing 1A; *AQP8*: Aquaporin 8; *YAP1*: yeast associated protein 1; *TNF*: Tumor necrosis factor; *TGF-β1*: transforming growth factor β1; *TNFRSF1B*: TNF receptor 2

gene is associated with type 2 diabetes mellitus and obesity (38). In studies evaluating the association of FTO and PCOS, FTO was mainly associated with BMI and anthropometric parameters in women of various races with PCOS (37-39, 44) (Table 1). Increasing evidence suggests the association of variants of FTO with hyperandrogenemia in PCOS patients (37, 38). The relationship between FTO variants and impaired glucose tolerance, insulin resistance, and hyperandrogenism, that are prominent features of PCOS, are mediated via obesity and BMI (38). So, FTO can be a main genetic factor in predisposing to PCOS, primarily via an effective role in obesity and BMI, and secondarily with influencing the metabolic parameters and hyperandrogenemia.

MC4R

Melanocortin-4 receptor (MC4R) gene encodes the G-protein coupled receptor that is dominantly expressed in the brain and mediates the signaling pathway of melanocortin (45). MC4R has an important role in energy homeostasis and appetite and is the main genetic cause of obesity in humans (45, 46) and animals (47-49). Due to the critical role of MC4R in obesity pathology, the association between different SNPs of the MC4R gene and BMI and obesity in PCOS patients were demonstrated (36, 39, 40). According to the evidence, the MC4R gene via a causal effect on obesity contributes to PCOS etiology. The same evidence has been shown in PCOS animal models, too (50).

SREBP-2 and LXR

Given that insulin resistance disrupts lipid metabolism and serum lipid parameters, central transcription factors in metabolic pathways appear to be likely candidates for PCOS abnormalities (28). The central transcription factors of lipid metabolism, are liver X receptor (NR1H3, LXRa), and sterol regulatory element binding protein-2 (SREBP-2) (51-53). These two transcription factors regulate the expression of effective genes on lipoprotein metabolism, cholesterol homeostasis, and lipogenesis. LXR α transcription factor also plays a key action in insulin secretion of pancreatic

beta cells (53). The SREBP-2, which is encoded by a single gene on human chromosome 22, also plays an important role in maintaining lipid homeostasis (54). In a case-control study on Chinese Han women, variants of these two transcription factors were associated with PCOS (28). Although more research is needed to prove this association.

Chronic inflammation-related genes in etiology of polycystic ovary syndrome

Inflammation can be the key marker of endothelial dysfunction, atherosclerosis and cardiovascular diseases, and also metabolic disruptions of PCOS. It seems inflammatory reactions are more often secondary pathways in PCOS etiology and are affected by obesity and hyperglycemia. Glucose-induced nuclear factor- κ B (NF- κ B) activation from mononuclear cells eventually leads to beta cell dysfunction and insulin secretion irregularities in PCOS (55). Adipokines derived adipocytes contribute to inflammation and insulin resistance development. For instance, decreased adiponectin and increased TNF α and IL-6 are contributed to insulin resistance development.

PCOS is an inflammatory state, and chronic inflammatory-related genes may be effective in the incidence of PCOS through mediating role in hyperandrogenism, obesity, insulin resistance, and anovulation (56). Chronic inflammation is involved in the development of PCOS. Dietary glucose-induced oxidative stress by the particular molecular signaling pathway leads to increased production of pro-inflammatory cytokines from mononuclear cells (MNC).

Hyperandrogenism may be the progenitor of low-grade chronic inflammation in PCOS and via increase of MNC sensitivity, stimulates glucose-induced inflammation (57). On the other hand, the pro-inflammatory cytokines, TNF α , can elevate the production of androgens with upregulation of steroidogenic enzymes and stimulation of proliferation of theca cells. Also, TNF α is a mediator

of insulin resistance, thus it is likely dietary-induced inflammation, is the base of insulin resistance in PCOS (57). Exposure of androgen excess prone the adipocytes to hypertrophy and hypertrophic adipocytes were observed in PCOS women. Hypertrophic adipocytes were more predisposed to inflammation (58). Adipose tissue adipocytes in an autocrine/paracrine manner by secretion of products, some of which are inflammation factors, contributed to low-grade inflammation related to PCOS (13). The low-grade chronic inflammation in PCOS may be related to hyperandrogenism and hypertrophic adipocyte (58).

The correlation between the incidence of PCOS and chronic inflammation has increased attention to the inflammatory factors coding genes and their association with PCOS (59). The imbalance between pro-inflammatory and anti-inflammatory cytokines and cytokine genes polymorphisms may be involved in etiology of PCOS (60). Therefore, inflammatory reactions act as mediators and contribute to the development and aggravation of metabolic properties of PCOS (Figure 2). The genes related to chronic inflammation are presented in Table 2.

TGF-β1

The beta-transforming growth factor- β (TGF- β 1) is a component of multifunctional cytokines family and mediates wound healing, tissue fibrosis, and embryonic development (68). In recent years, pathogenic immunity factors have been widely considered in PCOS, which shows PCOS is a low-grade chronic inflammatory condition (68). PCOS patients showed higher levels of lymphocytes, monocytes, and eosinophils, plus CRPs, TNF α , and IL-6 in serum, all of which are peripheral inflammatory factors (72, 73). Moreover, PCOS ovaries showed chronic inflammation and higher numbers of inflammatory cells compared to normal conditions. Cytokines seem to be important for folliculogenesis and

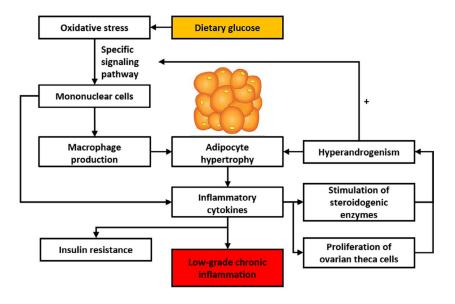


Figure 2. Pathophysiological pathways contribute to low-grade inflammation in polycystic ovary syndrome (PCOS). Glucose-induced oxidative stress by production of inflammatory cytokines leads to insulin resistance. Cytokines via effects on theca cells eventually create hyperandrogenism. Macrophage derived mononuclear cells resulted in adipocytes hypertrophy and in turn by secretion of inflammatory cytokine create low-grade chronic inflammation

ovulation and also participate in the process of follicular atresia and corpus luteum regression by activating immune cells (68, 74). Polymorphism of rs4803457C/T in *TGF* β gene was associated with susceptibility to PCOS and was demonstrated as the main constituent of PCOS development in Chinese women (68).

TNFα

It is reported that the TNF α cytokine is secreted by adipose tissue and plays an essential role in mediating insulin resistance and as a result of obesity (75). The TNF α pro-inflammatory cytokine is known as a mediator of insulin resistance in PCOS (57). Androgen excess promotes the release of TNFa from MNCs in vivo and *in vitro* (76). TNF α plays a cardinal role in oxidative stress and inflammation; in PCOS, production of $TNF\alpha$ is induced by hyperglycemia and hyperandrogenemia (77). Serine phosphorylation of Insulin receptor substrate 1 (IRS-1) seems to be a mechanism for insulin resistance by mediating $TNF\alpha$ (78). Overexpression of TNF α in peripheral tissues such as muscle and adipose, by reducing the tyrosine kinase activity in the insulin signaling pathway, is one of the mechanisms for insulin resistance development mediated by $TNF\alpha$ (78). Also, TNF α directly effects ovary and adrenal function (10) and then increases steroidogenesis in theca cells in vitro (79). Despite the finding that $TNF\alpha$ is the mediator of insulin resistance, a significant association of $TNF\alpha$ gene with PCOS was not observed.

TNFRS1*β*

The TNF receptor superfamily 1β is a member of TNF α receptors that are said to localize on the surface of all normal and tumor cells (80). Type 2 TNF receptor mediates most of the metabolic effects of TNF α . Its serum levels increase in obese subjects correlating with insulin resistance indices (81). The M196R variant in exon 6 of the TNF receptor superfamily 1β (*TNFRS1* β) gene was associated with hyperandrogenism and PCOS in the study of hyperandrogenic and PCOS Spanish women (82). It has also been suggested that TNF α cytokines were able to induce proliferation of theca internal cells in porcine ovaries (80).

IL-6

IL-6 is a pleiotropic cytokine and is secreted by numerous cells including lymphocytes, monocytes, and endothelial cells. Interleukin-6 is involved in reproductive physiologic processes, such as regulating of ovarian steroid production, follicular maturation, fertilization, implantation, and modulating of ovarian development and functions (70). Concentrations of the IL-6 and CRP increase in obese women, but not in PCOS patients (14). It is shown that the polymorphism of the promoter region of the IL-6 gene could be associated with the occurrence of metabolic abnormalities in Turkish PCOS women (60). Also, IL-6 may have a direct effect on ovaries and adrenal cells (10). In vitro studies have shown that IL-6 induced the function of human adrenal cells, in turn, elevated steroidogenesis in adrenals (83). The results of an investigation of IL-6 gene polymorphisms are very inconsistent (Table 2), which is possibly related to racial background, genetic variants, and epigenetic environmental factors among

different populations (70).

DENND1A

Differentially expressed in normal and neoplastic (cell) domain containing 1A (DENND1A) is a member of connecdenn proteins family. These proteins have differential expression in normal and neoplastic domains of cells (84). The DENND1A gene encodes the connecdenn-1 protein, which has a clathrin-binding domain and facilitates receptor-mediated endocytosis and participates in endosomes trafficking (84, 85). Overexpression of variant 2 DENND1A, alternative splicing of DENND1A, in PCOS theca cells was observed, which can contribute to androgen overproduction in these cells (86). High expression of variant 2 DENND1A leads to over mRNA expression of cytochrome P450 17A1 (CYP17A1) and hyperactivity of CYP11A1 and CYP17A1, which in turn increases the level of androgen production in theca cells (86). So, DENND1A can be a mediator for hyperandrogenism of PCOS, and maybe through this pathway connect to hyperandrogenism, the common loop of all hypotheses of PCOS. Previous studies reported DENND1A as a susceptibility locus in PCOS, but there are racial differences about this association (64). Even though the DENND1A locus in various GWAS among different populations is identified as a risk locus in PCOS and maybe a valuable gene in PCOS pathogenesis and diagnosis, it is not applicable to all populations.

Conclusion

To sum up, identifying the root cause of PCOS as a heterogeneous disorder is hard and at the same time, it provides the basis of many investigations. So far, the role of hyperandrogenism and its pre- and postpathways in PCOS development is better explained and confirmed; various backgrounds including genetic, environmental factors, and developmental origins can interfere in its creation. Hyperandrogenism can also be caused by insulin resistance. While the roles of hyperandrogenism and hyperinsulinemia are the major reasons for PCOS development, but abdominal obesity and low-grade inflammation can play a significant role through the mediation of some pathways leading to insulin resistance and hyperandrogenism. As foregoing, chronic inflammation and obesity are not necessarily primary factors, and sometimes their role exacerbates the syndrome or even themselves as a consequence of the syndrome. It seems that genes related to obesity pathology (FTO, ADIPOQ, and MC4R), in genomic research association, are related to metabolic aspects and BMI in PCOS patients. These findings suggest that obesity, especially abdominal phenotype, completes and exacerbates the phenotypic picture of PCOS. Inflammation also occurs, followed by obesity and has the same role in PCOS. It should be kept in mind that these hypotheses are better evaluated by clinical and experimental research such as animal models and therapeutic approaches.

Acknowledgment

The results presented in this paper were part of a student thesis. This review paper was financially

supported by the Department of Animal Science, College of Agriculture, Shiraz University, Shiraz, Iran.

References

1. Azziz R. PCOS in 2015: New insights into the genetics of polycystic ovary syndrome. Nat Rev Endocrinol. 2016;12:74.

2. Vink J, Sadrzadeh S, Lambalk C, Boomsma D. Heritability of polycystic ovary syndrome in a Dutch twin-family study. J Clin Endocrinol Metab. 2006;91:2100-2104.

3. Chen Z-J, Zhao H, He L, Shi Y, Qin Y, Shi Y, *et al*. Genome-wide association study identifies susceptibility loci for polycystic ovary syndrome on chromosome 2p16. 3, 2p21 and 9q33. 3. Nat Genet. 2011;43:55.

4. Shi Y, Zhao H, Shi Y, Cao Y, Yang D, Li Z, *et al.* Genome-wide association study identifies eight new risk loci for polycystic ovary syndrome. Nat Genet. 2012;44:1020-1025.

5. Goodarzi MO, Jones MR, Li X, Chua AK, Garcia OA, Chen Y-DI, *et al.* Replication of association of DENND1A and THADA variants with polycystic ovary syndrome in European cohorts. J Med Genet. 2012;49:90-95.

6. Welt CK, Styrkarsdottir U, Ehrmann DA, Thorleifsson G, Arason G, Gudmundsson JA, *et al.* Variants in DENND1A are associated with polycystic ovary syndrome in women of European ancestry. J Clin Endocrinol Metab. 2012;97:E1342-E1347.

7. Mutharasan P, Galdones E, Peñalver Bernabé B, Garcia OA, Jafari N, Shea LD, *et al.* Evidence for chromosome 2p16. 3 polycystic ovary syndrome susceptibility locus in affected women of European ancestry. J Clin Endocrinol Metab. 2013;98:E185-E190.

8. Brower MA, Jones MR, Rotter JI, Krauss RM, Legro RS, Azziz R, *et al*. Further investigation in europeans of susceptibility variants for polycystic ovary syndrome discovered in genome-wide association studies of Chinese individuals. J Clin Endocrinol Metab. 2015;100:E182-E186.

9. Diamanti-Kandarakis E, Piperi C. Genetics of polycystic ovary syndrome: searching for the way out of the labyrinth. Hum Reprod Update. 2005;11:631-643.

10. Escobar-Morreale HF, San Millán JL. Abdominal adiposity and the polycystic ovary syndrome. Trends Endocrinol Metab. 2007;18:266-272.

11. Huber-Buchholz M-M, Carey D, Norman R. Restoration of reproductive potential by lifestyle modification in obese polycystic ovary syndrome: role of insulin sensitivity and luteinizing hormone. J Clin Endocrinol Metab. 1999;84:1470-1474.

12. van Dam EW, Roelfsema F, Veldhuis JD, Hogendoorn S, Westenberg J, Helmerhorst FM, *et al.* Retention of estradiol negative feedback relationship to LH predicts ovulation in response to caloric restriction and weight loss in obese patients with polycystic ovary syndrome. Am J Physiol Endocrinol Metab. 2004;286:E615-E620.

13. Duleba AJ, Dokras A. Is PCOS an inflammatory process? Fertil Steril. 2012;97:7-12.

14. Escobar-Morreale H, Villuendas G, Botella-Carretero J, Sancho J, San Millan J. Obesity, and not insulin resistance, is the major determinant of serum inflammatory cardiovascular risk markers in pre-menopausal women. Diabetologia. 2003;46:625-633.

15. Pasquali R, Gambineri A. The Endocrine Impact of Obesity and Body Habitus in the Polycystic Ovary Syndrome. Androgen Excess Disorders in Women: Springer; 2006. p. 283-291.

16. Cresswell J, Barker D, Osmond C, Egger P, Phillips D, Fraser R. Fetal growth, length of gestation, and polycystic ovaries in adult life. Lancet. 1997;350:1131-1135.

17. Attie AD, Scherer PE. Adipocyte metabolism and obesity. J

Lipid Res. 2009;50:S395-S399.

18. Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. J Clin Invest. 2005;115:1111-1119.

19. Shaaban Z, Khoradmehr A, Jafarzadeh Shirazi MR, Tamadon A. Pathophysiological mechanisms of gonadotropins–and steroid hormones–related genes in etiology of polycystic ovary syndrome. Iran J Basic Med Sci. 2019;22:3-16.

20. Lee YS. Consequences of childhood obesity. Ann Acad Med Singapore. 2009;38:75-77.

21. Barber TM, Franks S. Genetics of polycystic ovary syndrome. Polycystic Ovary Syndrome. 40: Karger Publishers; 2013. p. 28-39.

22. Fenichel P, Rougier C, Hieronimus S, Chevalier N, editors. Which origin for polycystic ovaries syndrome: genetic, environmental or both? Ann Endocrinol; 2017: Elsevier.

23. Legro RS, editor Obesity and PCOS: implications for diagnosis and treatment. Semin Reprod Med; 2012: NIH Public Access.

24. Luque-Ramírez M, San Millán JL, Escobar-Morreale HF. Genomic variants in polycystic ovary syndrome. Clin Chim Acta. 2006;366:14-26.

25. Beatriz Motta A. The role of obesity in the development of polycystic ovary syndrome. Curr Pharm Des. 2012;18:2482-2491.

26. Xiong W, Lin Y, Xu L, Tamadon A, Zou S, Tian F, *et al.* Circulatory microRNA 23a and microRNA 23b and polycystic ovary syndrome (PCOS): the effects of body mass index and sex hormones in an Eastern Han Chinese population. J Ovarian Resh. 2017;10:10.

27. Legro RS, Kunselman AR, Dunaif A. Prevalence and predictors of dyslipidemia in women with polycystic ovary syndrome. Am J Med. 2001;111:607-613.

28. Zhao J, Hu Z, Cai L, Liu L, Jiang X, Wu L, *et al.* Association between single nucleotide polymorphisms of sterol regulatory element binding protein-2 and liver X receptor α gene and risk of polycystic ovary syndrome in a Chinese Han population. Cell Biochem Biophys. 2014;70:1421-1426.

29. Jia H, Yu L, Guo X, Gao W, Jiang Z. Associations of adiponectin gene polymorphisms with polycystic ovary syndrome: a metaanalysis. Endocrine. 2012;42:299-306.

30. Zhang W, Wei D, Sun X, Li J, Yu X, Shi Y, *et al.* Family-based analysis of adiponectin gene polymorphisms in Chinese Han polycystic ovary syndrome. Fertil Steril. 2014;101:1419-1423. e1413.

31. Baba T, Endo T, Sata F, Nagasawa K, Honnma H, Kitajima Y, *et al.* The contributions of resistin and adiponectin gene single nucleotide polymorphisms to the genetic risk for polycystic ovary syndrome in a Japanese population. Gynecol Endocrinol. 2009;25:498-503.

32. Xian L, He W, Pang F, Hu Y. ADIPOQ gene polymorphisms and susceptibility to polycystic ovary syndrome: a HuGE survey and meta-analysis. Eur J Obstet Gynecol Reprod Biol. 2012;161:117-124.

33. Zhang N, Shi Y-H, Hao C-F, Gu HF, Li Y, Zhao Y-R, *et al.* Association of + 45G15G (T/G) and + 276 (G/T) polymorphisms in the *ADIPOQ* gene with polycystic ovary syndrome among Han Chinese women. Eur J Endocrinol. 2008;158:255-260.

34. Li L, Yun J-H, Lee J-H, Song S, Choi B-C, Baek K-H. Association study of+ 45G15G (T/G) and+ 276 (G/T) polymorphisms in the *adiponectin* gene in patients with polycystic ovary syndrome. Int J Mol Med. 2011;27:283-287.

35. Barber T, Bennett A, Groves C, Sovio U, Ruokonen A, Martikainen H, *et al.* Association of variants in the fat mass and obesity associated (*FTO*) gene with polycystic ovary syndrome. Diabetologia. 2008;51:1153-1158.

36. Ewens KG, Jones MR, Ankener W, Stewart DR, Urbanek M, Dunaif A, *et al. FTO* and *MC4R* gene variants are associated

with obesity in polycystic ovary syndrome. PLoS ONE. 2011;6:e16390.

37. Wehr E, Schweighofer N, Möller R, Giuliani A, Pieber TR, Obermayer-Pietsch B. Association of *FTO* gene with hyperandrogenemia and metabolic parameters in women with polycystic ovary syndrome. Metabolism. 2010;59:575-580.

38. Song DK, Lee H, Oh J-Y, Hong YS, Sung Y-A. FTO gene variants are associated with PCOS susceptibility and hyperandrogenemia in young Korean women. Diabetes Metab J. 2014;38:302-310.

39. Yuan H, Zhu G, Wang F, Wang X, Guo H, Shen M. Interaction between common variants of FTO and MC4R is associated with risk of PCOS. Reprod Biol Endocrinol. 2015;13:55.

40. Bradnová O, Vejražková D, Vaňková M, Lukášová P, Včelák J, Stanická S, *et al.* Metabolic and hormonal consequencies of the obesity risk" MC4R variant (rs12970134) in Czech women. Physiol Res. 2015;64: S187-S195.

41. Ranjzad F, Mahmoudi T, Shemirani AI, Mahban A, Nikzamir A, Vahedi M, *et al.* A common variant in the adiponectin gene and polycystic ovary syndrome risk. Mol Biol Rep. 2012;39:2313-2319.

42. Zaki M, Kholoussi S, Ismail S, Raouf HA, Helwa I, Hassan N, *et al.* Metabolic abnormalities in young Egyptian women with polycystic ovary syndrome and their relation to ADIPOQ gene variants and body fat phenotype. Egyptian J Med Hum Gen. 2015;16:367-374.

43. Stratigopoulos G, Padilla SL, LeDuc CA, Watson E, Hattersley AT, McCarthy MI, *et al.* Regulation of *Fto/Ftm* gene expression in mice and humans. Am J Physiol Regul Integr Comp Physiol. 2008;294:R1185-R1196.

44. Attaoua R, El Mkadem SA, Radian S, Fica S, Hanzu F, Albu A, *et al. FTO* gene associates to metabolic syndrome in women with polycystic ovary syndrome. Biochem Biophys Res Commun. 2008;373:230-234.

45. Grant SF, Bradfield JP, Zhang H, Wang K, Kim CE, Annaiah K, *et al.* Investigation of the locus near *MC4R* with childhood obesity in Americans of European and African ancestry. Obesity. 2009;17:1461-1465.

46. Yang Y-k, Dickinson CJ, Zeng Q, Li J-Y, Thompson DA, Gantz I. Contribution of melanocortin receptor exoloops to Agoutirelated protein binding. J Biol Chem. 1999;274:14100-14106.

47. Sarvestani FS, Tamadon A, Hematzadeh A, Jahanara M, Shirazi MRJ, Moghadam A, *et al*. Expression of *melanocortin-4 receptor* and *agouti-related peptide* mRNAs in arcuate nucleus during long term malnutrition of female ovariectomized rats. Iran J Basic Med Sci. 2015;18:104-107.

48. Zandi MR, Jafarzadeh Shirazi MR, Tamadon A, Akhlaghi A, Salehi MS, Niazi A, *et al.* Hypothalamic expression of *melanocortin-4 receptor* and *agouti-related peptide* mRNAs during the estrous cycle of rats. Int J Mol Cell Med. 2014;3:183-189.

49. Asadi-Yousefabad S-L, Sabet Sarvestani F, Tamadon A, Jafarzadeh Shirazi MR, Ahmadloo S, Moghadam A, *et al. Agouti-related peptide* and *melanocortin-4 receptor* mRNAs expressions in arcuate nucleus during the pregnancy and lactation of rats. Vet Arh. 2015;85:689-700.

50. Nooranizadeh MH, Rahmanifar F, Ahmadloo S, Shaaban Z, Shirazi MRJ, Tamadon A. Enhancement of *melanocortin-4 receptor* (*MC4R*) and constancy of *Kiss1* mRNAs expression in the hypothalamic arcuate nucleus in a model of polycystic ovary syndrome rat. Galen Med J. 2018;7:e1070.

51. Raghow R, Yellaturu C, Deng X, Park EA, Elam MB. SREBPs: the crossroads of physiological and pathological lipid homeostasis. Trends Endocrinol Metab. 2008;19:65-73.

52. Wild RA, Rizzo M, Clifton S, Carmina E. Lipid levels in polycystic ovary syndrome: systematic review and meta-

analysis. Fertil Steril. 2011;95:1073-1079.

53. Zhao C, Dahlman-Wright K. Liver X receptor in cholesterol metabolism. J Endocrinol. 2010;204:233-240.

54. Horton JD, Goldstein JL, Brown MS. SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. J Clin Invest. 2002;109:1125-1131.

55. Malin SK, Kirwan JP, Sia CL, González F. Glucose-stimulated oxidative stress in mononuclear cells is related to pancreatic β -cell dysfunction in polycystic ovary syndrome. J Clin Endocrinol Metab. 2014;99:322-329.

56. Ojeda-Ojeda M, Murri M, Insenser M, F Escobar-Morreale H. Mediators of low-grade chronic inflammation in polycystic ovary syndrome (PCOS). Curr Pharm Des. 2013;19:5775-5791.
57. González F. Inflammation in polycystic ovary syndrome: underpinning of insulin resistance and ovarian dysfunction. Steroids. 2012;77:300-305.

58. Spritzer PM, Lecke SB, Satler F, Morsch DM. Adipose tissue dysfunction, adipokines, and low-grade chronic inflammation in polycystic ovary syndrome. Reproduction. 2015;149:R219-R227.

59. Deligeoroglou E, Kouskouti C, Christopoulos P. The role of genes in the polycystic ovary syndrome: predisposition and mechanisms. Gynecol Endocrinol. 2009;25:603-609.

60. Vural P, Değirmencioğlu S, Saral NY, Akgül C. *Tumor necrosis factor* α (- 308), *interleukin-6* (- 174) and *interleukin-10* (- 1082) gene polymorphisms in polycystic ovary syndrome. Eur J Obstet Gynecol Reprod Biol. 2010;150:61-65.

61. Eriksen MB, Brusgaard K, Andersen M, Tan Q, Altinok ML, Gaster M, *et al.* Association of polycystic ovary syndrome susceptibility single nucleotide polymorphism rs2479106 and PCOS in Caucasian patients with PCOS or hirsutism as referral diagnosis. Eur J Obstet Gynecol Reprod Biol. 2012;163:39-42.

62. Gao J, Xue J-D, Li Z-C, Zhou L, Chen C. The association of *DENND1A* gene polymorphisms and polycystic ovary syndrome risk: a systematic review and meta-analysis. Arch Gynecol Obstet. 2016;294:1073-1080.

63. Bao S, Cai J-H, Yang S-Y, Ren Y, Feng T, Jin T, *et al*. Association of DENND1A gene polymorphisms with polycystic ovary syndrome: a meta-analysis. J Clin Res Pediatr Endocrinol. 2016;8:135.

64. Dallel M, Sarray S, Douma Z, Hachani F, Al-Ansari AK, Letaifa DB, *et al.* Differential association of *DENND1A* genetic variants with polycystic ovary syndrome in Tunisian but not Bahraini Arab women. Gene. 2018;647:79-84.

65. Li Y, Liu H, Zhao H, Xu C, Zhao Y, Ma J, *et al.* Association of AQP8 in women with PCOS. Reprod Biomed Online. 2013;27:419-422.

66. Li T, Zhao H, Zhao X, Zhang B, Cui L, Shi Y, *et al*. Identification of *YAP1* as a novel susceptibility gene for polycystic ovary syndrome. J Med Genet. 2012;49:254-257.

67. Diao X, Han T, Zhang Y, Ma J, Shi Y, Chen Z-J. Family association study between *tumour necrosis factor a* gene polymorphisms and polycystic ovary syndrome in Han Chinese. Reprod Biomed Online. 2014;29:581-587.

68. Yang J, Zhong T, Xiao G, Chen Y, Liu J, Xia C, *et al.* Polymorphisms and haplotypes of the TGF- $\beta 1$ gene are associated with risk of polycystic ovary syndrome in Chinese Han women. Eur J Obstet Gynecol Reprod Biol. 2015;186:1-7.

69. Peral Bn, San Millán JL, Castello R, Moghetti P, Escobar-Morreale HcF. The methionine 196 arginine polymorphism in exon 6 of the *TNF receptor 2* gene (*TNFRSF1B*) is associated with the polycystic ovary syndrome and hyperandrogenism. J Clin Endocrinol Metab. 2002;87:3977-3983.

70. Tumu VR, Govatati S, Guruvaiah P, Deenadayal M, Shivaji S, Bhanoori M. An interleukin-6 gene promoter polymorphism is associated with polycystic ovary syndrome in South Indian women. J Assist Reprod Genet. 2013;30:1541-1546.

71. Guo R, Zheng Y, Yang J, Zheng N. Association of *TNF-alpha*, *IL-6* and *IL-1beta* gene polymorphisms with polycystic ovary syndrome: a meta-analysis. BMC Genetics. 2015;16:5.

72. Deligeoroglou E, Vrachnis N, Athanasopoulos N, Iliodromiti Z, Sifakis S, Iliodromiti S, *et al.* Mediators of chronic inflammation in polycystic ovarian syndrome. Gynecol Endocrinol. 2012;28:974-978.

73. Xiong Y-l, Liang X-y, Yang X, Li Y, Wei L-n. Low-grade chronic inflammation in the peripheral blood and ovaries of women with polycystic ovarian syndrome. Eur J Obstet Gynecol Reprod Biol. 2011;159:148-150.

74. Kim YY, Tamadon A, Ku S-Y. Potential use of antiapoptotic proteins and noncoding RNAs for efficient in vitro follicular maturation and ovarian bioengineering. Tissue Eng Part B Rev. 2017;23:142-158.

75. Hotamisligil G. The role of $TNF\alpha$ and TNF receptors in obesity and insulin resistance. J Intern Med. 1999;245:621-625.

76. González F, Sia CL, Bearson DM, Blair HE. Hyperandrogenism induces a proinflammatory $TNF\alpha$ response to glucose ingestion in a receptor-dependent fashion. J Clin Endocrinol Metab. 2014;99:E848-E854.

77. Szczuko M, Zapałowska-Chwyć M, Maciejewska D, Drozd A, Starczewski A, Stachowska E. High glycemic index diet in PCOS patients. The analysis of IGF I and TNF- α pathways in metabolic disorders. Med Hypotheses. 2016;96:42-47.

78. Hotamisligil GS, Peraldi P, Budavari A, Ellis R, White MF, Spiegelman BM. IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α -and obesity-induced insulin resistance. Science. 1996;271:665-670.

79. Spaczynski RZ, Arici A, Duleba AJ. Tumor necrosis factor- α stimulates proliferation of rat ovarian theca-interstitial cells.

Biol Reprod. 1999;61:993-998.

80. Hong L, Zhang Y, Wang Q, Han Y, Teng X. Effects of interleukin 6 and tumor necrosis factor- α on the proliferation of porcine theca interna cells: Possible role of these cytokines in the pathogenesis of polycystic ovary syndrome. Taiwan J Obstet Gynecol. 2016;55:183-187.

81. Fernández-Real JM, Gutiérrez C, Ricart W, Castiñeira Ma-J, Vendrell J, Richart C. Plasma levels of the soluble fraction of tumor necrosis factor receptors 1 and 2 are independent determinants of plasma cholesterol and LDL-cholesterol concentrations in healthy subjects. Atherosclerosis. 1999;146:321-327.

82. Peral B, San Millán JL, Castello R, Moghetti P, Escobar-Morreale HcF. The methionine 196 arginine polymorphism in exon 6 of the *TNF receptor 2* gene (*TNFRSF1B*) is associated with the polycystic ovary syndrome and hyperandrogenism. J Clin Endocrinol Metab. 2002;87:3977-3983.

83. Päth Gn, Bornstein SR, Ehrhart-Bornstein M, Scherbaum WA. Interleukin-6 and the interleukin-6 receptor in the human adrenal gland: expression and effects on steroidogenesis. J Clin Endocrinol Metab. 1997;82:2343-2349.

84. Marat AL, Dokainish H, McPherson PS. DENN domain proteins: regulators of Rab GTPases. J Biol Chem. 2011;286:13791-13800.

85. Tee MK, Speek M, Legeza B, Modi B, Teves ME, McAllister JM, *et al*. Alternative splicing of *DENND1A*, a PCOS candidate gene, generates variant 2. Mol Cell Endocrinol. 2016;434:25-35.

86. McAllister JM, Modi B, Miller BA, Biegler J, Bruggeman R, Legro RS, *et al.* Overexpression of a *DENND1A* isoform produces a polycystic ovary syndrome theca phenotype. Proc Natl Acad Sci U S A. 2014:201400574.