

Four main therapeutic keys for Parkinson's disease: A mini review

Daniel Hernandez-Baltazar ^{1*}, Rasajna Nadella ², Laura Mireya Zavala-Flores ³, Christian de Jesús Rosas-Jarquín ⁴, María de Jesús Roviroso-Hernández ⁴, Arnulfo Villanueva-Olivo ⁵

¹ CONACYT-Instituto de Neurootología, Universidad Veracruzana, Xalapa, Veracruz, Mexico

² IIT Srikakulam, Rajiv Gandhi University of Knowledge Technologies (RGUKT); International collaboration ID:1840; India

³ Centro de Investigación Biomedica del Noreste. IMSS. Monterrey, Nuevo Leon. Mexico

⁴ Instituto de Neurootología, Universidad Veracruzana. Xalapa, Veracruz, Mexico

⁵ Facultad de Medicina. Universidad Autonoma de Nuevo Leon. Monterrey, Nuevo Leon, Mexico

ARTICLE INFO

Article type:

Mini review article

Article history:

Received: Jul 24, 2018

Accepted: Jan 8, 2019

Keywords:

Cell death
Dopaminergic neurons
Inflammation
Survival
Therapeutics

ABSTRACT

Objective(s): Parkinson's disease (PD) is characterized by motor and cognitive dysfunctions. The progressive degeneration of dopamine-producing neurons that are present in the *substantia nigra pars compacta* (SNpc) has been the main focus of study and PD therapies since ages.

Materials and Methods: In this manuscript, a systematic revision of experimental and clinical evidence of PD-associated cell process was conducted.

Results: Classically, the damage in the dopaminergic neuronal circuits of SNpc is favored by reactive oxidative/nitrosative stress, leading to cell death. Interestingly, the therapy for PD has only focused on avoiding the symptom progression but not in finding a complete reversion of the disease. Recent evidence suggests that the renin-angiotensin system imbalance and neuroinflammation are the main keys in the progression of experimental PD.

Conclusion: The progression of neurodegeneration in SNpc is due to the complex interaction of multiple processes. In this review, we analyzed the main contribution of four cellular processes and discussed in the perspective of novel experimental approaches.

► Please cite this article as:

Hernandez-Baltazar D, Nadella R, Zavala-Flores LM, Rosas-Jarquín ChD, Roviroso-Hernández MDJ, Villanueva-Olivo A. Four main therapeutic keys for Parkinson's disease: A minireview. Iran J Basic Med Sci 2019; 22:716-721. doi: 10.22038/ijbms.2019.33659.8025

Introduction

Parkinson's disease (PD) is the second most common neurodegenerative disease worldwide, with high annual costs of treatment (1). This progressive neurological disorder is characterized by gradual progression of neuronal damage in various motor and non-motor circuits (2). Currently, PD affects the adult population (>65 years) and even young people (3). PD affects a wide variety of nuclei in the central nervous system (CNS), including the dorsal motor nucleus of the vagus, raphe nuclei, *locus coeruleus*, pontine peduncle nucleus, retrorubral nucleus, parabrachial nucleus, the ventral tegmental area, *substantia nigra pars compacta* (SNpc), and *substantia nigra pars reticulata* (SNpr) (4). The degenerative process develops mainly in the dopaminergic neurons (DN), which exhibit native susceptibility to degeneration (5). In humans and the experimental models of PD, the loss of dopaminergic neurons from the SNpc drastically reduces the striatal dopamine concentration (6, 7) promoting motor imbalance, the main characteristic feature that is explored in clinical treatments.

Conventional therapies for Parkinson's disease

Since years ago, the most commonly used PD treatments has included surgical methods like-pallidotomy or deep brain stimulation (DBS) and pharmacological therapy for each and every PD

symptom (8-10). DBS is good at reducing the neuronal loss, avoiding motor fluctuations and preventing damage to the adjacent neurons. On the other hand, DBS is expensive, may cause akinesia and dyskinesia and presence of high risk due to surgical intervention.

Pharmacological therapy with levodopa (11-13) is specific to the dopaminergic system and decreases motor symptoms; however, it promotes hypersensitivity of receptors and overdoses induce dyskinesia. On the other hand, adenosine A2A (14) decreases dyskinesia, inducing low neuroinflammation, but sleep disorders and anxiety are reported. Oral administration of monoamine oxidase type B (MAO-B) inhibitors (15-17) decreases motor disability, prevents the production of free radicals and increases the levels of trophic factors in neurons. However it is not specific for the dopaminergic system, and long-term use may lead to hypertensive crisis, cerebrovascular accident, and weight gain. The oral or subcutaneous use of dopamine agonists (18, 19) lead to neuroprotection of the nigrostriatal pathway, but hallucinations, edema, and addiction have been reported as adverse effects.

The effectiveness of both pharmacological and gene therapy treatments depends on the level of brain neurodegeneration, and thus, determination of cellular processes at neurodegeneration is the key to improving the treatment effectivity.

*Corresponding author: Daniel Hernandez-Baltazar. Avenida Dr Luis Castelazo S/N. Km. 3.5. Carretera Federal. Col. Industrial Animas. C. P. 91190. Xalapa, Veracruz, Mexico. +52 228 841-89-00 ext. 13619; Email: danielhernandez@uv.mx

Cellular process associated with degeneration in substantia nigra pars compacta

Oxidative stress

Many scientific reports have demonstrated that oxidative stress produces neurodegeneration (20, 21). In normal conditions of the cell, the reactive oxygen (ROS) and nitrogen (RNS) species act as secondary messengers in cell processes, however, an excess of ROS is responsible for cell degeneration (22, 23). Dopaminergic neurons of SNpc are more susceptible to oxidative/nitrosative damage because they have low levels of glutathione peroxidase and vitamin E; as well as high levels of free iron (pro-oxidant), monoamine oxidase, and neuromelanin (5, 24, 25), for this the intracellular accumulation of ROS can induce mitochondrial respiratory chain blocking, increase of glutamate, and stimulation of NMDA receptors (4) to finally produce excitotoxicity (26) and cell death by necrosis and apoptosis (27). Additionally, in PD animal models has been shown that complex axonal arborization, elevated mitochondrial bioenergetics (28-30), and selective vulnerability of neuronal populations (31) could contribute to the speed of progression of neurodegeneration. Reverting the damage might be possible by controlling or modifying the ROS/RNS, which is one possible key for PD therapy.

Cell death

PD is characterized by programmed cell death, which is a homeostatic regulatory function of cells that requires energy in the form of ATP. This programmed cell death is of three types: type I cell death or apoptosis, type II cell death or autophagic cell death, and type III cell death or cytoplasmic cell death (32). In all three types the imbalance of mitochondrial bioenergetics favors DN degeneration in PD (33), which results in alterations of genes such as *alpha-synuclein*, *SNCA*, *PINK 1*, *DJ-1*, *LRRK2*, *ATP13A2*, *PLA2G6*, *FBX07*, and *VPS35* (34-36). In experimental studies three cell death types associated with DN damage have been identified, which include mitophagy (37, 38), autophagy (39, 40), and caspase-3-related apoptosis. Cellular stress can induce activation of caspase-3 by extrinsic and intrinsic pathways of apoptosis in the SNpc (41, 42) and favor the expression of pro-apoptotic genes such as *Bax* and *Bad* similar to ischemic stroke (43). In experimental models, it has linked the role of caspase-3, glycogen synthase kinase 3-beta (GSK3 β) and protein kinase C δ (PKC δ) as a switch between neurodegeneration and regeneration (42, 44, 45). As apoptosis is the most reported, development of new drugs that could modulate the pathways and direct towards neuronal survival would be one possible key for PD therapy.

Neuroinflammation

As per Grunewald *et al.* (37), most studies exhibit the neurons as protagonists in PD. However, the participation of other brain populations gives evidence of a complex phenomenon. The neuroinflammation in PD is also characterized by the presence of increased number of activated microglia and astrocytes around the degenerated neurons (46).

Under high oxidative stress conditions, microglial cells release reactive oxygen / nitrogen species (H₂O₂,

-NO₃) and pro-inflammatory cytokines (IL-1 β , IL-6 and TNF α) (47), which serve as signals for the recruitment of more microglial cells, causing imbalance in both neuronal growth and in the release of neurotrophic factors (47). The microglia populations present in damaged SNpc can correspond to two opposite types of microglia, cytotoxic (M1 type) and neuroprotective (M2 type) (48). In experimental models of PD, the cytotoxic microglia (M1) have been evidenced during the progress of DN degeneration in SNpc as a consequence of ROS increase, Lewy bodies (LB) formation, and cell death; stimuli as aggregated alpha-synuclein in Lewy bodies may activate M1-microglia and favor the release of pro-inflammatory responses.

In human post-mortem samples, the alpha-synuclein protein, the main component of LB, has been found in the pre-synaptic terminals of neurons and axons (49). Based on the presence of LB three phases of degenerative damage have been described: 1) LB positive (LB+) neurons without microglia involvement, 2) LB+ neurons with recruited microglia, and 3) LB+ neurons with activated astrocytes. For treating PD the knowledge of the stage-specific switching of M1/M2 phenotypes could be used in therapeutic approaches (48, 50-52).

On the other hand, after neuronal injury, mature astrocytes proliferate and acquire stem cell properties (53-55) promoting neuronal regeneration by synthesizing neurotrophic factors such as glia-derived neurotrophic factor (GDNF) (56) and cerebral dopamine neurotrophic factor (CDNF) (57), and recovery of brain blood irrigation *via* angiotensin type 2 (AT2) (58), the most important effector peptide of the renin-angiotensin system (RAS) (59). Finding a drug that could induce any of the glia to produce more neurotrophic factor or to release anti-inflammatory cytokine production will be a possible key for PD therapy.

Renin-angiotensin 2 system (RAS)

The actions of angiotensin 2 (AT2) are mediated by AT1 and AT2-receptors. AT2 increases the differentiation of precursor cells in dopaminergic neurons *via* activation of AT2-receptor (60, 61). It has also been observed that activation of AT2-receptor may inhibit the production of NADPH oxidase (62), supporting the neuroprotective effect due to RAS. However, the overproduction of AT2 could induce inflammation by promoting oxidative stress derived from NADPH *via* AT1-receptors (63, 64), which proposes the amplifying effect of AT2 during dopaminergic degeneration (6, 62). Interestingly, in PD patients increased local and peripheral levels of angiotensin are associated with motor and non-motor symptoms (59, 65-69).

In experimental models of PD, the high levels of AT2 and ROS induce increased neuron/glia type 2 (NG2) populations (70, 71), precursor cells of immature neurons, oligodendrocytes, Bergmann glia, microglia, and astrocytes depending on the stimulus (46, 57, 72-74). NG2 cells respond very quickly after injury by upregulating the expression of contains chondroitin sulfate proteoglycan 4 (CSPG4) on their surface and exhibit migration and proliferative potential (75-78). Actually, there are no clinical trials evaluating the effect of RAS. Developing or finding a drug that could stimulate

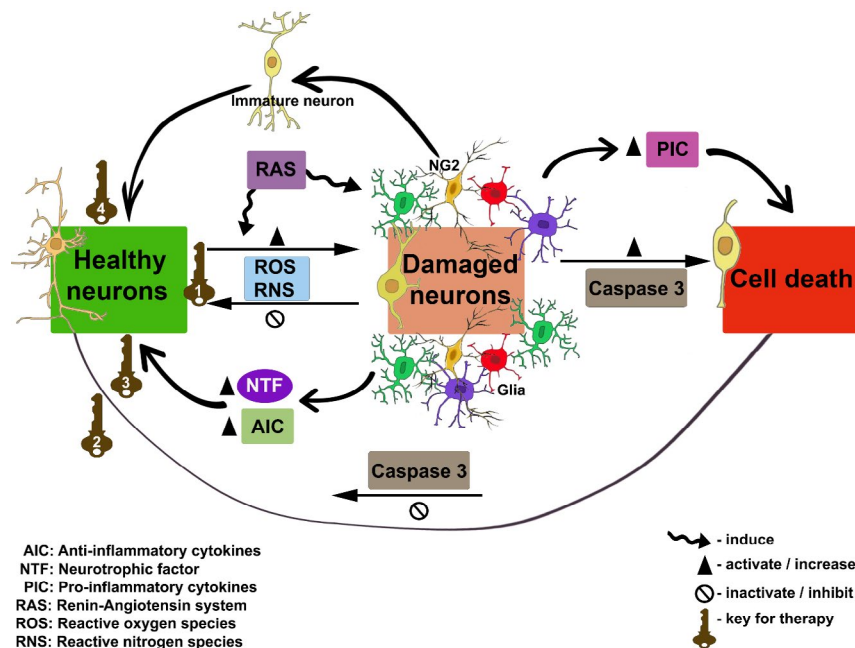


Figure 1. Flow chart of neurodegeneration and hot points for Parkinson's disease therapy

the conversion of NG2 cells to immature neurons would be another possible key for PD therapy.

Novel experimental approaches

As oxidative stress, cell death, neuroinflammation, and RAS system play crucial roles in the degeneration process, new drugs that could control or completely revert stress factors might act as keys for PD therapy (79). The Figure 1 shows the interaction of cellular processes above-revised, the new experimental approaches are focused on some of these hot points. Alternative experimental therapies such as targeted gene delivery, specific drugs, and plant-based anti-oxidant approaches are revised. In animal models, focusing the regulation of cell death, the use of GSK3 inhibitors and the upregulation of chaperone-mediated autophagy (CMA) by retinoic acid derivatives and micro RNAs (miRNAs) have yielded discrete results. The disadvantages of GSK3 inhibitors include the inhibition of kinase leading to severe side-effects due to its multiple cellular targets (44); while the upregulation of CMA could be promising by the use of safety administration route (39). Coupled with this, the use of melatonin as a neuroprotective agent continues to be evaluated (80, 81). In the field of control of ROS and neuroinflammation, pretreatment with synthetic neuromodulators (82), curcumin (83), or other plants derivatives (84, 85) could represent benefits, but further studies on bioavailability, dosage, and biosecurity will be required.

In clinical trials, the capability of GDNF and neurturin to rescue dopaminergic neurons in SNpc (86) has been tested, the results are promising, but due to the lack of safety and specificity, they did not turn out to be a therapeutic medicine (86, 87). In general, targeted gene delivery using viral vectors shows selectivity for dopaminergic neurons, averts neuronal loss, and local increase in the levels of neurotrophic factors that are produced by neurons and glial cells. Unfortunately,

currently, these types of strategies are expensive and require biosafety and must be regulated by turn on/off nanosystems expression.

Conclusion

The multifactorial nature of PD reflects the complex interaction of various cellular processes. The advance in the knowledge of the origin and impact of each related process (stress, neuroinflammation, and cell death) will allow us to better understand the degenerative process and consequently, progress in finding new therapeutic approaches.

Acknowledgment

This work was partially supported by Consejo Nacional de Ciencia y Tecnología (Catedra CONACyT # 1840) for DH-B, and the Instituto de Neuroetología from Universidad Veracruzana (DGI-174332015137) to MJR-H, and the Instituto Mexicano del Seguro Social (FIS/IMSS/PROT/G15/1480) to LMZ-F and PRODEP 29 (UANL-PTC-908) to AV-O. CJR-J received a fellowship from CONACyT for post-graduate studies in Neuroethology (#714879)

Conflicts of Interest

The authors declare that no competing interests exist.

References

- Ascherio A, Schwarzschild MA. The epidemiology of Parkinson's disease: risk factors and prevention. *Lancet Neurol* 2016; 15:1257-1272.
- Moustafa AA, Chakravarthy S, Phillips JR, Gupta A, Keri S, Polner B, *et al.* Motor symptoms in Parkinson's disease: A unified framework. *Neurosci Biobehav Rev* 2016; 68:727-740.
- DeMaagd G, Philip A. Parkinson's Disease and Its Management: Part 5: Treatment of Nonmotor Complications. *P T* 2015; 40:838-846.
- Sulzer D, Surmeier DJ. Neuronal vulnerability, pathogenesis,

- and Parkinson's disease. *Mov Disord* 2013; 28:715-724.
5. Hernandez-Baltazar D, Zavala-Flores LM, Villanueva-Olivo A. The 6-hydroxydopamine model and parkinsonian pathophysiology: Novel findings in an older model. *Neurologia* 2017; 32:533-539.
 6. Villar-Cheda B, Valenzuela R, Rodriguez-Perez AI, Guerra MJ, Labandeira-Garcia JL. Aging-related changes in the nigral angiotensin system enhances proinflammatory and pro-oxidative markers and 6-OHDA-induced dopaminergic degeneration. *Neurobiol Aging* 2012; 33:204 e201-211.
 7. Vogt Weisenhorn DM, Giesert F, Wurst W. Diversity matters - heterogeneity of dopaminergic neurons in the ventral mesencephalon and its relation to Parkinson's Disease. *J Neurochem* 2016; 139 Suppl 1:8-26.
 8. Mazzone P, Lozano A, Stanzione P, Galati S, Scarnati E, Peppe A, et al. Implantation of human pedunculopontine nucleus: a safe and clinically relevant target in Parkinson's disease. *Neuroreport* 2005; 16:1877-1881.
 9. Okun MS. Deep-brain stimulation for Parkinson's disease. *N Engl J Med* 2012; 367:1529-1538.
 10. Okun MS, Gallo BV, Mandybur G, Jagid J, Foote KD, Revilla FJ, et al. Subthalamic deep brain stimulation with a constant-current device in Parkinson's disease: an open-label randomised controlled trial. *Lancet Neurol* 2012; 11:140-149.
 11. Aquilonius SM, Nyholm D. Development of new levodopa treatment strategies in Parkinson's disease-from bedside to bench to bedside. *Ups J Med Sci* 2017; 122:71-77.
 12. Devos D, French DSG. Patient profile, indications, efficacy and safety of duodenal levodopa infusion in advanced Parkinson's disease. *Mov Disord* 2009; 24:993-1000.
 13. Hauser RA, Shulman LM, Trugman JM, Roberts JW, Mori A, Ballerini R, et al. Study of istradefylline in patients with Parkinson's disease on levodopa with motor fluctuations. *Mov Disord* 2008; 23:2177-2185.
 14. Hickey P, Stacy M. Adenosine A2A antagonists in Parkinson's disease: what's next? *Curr Neurol Neurosci Rep* 2012; 12:376-385.
 15. Cruz MP. Xadago (Safinamide): A Monoamine oxidase B inhibitor for the adjunct treatment of motor symptoms in Parkinson's disease. *P T* 2017; 42:622-637.
 16. deSouza RM, Schapira A. Safinamide for the treatment of Parkinson's disease. *Expert Opin Pharmacother* 2017; 18:937-943.
 17. Teo KC, Ho SL. Monoamine oxidase-B (MAO-B) inhibitors: implications for disease-modification in Parkinson's disease. *Transl Neurodegener* 2013; 2:19.
 18. Borovac JA. Side effects of a dopamine agonist therapy for Parkinson's disease: a mini-review of clinical pharmacology. *Yale J Biol Med* 2016; 89:37-47.
 19. Stacy M, Galbreath A. Optimizing long-term therapy for Parkinson disease: levodopa, dopamine agonists, and treatment-associated dyskinesia. *Clin Neuropharmacol* 2008; 31:51-56.
 20. Blesa J, Trigo-Damas I, Quiroga-Varela A, Jackson-Lewis VR. Oxidative stress and Parkinson's disease. *Front Neuroanat* 2015; 9:91.
 21. Dias V, Junn E, Mouradian MM. The role of oxidative stress in Parkinson's disease. *J Parkinsons Dis* 2013; 3:461-491.
 22. Di Meo S, Reed TT, Venditti P, Victor VM. Role of ROS and RNS sources in physiological and pathological conditions. *Oxid Med Cell Longev* 2016; 2016:1245049.
 23. Sovolyova N, Healy S, Samali A, Logue SE. Stressed to death - mechanisms of ER stress-induced cell death. *Biol Chem* 2014; 395:1-13.
 24. Gonzalez-Hernandez T, Cruz-Muros I, Afonso-Oramas D, Salas-Hernandez J, Castro-Hernandez J. Vulnerability of mesostriatal dopaminergic neurons in Parkinson's disease. *Front Neuroanat* 2010; 4:140-153.
 25. Haddad D, Nakamura K. Understanding the susceptibility of dopamine neurons to mitochondrial stressors in Parkinson's disease. *FEBS Lett* 2015; 589:3702-3713.
 26. Labandeira-Garcia JL, Rodriguez-Pallares J, Dominguez-Meijide A, Valenzuela R, Villar-Cheda B, Rodriguez-Perez AI. Dopamine-angiotensin interactions in the basal ganglia and their relevance for Parkinson's disease. *Mov Disord* 2013; 28:1337-1342.
 27. Ghavami S, Shojaei S, Yeganeh B, Ande SR, Jangamreddy JR, Mehrpour M, et al. Autophagy and apoptosis dysfunction in neurodegenerative disorders. *Prog Neurobiol* 2014; 112:24-49.
 28. Franco-Iborra S, Perier C. Neurodegeneration: The Size Takes It All. *Curr Biol* 2015; 25:R797-800.
 29. Giguere N, Trudeau LE. [Axon arborization size is a key factor influencing cellular bioenergetics and vulnerability of dopamine neurons in Parkinson's disease]. *Med Sci (Paris)* 2016; 32:342-344.
 30. Pacelli C, Giguere N, Bourque MJ, Levesque M, Slack RS, Trudeau LE. Elevated mitochondrial bioenergetics and axonal arborization size are key contributors to the vulnerability of dopamine neurons. *Curr Biol* 2015; 25:2349-2360.
 31. Giguere N, Burke Nanni S, Trudeau LE. On cell loss and selective vulnerability of neuronal populations in Parkinson's disease. *Front Neurol* 2018; 9:455.
 32. Venderova K, Park DS. Programmed cell death in Parkinson's disease. *Cold Spring Harb Perspect Med* 2012; 2.
 33. Van Laar VS, Berman SB. The interplay of neuronal mitochondrial dynamics and bioenergetics: implications for Parkinson's disease. *Neurobiol Dis* 2013; 51:43-55.
 34. Hernandez DG, Reed X, Singleton AB. Genetics in Parkinson disease: Mendelian versus non-Mendelian inheritance. *J Neurochem* 2016; 139 Suppl 1:59-74.
 35. Kalinderi K, Bostantjopoulou S, Fidani L. The genetic background of Parkinson's disease: current progress and future prospects. *Acta Neurol Scand* 2016; 134:314-326.
 36. Klein C, Westenberger A. Genetics of Parkinson's disease. *Cold Spring Harb Perspect Med* 2012; 2:a008888.
 37. Grunewald A, Kumar KR, Sue CM. New insights into the complex role of mitochondria in Parkinson's disease. *Prog Neurobiol* 2018.
 38. Yakhine-Diop SMS, Niso-Santano M, Rodriguez-Arribas M, Gomez-Sanchez R, Martinez-Chacon G, Uribe-Carretero E, et al. Impaired mitophagy and protein acetylation levels in fibroblasts from Parkinson's disease patients. *Mol Neurobiol* 2018.
 39. Campbell P, Morris H, Schapira A. Chaperone-mediated autophagy as a therapeutic target for Parkinson disease. *Expert Opin Ther Targets* 2018:1-10.
 40. Zhang J, Cao R, Cai T, Aschner M, Zhao F, Yao T, et al. The role of autophagy dysregulation in manganese-induced dopaminergic neurodegeneration. *Neurotox Res* 2013; 24:478-490.
 41. Hughes AJ, Daniel SE, Lees AJ. The clinical features of Parkinson's disease in 100 histologically proven cases. *Adv Neurol* 1993; 60:595-599.
 42. Hernandez-Baltazar D, Mendoza-Garrido ME, Martinez-Fong D. Activation of GSK-3beta and caspase-3 occurs in Nigral dopamine neurons during the development of apoptosis activated by a striatal injection of 6-hydroxydopamine. *PLoS One* 2013; 8:e70951.
 43. Blanco-Alvarez VM, Lopez-Moreno P, Soto-Rodriguez G, Martinez-Fong D, Rubio H, Gonzalez-Barrios JA, et al. Subacute zinc administration and L-NAME caused an increase of NO, zinc, lipoperoxidation, and caspase-3 during a cerebral hypoxia-ischemia process in the rat. *Oxid Med Cell Longev*

- 2013; 2013:240560.
44. Duda P, Wisniewski J, Wojtowicz T, Wojcicka O, Jaskiewicz M, Drulis-Fajdasz D, *et al.* Targeting GSK3 signaling as a potential therapy of neurodegenerative diseases and aging. *Expert Opin Ther Targets* 2018;1-16.
45. Shin EJ, Hwang YG, Sharma N, Tran HQ, Dang DK, Jang CG, *et al.* Role of protein kinase Cdelta in dopaminergic neurotoxic events. *Food Chem Toxicol* 2018; 121:254-261.
46. Kettenmann H, Kirchhoff F, Verkhratsky A. Microglia: new roles for the synaptic stripper. *Neuron* 2013; 77:10-18.
47. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal* 2014; 20:1126-1167.
48. Tang Y, Le W. Differential Roles of M1 and M2 Microglia in neurodegenerative diseases. *Mol Neurobiol* 2016; 53:1181-1194.
49. Stefanis L. alpha-Synuclein in Parkinson's disease. *Cold Spring Harb Perspect Med* 2012; 2:a009399.
50. Altenhofer S, Kleikers PW, Radermacher KA, Scheurer P, Rob Hermans JJ, Schiffers P, *et al.* The NOX toolbox: validating the role of NADPH oxidases in physiology and disease. *Cell Mol Life Sci* 2012; 69:2327-2343.
51. Ma MW, Wang J, Dhandapani KM, Brann DW. NADPH Oxidase 2 Regulates NLRP3 Inflammasome Activation in the Brain after Traumatic Brain Injury. *Oxid Med Cell Longev* 2017; 2017:6057609.
52. Flores-Martinez YM, Fernandez-Parrilla MA, Ayala-Davila J, Reyes-Corona D, Blanco-Alvarez VM, Soto-Rojas LO, *et al.* Acute neuroinflammatory response in the Substantia Nigra pars compacta of rats after a local injection of lipopolysaccharide. *J Immunol Res* 2018; 2018:1838921.
53. Buffo A, Rite I, Tripathi P, Lepier A, Colak D, Horn AP, *et al.* Origin and progeny of reactive gliosis: A source of multipotent cells in the injured brain. *Proc Natl Acad Sci U S A* 2008; 105:3581-3586.
54. Doetsch F, Caille I, Lim DA, Garcia-Verdugo JM, Alvarez-Buylla A. Subventricular zone astrocytes are neural stem cells in the adult mammalian brain. *Cell* 1999; 97:703-716.
55. Seri B, Garcia-Verdugo JM, Collado-Morente L, McEwen BS, Alvarez-Buylla A. Cell types, lineage, and architecture of the germinal zone in the adult dentate gyrus. *J Comp Neurol* 2004; 478:359-378.
56. Kitamura Y, Inden M, Minamino H, Abe M, Takata K, Taniguchi T. The 6-hydroxydopamine-induced nigrostriatal neurodegeneration produces microglia-like NG2 glial cells in the rat substantia nigra. *Glia* 2010; 58:1686-1700.
57. Nadella R, Voutilainen MH, Saarma M, Gonzalez-Barrrios JA, Leon-Chavez BA, Jimenez JM, *et al.* Transient transfection of human CDNF gene reduces the 6-hydroxydopamine-induced neuroinflammation in the rat substantia nigra. *J Neuroinflammation* 2014; 11:209.
58. Wright JW, Harding JW. Importance of the brain Angiotensin system in Parkinson's disease. *Parkinsons Dis* 2012; 2012:860923.
59. Labandeira-Garcia JL, Garrido-Gil P, Rodriguez-Pallares J, Valenzuela R, Borrajo A, Rodriguez-Perez AI. Brain renin-angiotensin system and dopaminergic cell vulnerability. *Front Neuroanat* 2014; 8:67.
60. Dominguez-Meijide A, Rodriguez-Perez AI, Diaz-Ruiz C, Guerra MJ, Labandeira-Garcia JL. Dopamine modulates astroglial and microglial activity via glial renin-angiotensin system in cultures. *Brain Behav Immun* 2017; 62:277-290.
61. Rodriguez-Pallares J, Quiroz CR, Parga JA, Guerra MJ, Labandeira-Garcia JL. Angiotensin II increases differentiation of dopaminergic neurons from mesencephalic precursors via angiotensin type 2 receptors. *Eur J Neurosci* 2004; 20:1489-1498.
62. Rodriguez-Pallares J, Rey P, Parga JA, Munoz A, Guerra MJ, Labandeira-Garcia JL. Brain angiotensin enhances dopaminergic cell death via microglial activation and NADPH-derived ROS. *Neurobiol Dis* 2008; 31:58-73.
63. Chao J, Yang L, Buch S, Gao L. Angiotensin II increased neuronal stem cell proliferation: role of AT2R. *PLoS One* 2013; 8:e63488.
64. Li J, Culman J, Hortnagl H, Zhao Y, Gerova N, Timm M, *et al.* Angiotensin AT2 receptor protects against cerebral ischemia-induced neuronal injury. *FASEB J* 2005; 19:617-619.
65. Garrido-Gil P, Rodriguez-Pallares J, Dominguez-Meijide A, Guerra MJ, Labandeira-Garcia JL. Brain angiotensin regulates iron homeostasis in dopaminergic neurons and microglial cells. *Exp Neurol* 2013; 250:384-396.
66. Labandeira-Garcia JL, Rodriguez-Pallares J, Rodriguez-Perez AI, Garrido-Gil P, Villar-Cheda B, Valenzuela R, *et al.* Brain angiotensin and dopaminergic degeneration: relevance to Parkinson's disease. *Am J Neurodegener Dis* 2012; 1:226-244.
67. Munoz A, Garrido-Gil P, Dominguez-Meijide A, Labandeira-Garcia JL. Angiotensin type 1 receptor blockage reduces l-dopa-induced dyskinesia in the 6-OHDA model of Parkinson's disease. Involvement of vascular endothelial growth factor and interleukin-1beta. *Exp Neurol* 2014; 261:720-732.
68. Rocha NP, Scalzo PL, Barbosa IG, de Campos-Carli SM, Tavares LD, de Souza MS, *et al.* Peripheral levels of angiotensins are associated with depressive symptoms in Parkinson's disease. *J Neurol Sci* 2016; 368:235-239.
69. Villar-Cheda B, Costa-Besada MA, Valenzuela R, Perez-Costas E, Melendez-Ferro M, Labandeira-Garcia JL. The intracellular angiotensin system buffers deleterious effects of the extracellular paracrine system. *Cell Death Dis* 2017; 8:e3044.
70. Jones LL, Yamaguchi Y, Stallcup WB, Tuszynski MH. NG2 is a major chondroitin sulfate proteoglycan produced after spinal cord injury and is expressed by macrophages and oligodendrocyte progenitors. *J Neurosci* 2002; 22:2792-2803.
71. Stallcup WB. The NG2 proteoglycan: past insights and future prospects. *J Neurocytol* 2002; 31:423-435.
72. Belachew S, Chittajallu R, Aguirre AA, Yuan X, Kirby M, Anderson S, *et al.* Postnatal NG2 proteoglycan-expressing progenitor cells are intrinsically multipotent and generate functional neurons. *J Cell Biol* 2003; 161:169-186.
73. Chung SH, Guo F, Jiang P, Pleasure DE, Deng W. Olig2/Plp-positive progenitor cells give rise to Bergmann glia in the cerebellum. *Cell Death Dis* 2013; 4:e546.
74. Tripathi RB, Rivers LE, Young KM, Jamen F, Richardson WD. NG2 glia generate new oligodendrocytes but few astrocytes in a murine experimental autoimmune encephalomyelitis model of demyelinating disease. *J Neurosci* 2010; 30:16383-16390.
75. Chari DM, Blakemore WF. Efficient recolonisation of progenitor-depleted areas of the CNS by adult oligodendrocyte progenitor cells. *Glia* 2002; 37:307-313.
76. Magnus T, Carmen J, Deleon J, Xue H, Pardo AC, Lepore AC, *et al.* Adult glial precursor proliferation in mutant SOD1G93A mice. *Glia* 2008; 56:200-208.
77. Nait-Oumesmar B, Decker L, Lachapelle F, Avellana-Adalid V, Bachelin C, Baron-Van Evercooren A. Progenitor cells of the adult mouse subventricular zone proliferate, migrate and differentiate into oligodendrocytes after demyelination. *Eur J Neurosci* 1999; 11:4357-4366.
78. Tamura Y, Kataoka Y, Cui Y, Takamori Y, Watanabe Y, Yamada H. Multi-directional differentiation of doublecortin- and NG2-immunopositive progenitor cells in the adult rat neocortex in vivo. *Eur J Neurosci* 2007; 25:3489-3498.
79. Charvin D, Medori R, Hauser RA, Rascol O. Therapeutic strategies for Parkinson disease: beyond dopaminergic drugs.

Nat Rev Drug Discov 2018; 17:804-822

80. Carrascal L, Nunez-Abades P, Ayala A, Cano M. Role of melatonin in the inflammatory process and its therapeutic potential. *Curr Pharm Des* 2018; 24:1563-1588.

81. Sanchez-Barcelo EJ, Rueda N, Mediavilla MD, Martinez-Cue C, Reiter RJ. Clinical Uses of Melatonin in Neurological Diseases and Mental and Behavioural Disorders. *Curr Med Chem* 2017; 24:3851-3878.

82. Martinez B, Peplow PV. Neuroprotection by immunomodulatory agents in animal models of Parkinson's disease. *Neural Regen Res* 2018; 13:1493-1506.

83. Wang YL, Ju B, Zhang YZ, Yin HL, Liu YJ, Wang SS, *et al*. Protective effect of curcumin against oxidative stress-induced injury in rats with Parkinson's disease through the Wnt/beta-catenin signaling pathway. *Cell Physiol Biochem* 2017;

43:2226-2241.

84. Ullah H, Khan H. Anti-Parkinson potential of silymarin: mechanistic insight and therapeutic standing. *Front Pharmacol* 2018; 9:422.

85. Zanforlin E, Zagotto G, Ribaldo G. The medicinal chemistry of natural and semisynthetic compounds against Parkinson's and Huntington's diseases. *ACS Chem Neurosci* 2017; 8:2356-2368.

86. Kirik D, Cederfjall E, Halliday G, Petersen A. Gene therapy for Parkinson's disease: Disease modification by GDNF family of ligands. *Neurobiol Dis* 2017; 97:179-188.

87. Pignataro D, Sucunza D, Rico AJ, Dopeso-Reyes IG, Roda E, Rodriguez-Perez AI, *et al*. Gene therapy approaches in the non-human primate model of Parkinson's disease. *J Neural Transm (Vienna)* 2018; 125:575-589.