

## The genus *Cuscuta* (Convolvaceae): An updated review on indigenous uses, phytochemistry, and pharmacology

Shazia Noreen<sup>1</sup>, Sobia Noreen<sup>1\*</sup>, Shazia Akram Ghumman<sup>2</sup>, Fozia Batool<sup>1</sup>, Syed Nasir Abbas Bukhari<sup>3</sup>

<sup>1</sup> Department of Chemistry, University of Sargodha, Sargodha-40100, Pakistan

<sup>2</sup> College of Pharmacy, University of Sargodha, Sargodha-40100, Pakistan

<sup>3</sup> Department of Pharmaceutical Chemistry, College of Pharmacy, Jouf University, Aljouf, Sakaka2014, Saudi Arabia

### ARTICLE INFO

#### Article type:

Review article

#### Article history:

Received: Oct 23, 2018

Accepted: May 10, 2019

#### Keywords:

Bioactive

*Cuscuta*

Folk medicines

Pharmacological activities

Phytochemicals

### ABSTRACT

*Cuscuta*, commonly known as dodder, is a genus of family convolvaceae. Approximately 170 species of *Cuscuta* are extensively distributed in temperate and subtropical areas of the world. Species of this genus are widely used as essential constituents in functional foods and traditional medicinal systems. Various parts of many members of *Cuscuta* have been found efficacious against a variety of diseases. Phytochemical investigations have confirmed presence of biologically active moieties such as flavonoids, alkaloids, lignans, saponines, phenolics, tannins, and fatty acids. Pharmacological studies and traditional uses of these plants have proved that they are effective antibacterial, antioxidant, antiostoporotic, hepatoprotective, anti-inflammatory, antitumor, antipyretic, antihypertensive, analgesic, anti hair fall, and antisteriogenic agents.

#### ► Please cite this article as:

Noreen Sh, Noreen S, Ghumman ShA, Batool F, Bukhari SNA. The genus *Cuscuta* (Convolvaceae): An updated review on indigenous uses, phytochemistry, and pharmacology. Iran J Basic Med Sci 2019; 22:1225-1252. doi: 10.22038/ijbms.2019.35296.8407

### Introduction

Plant-based medicines are an integral part of virtually all cultures since immemorial times. The journey of information from prehistoric texts to various indigenous folklores and modern preparations have witnessed the presence of bioactive moieties with therapeutic potential in these herbs (1-4). The immense population of current allopathic products is embedded in nature. More than half of the clinically approved drugs in the world are either natural products or their modifications. Higher plants being an endless reservoir contribute above one fourth. The remarkable resurgence of interest in nature to explore pharmaceutical and nutraceutical agents is still marching towards new horizons (5-7).

Ever growing consumption of natural products by local masses has forcefully motivated the scientists to acquire systematic, elaborated, and practical knowledge about their constituents by using advanced technologies (8). Herbal products, both as purified compounds and in the form of standard extracts, offer infinite odds for novel pharmaceutical products due to the matchless accessibility to different chemical species (9). Target-based phytochemicals have transfigured the medicinal industry because these are not only directly utilized for treatment purposes but also act as leads and standard template for synthetic drugs (10-11). Therefore, modern scientific investigations are turning towards traditional medicines to look for new windows of opportunities giving rise to superior pharmacologically active agents against diseases (12).

The genus *Cuscuta* L. commonly known as dodder is one of the essential herbal constituents of pharma foods and curative tonics that are frequently prescribed to nourish various body parts. It is used to enhance the

nutritional value of porridge and alcoholic beverages (13). The genus has a rich history of folk medicinal uses, and numerous phytoconstituents of therapeutic value have been isolated and identified (14). Various species are indigenous used to cure fits, melancholy, insanity (15), fertility problems (16), tumors (17), scabies, eczema (18), chronic ulcer, jaundice, inflammation (19), chest pain (20), fever, itching (21), osteoporosis (22), diarrhea, oedema, stomach ache, infections, measles, sores, kidney problems (23), sprain (24), alleviation of high blood pressure, leucorrhoea (25), obesity (26), migraine, amnesia, epilepsy, and constipation (27).

Pharmacological analysis of various *Cuscuta* species unveiled their antitumor, antimicrobial (28-31), hepatoprotective (32-33), anticonvulsant (34), immunostimulatory, antioxidant (14, 35-37),  $\alpha$ -glucosidase inhibition (38), psychopharmacological (39), hair-growth promoting (40-41), anti-steroidogenic (42), anti-inflammatory (43-44), diuretic (45), analgesic (46), antipyretic (47-48), anti-HIV (49), antidiabetic (50), neuroprotective (51), antiulcer (52), antispasmodic, hemodynamic, bradycardia<sup>1</sup>, antihypertensive, cardiotoxic, and muscle relaxant activities (53).

*Cuscuta* species are rich in bioactive constituents that exhibit a wide variety of pharmacological activities. Presence of a good deal of valuable components, broad range of biological attributes and remedial value of these plants in folk medicinal systems gives stimulation toward the concept that this genus can play an important role in discovery of new and more efficient therapeutic agents. This review is an effort to edify knowledge of its phytochemical richness, pharmacological and biological significance, and folk medicinal uses, which will enhance its value as a potent pharmaceutical precursor.

\*Corresponding author: Sobia Noreen. Department of Chemistry, University of Sargodha, Sargodha, 40100, Pakistan. Tel: +923018434400; Email: sobianoreen@uos.edu.pk

## Methods

This review on *Cuscuta* genus has been written according to the information collected from various scientific databases such as Scopus, Researchgate, Web of Science, ScienceDirect, and PubMed up to August 2018.

## Distribution and botanical description

*Cuscuta*, a flowering parasitic genus was previously placed in the *Convolvulaceae* family, but later it was segregated as the separate family *Cuscutaceae* (54-57). Global distribution record indicates that most of the species are concentrated in tropical and subtropical areas and fewer in temperate regions. This parasitic genus is known by many common names such as dodder, gold-thread, hair-weed, devil's hair, hell-vine, strangle-vine, love-vine, pull-down, etc. in different regions of the world. The number of species documented by various authors varies from 100 to 170 (58-66). Medicinally important species are *C. reflexa* Roxb. (67), *C. chinensis* Lam. (68), *C. japonica* Choisy (69), *C. australis* R. Br. (70), *C. europaea* Linn. (71), *C. gigantea* Griff. (72), *C. hyalina* Roth. (73), *C. campestris* Yuncker. (47), *C. racemosa* Mart. (52), *C. pedicellata* Ledeb. (74), *C. epithymum* L. (75), *C. kilimanjari* Oliv. (76), *C. kotschyana* Boiss. (77), *C. mitraeformis* Engelm. (78), *C. tinctoria* Mart (79), and

*C. capitata* Roxb (80).

*Cuscuta* species are holoparasitic, annual or perennial, herbaceous vines. The thread-like slender, twining stems have orange, red, or yellow color. Majority of the members have achlorophyllous, scaly leaves while some of them are with reduced synthetic apparatus and can perform localized and limited photosynthesis. Bisexual flowers in multiple colors like cream, yellow, white, and pink are pollinated by insects. Roots are absent, and haustoria are used to suck water and nutrients. Several morphological and physiological simplifications, for instance absence of cotyledons or radicles in their embryos, scaly leaves without vascular tissue and haustoria represent an adaptation to parasitism. They are obligate parasitic plants (54, 61, 81-84). These stem and leaf parasites depend entirely on their host plant, thus reducing the growth and yield of the host. They mostly infect many broadleaf crops, ornamentals plants, weeds, and a few monocot crops. Some of the species are strictly host-specific while others thrive on diverse hosts (85, 86). The usual growing season is early summer; germination starts in May, parasites invade the host by haustoria and may wither and die in the absence of a suitable host within two weeks (87). Flowering starts in June and seed production in November (88).

**Table 1.** Common names and global distribution of some medicinally important *Cuscuta* species

Name	Common name	Distribution	References
<i>C. reflexa</i>	Hell weed, devil's gut, beggar weed, strangle tare, scald weed, dodder of thyme, greater dodder, lesser dodder	Pakistan, India, China, E. Asia, Afghanistan, Bangladesh,	(27, 29, 89-90)
<i>C. chinensis</i>	Chinese dodder	Ethiopia, Kazakhstan, Kyrgyzstan, Tajikistan, Turkmenistan, Uzbekistan, Mongolia; Russia, China, Iran, Iraq, Afghanistan, India, Sri Lanka, Indonesia, Korea, Japan, Taiwan, Thailand, Australasia,	(68, 91)
<i>C. japonica</i>	Japanese dodder	Korea	(92-93)
<i>C. australis</i>	Australian dodder, Omonigelegele, southern dodder	Taiwan, Africa, Japan, Australia, Madagascar, Europe, Asia, Senegal, Ethiopia,	(23, 70, 94-96)
<i>C. europaea</i>	.....	India, Romania, Bulgaria, Iran	(97-99)
<i>C. gigantea</i>	.....	Pakistan, China, Afghanistan, Tajikistan.	(62, 72)
<i>C. hyaline</i>	.....	Pakistan, Ethiopia, Sudan, Kenya, Uganda, Burundi, Rwanda, Zimbabwe, India, Botswana, Namibia, South Africa,	(100)
<i>C. planiflora</i>	Small seed dodder, red dodder	North Africa, Southwestern and southern Asia, Ethiopia, Madagascar, Angola	(23, 101-102)
<i>C. campestris</i>	Field dodder, common dodder, prairie dodder, yellow dodder, gewone dodder,	Saudi Arabia, Nigeria, South America, Europe, Asia, Africa, Australia, Taiwan	(81, 86, 103-105)
<i>C. racemosa</i>	Chilean dodder, lead-vine, golden thread	Brazil, Chile	(52, 106)
<i>C. pedicellata</i>	Clover dodder	Pakistan, Egypt, Qatar, Saudi Arabia, UAE, Iran	(26, 99, 107-109)
<i>C. epithymum</i>	Common dodder, Clover dodder, lesser dodder, flax dodder	Pakistan, Ireland, Iran, Poland	(95, 106, 110-112)
<i>C. kilimanjari</i>	Dodder	Sudan, Etopia, Congo, Malawi, Zimbabwe, Mozambique, Limpopo, Madagascar	(23, 95)
<i>C. monogyne</i>	Eastern dodder	Iran	(113)
<i>C. approximata</i>	Alfalfa dodder Smooth seed alfalfa dodder	Turkey, Iran	(14, 114-115)
<i>C. kotschyana</i>	.....	Iran,	(99)
<i>C. capitata</i>	.....	India, Nepal	(80, 116)
<i>C. mitraeformis</i>	.....	México	(78)

C: *Cuscuta*

## Medicinal uses

The local inhabitants of rural areas are aware of inherent properties of various plants. They preferentially use these herbs and their products to treat multiple types of diseases due to their handiness and low cost (117). Potentially useful plants have been acknowledged and sequentially conveyed throughout the centuries in all societies. Some of them are used through self-medication, while others are recommended by traditional healers (118). Plant utilization as medicine ranges from the direct administration of the leaves, seeds, barks, roots, and stems to the extracts and decoctions from different parts of the plants (119).

Many *Cuscuta* species being rich sources of diverse phytochemicals are popular components of various folk medicinal systems. *Cuscuta* species are used in traditional medicine as a purgative, diaphoretic,

anthelmintic, diuretic, and tonic as well as a treatment for itching and bilious disorders (120, 121). Seeds, stem, and whole plant are utilized as prescription to treat different types of ailments. Medicinal uses of several parts of *Cuscuta* members are given in Table 2.

*C. reflexa* is a treasured medicinal herb and widely used in conventional medicinal system of various Asian countries including China, India, Bangladesh, and Thailand for treating multiple disorders (122). It is called a miracle therapeutic plant in the ethnobotany, and a wide array of chemical compounds has been isolated with diverse medicinal properties (123). *C. reflexa* whole plant is used to treat conjunctivitis, respiratory disorders, piles, ulcers, and stomach problems (124). The paste of whole plant mixed with latex *Carica papaya* causes abortions (125). In rural areas of India its juice is used against jaundice. Paste of plant is effective to

**Table 2.** Traditional medicinal uses of some *Cuscuta* species

Species	Plant part	Preparation	Traditional use	References	
<i>C. reflexa</i>	WP*	Paste	Treatment of swollen testicles, gout and joint pain, causes abortion, anti-rheumatic, analgesic	(67, 125, 127-128, 132, 169-170)	
		Maceration	Infection treatment	(149)	
		Infusion	Anti-poisonous	(142)	
		Juice	Antiseptic, useful in itching skin and jaundice	(127, 171)	
		Powder	Anti-fertility agent, astringent, diaphoretic	(136)	
		Pills	Anti-tuberculosis	(89)	
		Decoction	Useful in skin disease, used for jaundice, cough, blood purification, bronchitis, fever, sex stimulation	(171-172)	
		-----	Antidiarrheal, anti-inflammatory, anti-ulcer, purgative, antidiarruff, conjunctivitis, analgesic, hepatoprotective, useful in cough, cephalgia, fever, leucorrhoea, and paralysis, respiratory disorders, piles, stomach problem, constipation, spleen diseases, helminthiasis, fracture joining	(124, 131, 144, 150, 169, 173-175)	
		Stem	Decoction	Hepatoprotective, antidiarrheal, useful in constipation, stomach disorders, urinary tract infections, jaundice, epilepsy, cholera, asthma	(144, 169)
			Paste	Anti-hair fall, anti-rheumatic, useful in skin diseases	(29, 128, 144)
	Seeds	Juice	Jaundice treatment	(126, 176)	
		Crushed	Blood purifier, purgative, good for brain, fever, anthrax in cattle	(135, 138)	
		-----	Effective in bilious disorders and fever	(133-134)	
		Decoction	Cause abortion	(144)	
		-----	Carminative, anthelmintic, alterative, emmenagogue, sedative, diuretic, useful in ulcer, liver disorders	(129, 170)	
		Poultice	Pain reliever	(177)	
		Leaves	Extract	Cold treatment	(178)
	Juice		Anti-hypertensive, anti-diarrheal, useful in jaundice.	(179)	
	-----		Effective in scabies, eczema, inducing sterility	(18, 180)	
<i>C. chinensis</i>	Fruits	-----	Antipyretic, cough reliever	(67)	
		Juice	Anti-ulcer, anti-inflammatory, wound healer, jaundice treatment	(19)	
	WP	Dressing	Useful in painful inflammations	(151)	
		Paste	Anti-ulcer and wound healer	(151)	
	Seeds	-----	Carminative, tonic, diuretic, sedative, diaphoretic	(158)	
		Paste	Joining fractures	(155)	
	Stem	-----	Expectorant, carminative, tonic, anthelmintic, purgative, diaphoretic, anti-inflammatory, analgesic	(158)	
		-----	Antihypertensive	(93)	
	<i>C. japonica</i>	Leaves	-----	Antihypertensive	(93)

Continued Table 2.

<i>C. australis</i>	-----	-----	Laxative, anthelmintic, astringent, emollient, sedative, sudorific, liver and kidney tonic, useful in sores and measles	(23)
<i>C. austrais</i>	seeds	Decoction	Brain tonic	(181)
<i>C. europaea</i>	Sap	-----	Carminative	(71)
	WP	Extract	Anti-psoriasis	(71)
		Juice	Useful in skin diseases	(167)
	Seed,	-----	Laxative, diuretic, analgesic	(116, 166)
	vegetative			
	pant			
<i>C. gigantea</i>	-----	Juice	Antipoisonous	(72, 164)
	-----	-----	Anti-septic	(116)
<i>C. hyalina</i>	WP	-----	Purgative, useful externally against itching and internally in protracted fevers	(21)
		Infusion	Sores washers	(21)
		-----	Abortion treatment	(73)
		-----	Antiulcer, against culex mosquito,	(23)
<i>C. planiflora</i>	WP	-----	Carminative, laxative	(130)
	Stem	-----	Anti-diarrheal	(23)
<i>C. campestris</i>	WP	Decoction	Purgative, useful in constipation,	(105)
		poultice		
<i>C. racemosa</i>	-----	-----	Anti-inflammatory, diuretic, effective in the stomach and hepatic disorders and fresh wounds	(52)
<i>C. pedicellata</i>	-----	-----	Anti-obesity	(26)
	Stem	-----	Purgative, wound healer, anti-inflammatory, antihypertensive, useful in Stomachache	(168)
<i>C. epithimum</i>	WP	-----	Diuretic, laxative, liver and kidney tonic, to treat sciatica, scurvy and scrofula derma	(163, 182)
		-----	Astringent, Laxative, deterrentive	(75)
		Extract	Scleroderma treatment	(162)
	Stem	-----	Useful in epilepsy	(183)
<i>C. kilimanjari</i>	Stem	Sap	Useful in ear, nose and throat diseases	(76)
		-----	Effective in stomach ache, edema, veterinary treatment, agalactia	(23)
	WP	-----		
		Sap	Treatment of ringworm and warts	(79)
<i>C. capitata</i>	WP	Powder	Reduces irritation of bladder and improves urinary function	(80)
			Useful in kidney problems	(116)
<i>C. approximata</i>	WP	-----	Laxative, carminative, hepatoprotective	(130)
	-----	-----	Useful in sin disease	(116)

C: *Cuscuta*; \*Whole plant

treat headache, gout, and rheumatism (67, 126-128). Plant juice mixed with other decoctions is purgative. Seeds of *C. reflexa* are carminative, anthelmintic, alterative, emmenagogue, sedative, and diuretic. It is effective against warts (116, 129). Leaves are used to treat eczema, scabies, cold, and to induce sterility (18, 130). Rabha tribes of west Bengal use the whole plant to treat leucorrhoea (131). It is applied internally to cure protracted fevers and externally on itchy skin. The plant is frequently used in Ayurvedic medicine to give relief in urinating difficulties, muscle pain, and coughs (132, 133). Pills prepared from the dried plant are used for treatment of tuberculosis (89). Its stem is a blood purifier, good for brain and fever (134-135). Tribal people use its various parts to treat fits, insanity, melancholy, and to control fertility (15). It is commonly used in veterinary medicines as poultice and sprains. The powder is used as astringent and diaphoretic for cattle (136-137).

*C. reflexa* stems are crushed with *Clerodendrum viscosum* leaves and fed to cattle to treat anthrax (138). The plant is used for skin infections and dandruff (139-140). The paste of whole plant with *Achyranthes aspera* is used to control excessive bleeding during menstruation (141). It is also used for treatment of bone fracture and body pain (142). In folk medicine of Bangladesh, it is used to cure tumors (17). The Tripura community of Bangladesh and Satar tribes in Nepal use this plant to cure edema, body ache and for maintenance of liver function. It is used for treating constipation, spleen diseases, diarrhea, and inflammation. Paste mixed with sesame oil is applied for curing hair fall. The decoction of stem is used to cure diarrhea, cholera, and asthma, while decoction of seeds causes depression, nausea, and vomiting (29, 143-145). Whole plant powder is used to treat jaundice by tribal people of nallamalais in Andhra Pradesh (146).

It is also used as expectorant, aphrodisiac, is useful

in vomiting, and purifies the blood (32). *C. reflexa* is an essential constituent of several medical compositions, which are used in the treatment of migraine, headache, chronic catarrh, epilepsy, amnesia, and to prolong fever (27, 147-148). Maceration of whole plant is used to treat infections (149). The whole plant is also useful in cephalgia, paralysis, stomach pain and helminthiasis (89, 150).

*C. chinensis* Lam. also known as Chinese dodder or *Tu-Si-Zi*, also has a wide range of uses. It has been mentioned in various old Chinese scripts and recommended by many herbal practitioners (68). Besides China it is also a famous prescription in many other countries. In Pakistan dressing made of plant is used on painful inflammations. Moreover, paste is useful for chronic ulcers and wounds (151). In traditional Indian system, leaves and stems are used to enhance lactation (152). In Vietnam people use whole plant in back pain and constipation (153). In Korea, seeds with other herbal prescriptions are effective to improve sexual function and health (154). Stem paste of *C. chinensis* is applied to fractured bone to promote the joining (155). Whole plant juice is used to treat inflammation and jaundice (19, 156). A lotion prepared from stem is used to treat sore heads and inflamed eyes. It has been found useful in the treatment of impotence, nocturnal emissions, dizziness, lumbago, leucorrhoea, decreased eyesight, abortion, and chronic diarrhea (133). *C. chinensis* is used in treatment of mania, epilepsy, and insanity (157). Its stem and seeds are considered tonic, expectorant, purgative, sedative, diuretic, diaphoretic, carminative, anthelmintic, and advantageous in muscles and joints pain (158-159). Prescriptions containing *C. chinensis* are used to treat impairment of sexual function, cure cardiovascular diseases and osteoporosis, treatment of premature ejaculation, to treat lower abdominal and back pain, infertility, wet dreams, impotence, urinary retention, and urinary incontinence (68). It is also used to cure melisma, freckles and considered as anti-dandruff agent (160-161).

*C. epithymum* is a mild diuretic and used to treat sciatica and scurvy. The fresh plant is applied to the skin against scrofula derma and scleroderma. It is associated with the health of liver and kidneys and used in various formulas. It is considered a mild laxative (162-163). The whole plant is dried and used as astringent and detergent (75). Whole plant decoction of *C. campestris* is used as purgative and poultice (105). The sap of *C. tinctoria* is used to cure ringworm and warts (79). Juice of *C. gigantea* plant is famous as an anti-poisonous agent (140, 164). The sap of *C. europaea* is used as a carminative, and the extract is applied to treat psoriasis (165). Seeds and vegetative parasitic plant is used as laxative, diuretic, and pain reliever and is poisonous. The juice is used for skin treatment (166-167). *C. capitata* whole plant reduces irritation of bladder and improves urinary function (80). *C. hyaline* is used to treat chest pain (20, 24). Its infusion is used as sores washer and to prevent abortion (21, 73). It is antiulcer and used against culex mosquito. *C. australis* is used as laxative, anthelmintic, astringent, for treatment of sores, measles and as kidney and liver tonic, emollient, sedative, and sudorific (23).

Leaves of *C. japonica* are considered antihypertensive

(93). The sap of *C. kilimanjari* collected from stems is directly installed to treat ear, nose, and throat diseases in central Kenya. The whole plant is used to treat stomach ache, edema, agalactia, and in veterinary medicines (23, 76). *C. pedicellate* is used for treatment of obesity, stomachache, to cure wounds, hypertension, as purgative, and anti-inflammatory agent (26, 168). The whole plant of *C. planiflora* is carminative and laxative, and the stem is anti-diarrheal (23, 130). *C. racemosa* has anti-inflammatory and diuretic effects, is also used for stomach and hepatic complaints and treatment of fresh wounds (52).

### Phytochemistry

Exploration of nature's garden of medication to expose more acceptable solutions with safety is a subject of interest from prehistoric era as more than half of world population still relies on medicinal plants to sustain life. The capability of these odds to appease and treat various diseases and infirmity is undoubted. The curative plants are extensively used in pharmaceuticals, food industry mostly as functional food, agricultural, and cosmetics. Various herbs, their extracts, and prescriptions are loaded with different biologically active constituents particularly alkaloids, steroids, saponins, flavonoids, and terpenoids that are responsible for their therapeutic outcomes (27, 184-189). Phytochemical screening of ever more medicinal plants is extremely momentous in detecting and identifying innovative sources of healing as well as commercially important compounds (190).

Genus *Cuscuta* is rich in many phytoconstituents representing a varied spectrum of secondary metabolites including flavonoids, alkaloids, lignans, polysaccharides, steroids, volatile oils, and resin glycosides (191-199). In a comparative study it was suggested that the plants in the *Cuscuta* species are blessed with almost same soluble phenolic secondary metabolites as Chlorogenic acid, 3,5-dicaffeoylquinic acid, 4,5-dicaffeoylquinic acid, hyperoside, quercetin, astragalin, kaempferol-3-O-galactoside, and quercetin-3-O-glucoside but with varying quantities (200).

Chemical constituents of *Cuscuta* species are host-dependent. For instance, a large number of alkaloids identified in these parasitic plants are the same as those found in their alkaloid containing hosts except a very few (201). These species can synthesize flavonoids, while the study of relation between flavonoids of host and parasite is under consideration. Preliminary determination indicates that flavonoid content of various *Cuscuta* samples growing on different hosts is quite different (202). The most thoroughly characterized species of this genus are *C. reflexa* and *C. chinensis* (67-68, 203).

Essential component of many medicinal compositions of *C. reflexa* has an extensively varied array of phytochemicals identified as phenolic compounds, flavonoids, alkaloids, phytosterols, amarbelin, betasterol, stigmasterol, glycosides, saponins, cuscutine, myricetin, dulcitol, coumarin, cuscutamine, luteolin, bergenin, proteins, fixed oils, fats, and carbohydrates (27, 67, 204).

This genus is a source of many novel metabolites. Qualitative analysis of methanolic extract of *C. reflexa* isolated two new compounds named as 7'--(3',4'-dihydroxyphenyl)-N-[(4-methoxyphenyl)ethyl]

propenamide and 7'-(4'-hydroxy,3'-methoxyphenyl)-N-[(4-butylphenyl)ethyl]propenamide (38). From aerial parts of same plant two novel tetrahydrofuran derivatives, namely Swarnalin and Cis-swarnelin were separated (205) while a flavanon, reflexin chemically named as 5-hydroxy-7-methoxy-6-(2,3-epoxy-3-methylbutyl)-flavanone, was isolated from the stem (206). Moreover, 3'-methoxy-3,4',5,7-tetrahydroxy flavone and 3'-methoxy-4',5,7-trihydroxy flavone-3-glucoside were isolated from whole plant (207). An antiviral protein with molecular weight about 14,000–18,000 Daltons was separated and evaluated against several isometric and anisometric viruses (208).

Phytochemical investigations of *C. chinensis* have shown that flavonoids, alkaloids, poly-saccharides, steroids, lignans, and volatile oils are mostly reported in its various parts (68). The active moieties responsible for various pharmacological activities of the *C. chinensis* mostly include flavonoids, lignans, quinic acid, and polysaccharide. Flavonoids are the prime biologically active components in *C. chinensis*. Additionally, quercetin, kaempferol, and hyperoside can serve as an index to evaluate the quality of the crude drug (209).

*C. chinensis* extract afforded four new lignans cuscutoside A (2'-hydroxyl asarinin 2'-O-β-D-apiofuranosyl-(1→2)-β-D-glucopyranoside), cuscutoside B (2'-hydroxyl asarinin 2'-O-β-xylopyranosyl-(1→6)-β-

glucopyranoside), cuscutoside C (2'-hydroxyl asarinin 2'-O-β-D-glucopyranoside), cuscutoside D (2'-hydroxyl asarinin 2'-O-β-D-apiofuranosyl-(1→2)-[β-D-glucopyranosyl-(1→6)]-β-D-glucopyranoside) and neosamin (188, 193, 210). *C. chinensis* and *C. australis* are used to prepare the famous Chinese herbal prescription Tu-Si-Zi. Phytochemical analysis was done to compare the phenolic constituents of both plants. Principal compounds of *C. australis* were kaempferol and astragalol while hyperoside was predominant in *C. chinensis* (211). Several Phytoestrogens were isolated and identified from *C. chinensis*. Ethanolic extract of seeds afforded three new lignans named cuscutarensinols A–C (212). In another investigative study, four new glycosidic acids called cuscuteic acids A-D were isolated from the alkaline hydrolysate of the ether-insoluble resin glycoside (191). Up till now bulk of the phytochemical investigations on *C. chinensis* targeted the seeds while other parts of the plant have had much less attention by the researchers.

An ether insoluble resin glycoside fraction was separated from seeds of *C. australis* and identification and characterization of resin matrix revealed the presence of three new glycosidic acids, cuscuteic acids A<sub>1</sub>–A<sub>3</sub> (213). *C. racemosa* like other species of the genus offers flavonoids as the chief constituent along with tannins. In another experiment alkaloids, flavonoids, tannins, and saponins have been identified (52, 214).

**Table 3.** Phytochemical profile of various *Cuscuta* species

Name	Plant part	Solvent	Extraction	Separation technique	Phytochemicals	References			
<i>C. reflexa</i>	WP	MeOH	Maceration	CC	7'-(3',4'-dihydroxyphenyl)-N-[(4-methoxyphenyl)ethyl]propenamide 7'-(4'-hydroxy,3'-methoxyphenyl)-N-[(4-butylphenyl)ethyl]propenamide 6,7-dimethoxy-2H-1-benzopyran-2-one 2-(3-hydroxy-4-methoxyphenyl)-3,5-dihydroxy-7-O-β-D-glucopyranoside-4H-1-benzopyrane-4-one, 3-(3,4-dihydroxyphenyl)-2-propen-1-ethanoate 6,7,8-trimethoxy-2H-1-benzopyran-2-one 3-(4-O-β-D-glucopyranoside-3, dimethoxyphenyl)-2-propen-1-ol	(38)			
					-----	-----	HPLC	Kaempferol Quercetin Lupeol β-sitosterol	(215)
					Aq. EtOH	Soxhlet	TLC	Gallic acid Quercetin	(53)
<i>C. chinensis</i>	WP	EtOH	-----	VLC	Odoroside H 21-hydroxyodoroside H Neritaloside Strosposide 16-_-hydroxydigitoxin N-trans and cis feruloyl tyramines Ethyl caffeate Coumarins Ursolic acid_-sitosterol Glucoside 4-O-p-coumaroyl_-D-glucoside	(216)			
					-----	-----	GC-MS	Heneicosanoic acid Pentadecanoic acid	(217)
					n-hex	Soxhlet	-----	-----	-----

Continued Table 3.

Stem	EA	Maceration	GC-MS			
				Hexadecenoic acid Heptadecanoic acid Octadecanoic acid 1, 2, 3 Propanetriol, 1- acetate, Benzofuran 2, 3, dihydro Glycerol 1, 2- diacetate 1 H- 1, 2, 4-triazol-5-amine 1- ethyl- 2-methoxy-4-vinylphenol Triacetin D - glucitol, 4 - O-hexyl 3,4,5-trimethoxy cinnamic acid Hexadecanoic acid, ethyl ester 3,6 -di methoxy phenanthrene 3, 5 - di - tert -Butyl -4 -hydroxyanisol Vanillin 3 - aminopyrrolidine Cetene Sarcosine, N -isobutyryl, tetradecyl ester 4 - ((1E) - 3 - hydroxyl -1-propenyl)-2- methoxy phenol 1,5-diphenyl-2H-1,2,4-triazoline-3-thione 1-octadecene Heptanamide, N-(1-cyclohexylethyl)-2- methyl Scoparone Hexadecanoic acid, ethyl ester 3'-Methyl-2-benzylidene-coumaran-3-one 5-hydroxy-7-methoxy-6-(2,3-epoxy-3- methylbutyl)-flavanone (reflexin) Isorhamnetin Isorhamnetin-3-O-glucoside Isorhamnetin-3-O-robinobioside 2-Methoxy-4-vinyl phenol Benzofuran-2,3-dihydro 3,5-di-tert-Butyl-4-hydroxyanisole Hexatriacontane n-Hexadecanoic acid Scoparone Hexadecanoic acid methyl ester 1,3-Benzenediamine, N, N, N', N' tetramethyl- Phenol, 4(3-hydroxypropenyl), 2-methoxy Phenol, 2,4 bis (1,1dimethylethyl); 2,3,5,6- Tetramethyl para phenylene diamine Retinoic acid-5,6-epoxy-5,6-dihydro 2,4-Dihydroxy- 2,5-dimethyl-3(2H) furan-3-one 2,3-dihydro-3,5-dihydroxy-6-methyl-2- Propyl-tetrahydro-pyran-3-ol Pregn-4-ene-18-oic acid Swarnalin Cis-swarnelin Coumarin 5, 6, 7-trimethoxycoumarin Aromadendrin Taxifolin Aromadendrin-7-O-β-D-glucopyranoside 3,5,7,8,4'-pentahydroxyflavanone Taxifolin-7-O-β-D-glucopyranoside Coccinoside B Pruning 3-O-dicaffeoyl quinic acid 3-4-O-dicaffeoyl quinic acid 3, 4, 5-O-Tricaffeoylquinic acid	(218)	
	Pet Eth	Soxhlet	CC	5-hydroxy-7-methoxy-6-(2,3-epoxy-3- methylbutyl)-flavanone (reflexin) Isorhamnetin Isorhamnetin-3-O-glucoside Isorhamnetin-3-O-robinobioside	(206) (122)	
	MeOH	Maceration	GC-MS	2-Methoxy-4-vinyl phenol Benzofuran-2,3-dihydro 3,5-di-tert-Butyl-4-hydroxyanisole Hexatriacontane n-Hexadecanoic acid Scoparone Hexadecanoic acid methyl ester 1,3-Benzenediamine, N, N, N', N' tetramethyl- Phenol, 4(3-hydroxypropenyl), 2-methoxy Phenol, 2,4 bis (1,1dimethylethyl); 2,3,5,6- Tetramethyl para phenylene diamine Retinoic acid-5,6-epoxy-5,6-dihydro 2,4-Dihydroxy- 2,5-dimethyl-3(2H) furan-3-one 2,3-dihydro-3,5-dihydroxy-6-methyl-2- Propyl-tetrahydro-pyran-3-ol Pregn-4-ene-18-oic acid	(219)	
AP	MeOH	Maceration	RHPLC HPLC	Swarnalin Cis-swarnelin Coumarin 5, 6, 7-trimethoxycoumarin	(205)	
-----	Water	-----	-----	Aromadendrin Taxifolin Aromadendrin-7-O-β-D-glucopyranoside 3,5,7,8,4'-pentahydroxyflavanone Taxifolin-7-O-β-D-glucopyranoside Coccinoside B Pruning 3-O-dicaffeoyl quinic acid 3-4-O-dicaffeoyl quinic acid 3, 4, 5-O-Tricaffeoylquinic acid	(49)	
-----	DCM	Maceration	HPLC	Violaxanthin Lutein Lycopene β, ψ-carotene Rubixanthin	(220)	

Continued Table 3.

<i>C. chinensis</i>	Fil.	Water	Maceration	CC	β, β - carotene Esterified rubixanthin Lutein violaxanthin β-cryptoxanthin		
	Fruit	50 % MeOH	-----	CC	An antiviral protein with molecular weight about 14,000---18,000 daltons	(219)	
					Cuscutamine	(194)	
	Stem	Pet. eth CF	-----	-----	Cuscutoside A (2'-hydroxyl asarinin 2'-O-β-D-apiofuranosyl-(1 → 2)-β-D-glucopyranoside Cuscutoside B (2'-hydroxyl asarinin 2'-O-β-xylopyranosyl-(1 → 6)-β-glucopyranoside Hyperoside Astragalol Quercetin Quercetin-3-O-apiosyl (1-2)-galactoside Pinoresinol-4-O-glucoside Einoresinol Epipinoresinol p-coumaric acid Caffeic acid Chlorogenic acid Arbutin		
					β-sitosterol	(221)	
	Seed	Pet. eth	Reflux	CC	9(R) - hydroxy-d-sesamin D-pinoresinol daucosterol		
					Cuscutoside C (2'-hydroxyl asarinin 2'-O-β-D-glucopyranoside) Cuscutoside D (2'-hydroxyl asarinin 2'-O-β-D-apiofuranosyl-(1 → 2)-[β-D-glucopyranosyl-(1 → 6)]-β-D-glucopyranoside	(196)	
						Neo-sesamin	(210)
						Kaempferol Kaempferol-3-O-β-D-glucopyranoside 4', 4', 6-trihydroxyaurone Quercetin Hyperoside Palmitic acid Stearic acid β-sitosterol Daucosterol	
					RHPLC	quercetin 3-O-β-D-galactoside-7-O-β-D-glucoside quercetin 3-O-β-D-apiofuranosyl-(1etin3D-galactoside hyperoside quercetin kaempferol	(218)
	Ether Water	Saponi- fication		CC	A trisaccharide	(194)	
					Four new glycosidic acids (cuscutic acids A-D) Acetic acid Propionic acid 2-methylbutyric acid Tiglic acid Nilic acid Convolvulinolic acid Jalapinolic acid		
	95 % EtOH			CC	Cuscuta-resinols A-C (+)-sesamin (+)-xanthoxylol 9-hydroxysesamin (+)-pinoresinol Kaempferol	(212)	



Continued Table 3.

		95 % EtOH	-----	CC	kaempferol Isorhamnetin Kaempferol Quercetin astragalin, isorhamnetin hyperoside	(22)
		n-hex	-----	Capillary GC	Sixteen fatty acids including Palmitic acid Linoleic acid Oleic acid Linolenic acid	(222)
		MeOH	-----	-----	Methyl 4-hydroxy-3,5-dimethoxycinnamate, Caffeic acid Quercetin Kaempferol Calycopteretin	(223)
		EtOH	-----	CC	neocucucosides A, B and C	(224)
		-----	-----	-----	Octadecyl (E)-p-coumarate Methyl 3-O-β-D-glucopyranosyl-5- hydroxycinnamate Quercetin-3-O-(6"-galloyl) β-D-glucoside Kaempferol Astragalin Hyperoside Astragalin 6"-O-gallate β-sitosterol Daucosterol	(225)
<i>C. japonica</i>	Seed	MeOH	-----	FCC	3, 5-Di-O-caffeoylquinic acid 3, 4-Di-O-caffeoylquinic acid Methyl 3, 5-Di-O-caffeoylquinic acid Methyl 3, 4-Di-O-caffeoylquinic acid	(226)
<i>C. australis</i>	Stem	80 % acetone	-----	CC	α-caroten-5 6-epoxide β-and γ-carotene Xanthophylls Taraxanthin Lutein Kaempferol	(227)
	Seed	-----	-----	GC	Cuscutic acids A <sub>1</sub> -A <sub>3</sub> Acetic acid Isobutyric acid 2-methylbutyric acid Tiglic acid Nitic (3-hydroxy-2-methylbutyric) acid	(217)
		-----	-----	-----	β-sitosterol Sesamin Hexadecanoic acid Hexadecanoic acid Kaempferol Quercetin Astragloside Hyperoside caffeic acid Quercetin-3-O-β-D-galactopyranosyl-β-D- apiopyranoside	(228-229)
<i>C. europaea</i>	-----	-----	-----	-----	Glycoside Flavonoids	(166)
<i>C. campestris</i>	AP	MeOH	Maceration	HPLC	Sinapic acid Quercetin Hesperidin Eugenol	(14)
<i>C. racemosa</i>	WP	70 % EtOH	Percolation	TLC	Flavonoids Tannins Flavonol (4' methoxyquercetin)	(214)

Continued Table 3.

		MeOH	Soaked	DCCC	Kaempferol Quercetin Pinoresinol 9- $\alpha$ -hydroxyxsesamin 9- $\beta$ -hydroxyxsesamin Acuminatolide	(230)
		-----	-----	-----	Quercetin 5, 7, 3', 4'-tetramethyl ether	(231)
<i>C. pedicellata</i>	WP	EtOH	-----	CC	Naringenin Kaempferol Aromadenderin Quercetin 3,5,7,30,50-pentahydroxy flavanone, Naringenin -7-O-b-D-glucoside Aromadenderin -7-O-b-D-glucoside, Taxifolin -7-O-b-D-glucoside, Kaempferol -3-O-b-D-glucoside Quercetin -3-O-b-D-glucoside	(26)
	Seed	Pet. eth	Soxhlet	CC	Quercetin Kaempferol Genkwanin Astragaln Palmitic acid	(232)
<i>C. epithymum</i>	WP	MeOH	Soxhlet	-----	Alkaloids Carbohydrates Flavonoids Glycosides Phytosterols Triterpenoids	(182)
<i>C. approximata</i>	AP	MeOH	Maceration	HPLC	Gallic acid Catechin Caffeic acid Chloregenic acid, Quercetin Coumarin, Vanilin, Eugenol	(14)
<i>C. monogyna</i>	AP	MeOH	Maceration	HPLC	Sinapic acid Catechin Caffeic acid Chloregenic acid Rutin Coumarin Vanilin Hesperidin Ellagic acid	(14)
<i>C. mitraeformis</i>	Stem	n-hex	-----	GC-FID GC-MS HPLC-DAD	Nonanal Thymol Eugenol $\beta$ - carotene Lutein	(78)
<i>C. kotschyana</i>	-----	-----	-----	-----	Quercetin kaempferol	(233)

*C. Cuscuta*; WP: whole plant; AP: aerial parts; Fil: filament; Aq: aqueous; MeOH: methanol; EtOH: ethanol; Pet. eth: petroleum ether; n-hex: n-hexan; EA: ethyl acetate; DMC: dichloromethane; CC: column chromatography; HPLC: high performance liquid chromatography; RHPLC: reverse phase high performance liquid chromatography; TLC: thin layer chromatography; VLC: vacuum liquid chromatography; GC-MS: gas chromatography-mass spectrometry; FCC: Flash Column Chromatography; DCCC: Droplet counter-current chromatography; FID: flame ionization detector; DAD: diode array detector

### Pharmacological attributes

Impressive medicinal background of *Cuscuta* species has attracted the attention of many pharmacological researchers. A good deal of biological attributes has been studied and is listed in tabular form in Table 4.

### Antioxidant

Medicinally important plants are endless reservoirs of antioxidants that enhance the antioxidant capacity of

the body, which lead to a reduced risk of many diseases (234-235). Although a diverse population of synthetic analogs is commercially available due to side effects (liver impairment and carcinogenesis) blind reliance on these formulations has been over. Therefore, plants can play a key role to fulfill prerequisite for exploration of effective, biocompatible, and economic antioxidants (236).

Many investigators have employed different

qualitative and quantitative approaches to detect antioxidants in various *Cuscuta* species. Stem collected from different hosts and extracted with various solvents (100% methanol, 80% methanol, 100% ethanol, 80% ethanol, water, and n-hexane) were analyzed for quantity of phenolics and flavonoids content. Their antioxidant capacity was measured by using a variety of assays including reducing power, DPPH scavenging activity, percent inhibition of linoleic acid peroxidation and  $\delta$ -tocopherol. It was observed that there was a strong correlation between amount of total phenolics and antioxidant capacity (13).

*C. reflexa* has been reported for its antioxidant

potential (37, 237). Free radical scavenging capacity of methanolic extract of *C. reflexa* was evaluated by DPPH and reducing power assays. Results of DPPH assay, illustrated as IC50 value demonstrated its antioxidant activity 359.48  $\mu\text{g/ml}$  as compared to 9.22  $\mu\text{g/ml}$  value for ascorbic acid used as standard. The reducing power of extract was found dose-dependent and increased by increasing concentration (35). Ethyl acetate fraction of ethanolic extract of *C. reflexa* was significantly antioxidant. Activity may be related to presence of flavonoids, alpha tocopherol, and rutin, which were confirmed in preliminary phytochemical screening (238).

**Table 4.** Pharmacological attributes exhibited by *Cuscuta* species

Species	Activity	Plant part	Method	Extract type	Test applied	Testing model	Effective dose/conc.	Reference			
<i>C. reflexa</i>	Antioxidant	St	Soxhlet	MeOH	DPPH and FRAP assay	-----	600 $\mu\text{g/ml}$	(35)			
		L		EtOH	Non-Enzymatic Glycolysation of Haemoglobin	Hemoglobin	-----	(238)			
	Antibacterial	Fl	-----		MeOH	DPPH assay	-----	-----	(323)		
		L	Soxhlet		50% EtOH	Disc diffusion method	<i>Escherichia coli</i>	-----	(29)		
		St	-----		MeOH	Cup plate method	<i>Staphylococcus aureus</i> <i>Escherichia coli</i> <i>Bacillus punilus</i> <i>Salmonella typhi</i> <i>Salmonella thyphimurium</i> <i>Salmonella boydii</i> <i>Salmonella sonnei</i> <i>Salmonella dysenteriae</i> <i>Pseudomonas aeruginosa</i> <i>Klebsiella pneumoniae</i> <i>Vibrio cholerae</i>	125 $\mu\text{g/ml}$	(259)		
		WP	-----		DCM pet. eth	Disc diffusion method	<i>Bacillus subtilis</i> <i>Staphylococcus lutea</i> <i>Xanthomonas campestris</i> <i>Escherichia coli</i> <i>Klebsiella pneumoniae</i> <i>Proteus vulgaris</i> <i>Proteus denitrificans</i>	16 to 512 $\mu\text{g/ml}$	(235)		
				Soaked		EtOH	Agar well diffusion assay	<i>Bacillus subtilis</i> <i>Staphylococcus aureus</i> <i>Escherichia coli</i> <i>Salmonella typhi</i>	500 $\mu\text{g/ml}$	(324)	
				-----	-----		MeOH	Agar well diffusion	<i>Staphylococcus epidermidis</i> , <i>Staphylococcus aureus</i> <i>Escherichia coli</i> <i>Pseudomonas sp.</i> <i>Klebsiella pneumoniae</i>	-----	(260)
		Antifungal	L	soxhlet		50% EtOH	-----	<i>Aspergillus niger</i> <i>Candida albicans</i>	-----	(29)	
				-----	-----		Water	well diffusion method	<i>Aspergillus alternate</i> <i>Aspergillus niger</i> <i>Fusarium solani</i> <i>Fusarium oxysporium</i> <i>Macrophomina phaseolina</i>	30% (w/v)	(107)
		Antihypertensive	WP	Socked		EtOH	-----	Wistar rats	0.1 ml bolus injection	(283)	
		Psychopharmacological effect	St	Soxhlet		Pet. eth	General and exploratory behavior study	Swiss albino mice	-----	(39)	
		Anti-inflammatory	St	Suc. Ex		MeOH Pet. eth	Membrane stabilizing activity	Red blood cells	-----	(44)	
					Soxhlet	EtOH water	Percentage volume reduction	Albino rats	200, 400 mg/kg	(254)	

Continued Table 4.

	WP	Decoc.	Water	SQ-RT-PCR analysis	Murine macrophage cell line RAW264.7	-----	(253)	
Diuretic activity	AP	-----	EtOH water	Urine volume and electrolyte content	Wister rats	300 mg/kg	(45)	
Hepatoprotective	WP	Suc. Ex	Aq.	Biochemical parameters	Albino rats	200 mg/kg	(242)	
	AP	Soxhlet	Methanol	Biochemical parameters	Albino rats	-----	(32)	
Antitumor/anticancer	WP	Suc. Ex	MeOH CF	-----	Swiss albino mice MCF-7 cancer cell line	40 mg/kg	(15)	
		Decoc.	Water	MTT assay DAPI staining Annexin V staining SQ-RT-PCR analysis	Human hepatocellular carcinoma cell line Hep3B	-----	(253)	
	-----	-----	CF	Annexin V-FITC Apoptotic assay PARP cleavage Caspase activation	Hep 3B cell line	-----	(325)	
Antisteroidogenic	St	Soxhlet	MeOH	Ovary and uterus weight Biochemical parameters	Swiss albino mice	-----	(42)	
Hair growth	St	Soxhlet	Pet. eth	Visual observation Skin biopsy Histopathological observation	----- Swiss albino rats	2% extract in vehicle 250 mg/kg	(40) (313)	
Antidiabetic	St	Macera.	MeOH CF	Oral glucose tolerance test	Long Evans rats and Swiss albino mice	50-200 mg/kg bw	(50)	
	AP	Macera.	MeOH water	Oral glucose tolerance test	Swiss albino rats	400 mg/kg	(245)	
Antimutagenic	St	Soxhlet	MeOH	Ames test	<i>Salmonella typhimurium</i> TA 98 and TA 100	-----	(122)	
Anthelmintic	WP	-----	Pet. eth CF MeOH	-----	Pheritima posthuma	20-50 mg/ml	(44)	
Anxiolytic effect	WP	Macera.	MeOH	Elevated plus-maze Light and dark chamber	<i>Swiss albino mice</i>	400 mg/kg	(305)	
Anti-arthritis	St	-----	70% MeOH	Percentage inhibition of oedema Percentage inhibition of protein denaturation	Sprague-Dawley rats	600 mg/kg	(321)	
Nephroprotective	St	-----	70% MeOH	Biochemical parameters and pathological symptoms	Sprague-Dawley rats	600 mg/kg	(321)	
Anticonvulsant	L	Macera.	EOH	Delay the onset of convulsions	Albino mice	200 and 400 mg/kg	(238)	
Genotoxic effects	-----	-----	MeOH	Root growth, root apical meristem mitotic index (MI), chromosomal aberrations	<i>Allium cepa</i> L. <i>Allium sativum</i> L.	-----	(326)	
<i>C. chinensis</i>	anti-histaminic	-----	-----	EtOH	Albino rats	100 mg/kg	(327)	
	Anticancer	WP	Soaked	Water	Histological study	Swiss albino mice	1 g/kg	(30)
	Neuronal differentiation	Sd	Percola.	MeOH	Neurite assay	Rat pheochromocytoma PC12 cells	200 mg/l	(277)
	Adjuvant effect	Sd	-----	70% EtOH	Splenocyte proliferation assay Indirect ELISA	ICR mice	200 µg	(272)
	Hepatoprotective	Sd	Decoc.	EtOH	Liver function markers and histopathological study	Wistar-albino rats	125 and 250 mg/kg	(33)
	Antioxidant	Sd	Decoc.	EtOH	Antioxidant enzyme levels	Wistar-albino rats	125 and 250 mg/kg	(33)
	Antiosteoporotic	Sd	-----	95% EtOH	Alkaline phosphatases activity Alamar-Blue cell proliferation assay Reporter assays	UMR-106 cells	-----	(22)
	Improve erectile dysfunction	Sd	-----	-----	Radioimmunoassay	New Zealand white rabbits	1-5 mg/ml	(288)
	Anti-inflammatory	Sd	-----	80% EtOH	Griess assay ELIZA	Mouse microglia line BV-2 cells	-----	(255)
	Anti-apoptosis	Sd	-----	95% EtOH	Annexin V-FITC method	SD rats	-----	(303)
Effect on Melanogenesis	Sd	Hot Ex	EtOH water	Melanin contents and tyrosinase activity	B16F10 mouse melanoma cells Zebrafish	-----	(160)	

Continued Table 4.

	Anti-arthritic	St	-----	70% MeOH	Light and dark chamber Percentage inhibition of oedema Percentage inhibition of protein denaturation	Sprague-Dawley rats	600 mg/kg	(321)
	Nephroprotective	St	-----	70% MeOH	Biochemical parameters and pathological syptoms	Sprague-Dawley rats	600 mg/kg	(321)
	Anticonvulsant	L	Macera.	EOH	Delay the onset of convulsions	Albino mice	200 and 400 mg/kg	(238)
	Genotoxic effects	-----	-----	MeOH	Root growth, root apical meristem mitotic index (MI), chromosomal aberrations	<i>Allium cepa</i> L. <i>Allium sativum</i> L.	-----	(326)
<i>C. chinensis</i>	anti-histaminic	-----	-----	EtOH		Albino rats	100 mg/kg	(327)
	Anticancer	WP	Soaked	Water	Histological study	Swiss albino mice	1 g/kg	(30)
	Neuronal differentiation	Sd	Percola.	MeOH	Neurite assay	Rat pheochromocytoma PC12 cells	200 mg/l	(277)
	Adjuvant effect	Sd	-----	70% EtOH	Splenocyte proliferation assay Indirect ELISA	ICR mice	200 µg	(272)
	Hepatoprotective	Sd	Decoc.	EtOH	Liver function markers and histopathological study	Wistar-albino rats	125 and 250 mg/kg	(33)
	Antioxidant	Sd	Decoc.	EtOH	Antioxidant enzyme levels	Wistar-albino rats	125 and 250 mg/kg	(33)
	Antiosteoporotic	Sd	-----	95% EtOH	Alkaline phosphatases activity Alamar-Blue cell proliferation assay Reporter assays	UMR-106 cells	-----	(22)
	Improve erectile dysfunction	Sd	-----	-----	Radioimmunoassay	New Zealand white rabbits	1-5 mg/ml	(288)
	Anti- inflammatory	Sd	-----	80% EtOH	Griess assay ELIZA	Mouse microglia line BV-2 cells	-----	(255)
	Anti-apoptosis	Sd	-----	95% EtOH	Annexin V-FITC method	SD rats	-----	(303)
Effect on Melanogenesis	Sd	Hot Ex	EtOH water	Melanin contents and tyrosinase activity	B16F10 mouse melanoma cells Zebrafish	-----	(160)	
	Cytotoxic	WP	-----	-----	Methyl tetrazolium bromide test	Human Acute Lymphoblastic Leukemia Cell Line	3 µg/ml in 24 hr	(267)
<i>C. japonica</i>	Antihypertensive	Sd	-----	EA MtOH	Plasma ACE activity	Rats	400 mg/ml	(226)
	Melanogenesis inhibition	Sd	Hot Ex	Water	Tyrosinase activity assay melanin contents cAMP assay Western blot analysis	B16F10 mouse melanoma cells (CRL 6323)	-----	(69)
	Memory enhancing	Sd	Sonicat.	Water	Novel object recognition test The step-through passive avoidance test Immunohistochemistry	ICR mice	50 and 100 mg/kg/day	(315)
<i>C. australis</i>	Melasma elimination	AP	Heating	Water	Melasma Area Severity Index degree of hyperpigmentation	Patients	4.8 g/day	(311)
	Hepatoprotective	St	Soxhlet	EtOH	Hepatic injury markers	Wistar rats	125 and 250 mg/kg	(70)
<i>C. europaea</i>	Antibacterial	WP	Shaking	EtOH	Agar well method	<i>S. aureus</i> <i>E. coli</i>	20 mg/ml	(263)
<i>C. planiflora</i>	Antidepressant	AP	-----	-----	Triple-blind controlled clinical trial	Depression patients	500 mg capsule	(308)
<i>C. campestris</i>	Analgesic	WP	-----	95% EtOH	Writhing Test	Albino mice	50 and 100 mg/kg.	(47)
				Cold Macera.	MeOH	Writhing Test	Swiss Albino mice	400 mg/kg
	Antipyretic	WP	-----	95% EtOH	Heat conduction method electric thermocouple	Albino mice	50 and 100 mg/kg	(47)
	Antiinflammatory	WP	-----	95% EtOH	Volume plethysmographically	Albino mice	100 mg/kg	(47)
	CNS-depressant	WP	-----	95% EtOH	Behavioural study	Albino mice	50 and 100 mg/kg	(47)
	Anticancer	WP	-----	EA MeOH	-----	Hepatocellularcarcinoma cell line	-----	(270)
		AP	Macera.	MeOH	RT PCR analysis	MCF 10A, MCF-7 and MDA- MB-231 cell lines	-----	(271)

Continued Table 4.

	Antiviral	AP	Shaking	MeOH	RT-PCR analysis	Peripheral blood mononuclear cell	1000 mg/kg	(264)
	Hepatoprotective	WP	-----	75% EtOH	Biochemical parameters and histological	Mice	20, 100 and 500 mg/kg	(104)
<i>C. racemosa</i>	Antimicrobial	WP	Percola.	70% EtOH	Dilution in a liquid medium	<i>Staphylococcus aureus</i>	2 mg/ml	(214)
<i>C. pedicellata</i>	Anti-obesity	WP		EtOH	Biochemical measurements	Albino rats	400 mg/kg	(26)
	Antioxidant	Sd	Soxhlet Macera.	MeOH	DPPH assay	-----	-----	(232)
	Antibacterial	L	Decoc. infusion	Water	-----	<i>Xanthomonas campestris</i>	-----	(74)
		L	-----	MeOH	Agar well diffusion method	<i>Staphylococcus aureus</i> <i>Pseudomonas aeruginosa</i> <i>Klebsiella pneumonia</i> <i>Acinetobacter baumannii</i>	100 µl	(168)
	Antifungal	L	-----	MeOH	Agar tube dilution method	<i>Aspergillus fumigatus</i> , <i>Aspergillus flavus</i> <i>Rhizopus oryzae</i>	67 µl	(168)
	Cytotoxic potential	L	-----	MeOH	Brine shrimp assay	-----	-----	(168)
		St						
		Fr						
	anti-inflammatory	L	-----	MeOH	Albumin denaturation, membrane stabilization proteinase inhibitory assays	-----	200 µg/ml	(168)
		St						
		Fr						
<i>C. epithymum</i>	Hepatoprotective	WP	Soxhlet	MeOH	Blood serum parameters	Wistar albino rats	400 mg/kg BW	(182)
<i>C. kotschyana</i>	Anticancer	Sd	Decoc.	2N HCl EA	MTT assay	MCF-7 cell line	100 µg/ml	(77)
		St			Annexin V			
<i>C. mitraeformis</i>	Antioxidant	St	Hydro. dil	n-hex	DPPH assay	-----	-----	(78)
	Antimicro-bial	St	Hydro. dil	n-hex acetone	Broth microdilution method	<i>Clavibacter michiganensis</i> <i>Erwinia carotovora</i> <i>Pseudomonas syringae</i>	-----	(78)
<i>C. arvensis</i>	Analgesic activity	WP	-----	n-hex, DCM, EA, MeOH, water	Writhing test	Swiss albino mice	100 mg/kg	(257)

WP whole plant; AP aerial parts; Fl flower; St stem; Sd seed; Fr fruit; L leaves; Aq. aqueous; MeOH methanol; EtOH ethanol; Pet. eth petroleum ether; n-hex n-hexan; EA ethyl acetate; DMC dichloromethane; CF chloroform; Macera maceration; Decoc decoction; Hydro. Dil hydro distillation; Percola percolation; Sonicat sonication; Hot Ex hot extraction; Suc. Ex successive extraction

Seed oil of *C. pedicellata* was extracted with petroleum ether (pet. ether) and lipid contents were saponified to separate unsaponifiable materials and fatty acids. The extract was fractionated by using various solvents, and antioxidant activity of all extracts (pet. ether, unsaponified, fatty acids, 70 % methanol, ethyl acetate, and chloroform) was appraised by DPPH free radical assay. The methanol extract was found most potent (230).

In another study, a correlation was established between antioxidant activity and total phenolic content of aerial parts of three Iranian *Cuscuta* species. *C. approximata*, *C. monogyna* and *C. campestris* were estimated by using DPPH microplate method. The highest concentration of phenolic compounds was found in *C. monogyna* and *C. approximata*. TPC of

plant methanolic extracts was determined. Methanolic extracts of *C. approximata* and *C. monogyna* contain highest amounts of total phenolic, 56.67 mg/g and 49.59 mg/g, respectively, while antioxidant potential was in the order *C. monogyna* > *C. approximata* > *C. campestris* (14).

Ethyl acetate fraction of ethanolic extract of *C. chinensis* seeds possesses strongest antioxidant effect with kaempferol and quercetin as its main constituents. It hunts free radicals and inhibits lipid peroxidation (198, 239). The same fraction of methanolic extract was ascertained as an effective antioxidant by DPPH free radical scavenging assay (222). Moreover, aqueous extract of *C. chinensis* can protect murine osteoblastic MC3T3-E1 cells against tertiary butyl hydroperoxide induced injury because of its oxidation

stress management potential and functioning against mitochondria-dependent pathways (240). In another experiment, flavonoids of *C. chinensis* were evaluated for their protective effect against oxidative stress. The survival rate of PC12 cells having H<sub>2</sub>O<sub>2</sub>-induced apoptosis was measured. The protective effect was possibly due to scavenging of reactive oxidative species and enhanced activity of antioxidant enzyme (241). Essential oils and carotenoids separated from *C. mitraeformis* also showed antioxidant activity (78). These results suggest that *Cuscuta* plants are enriched with highly important natural antioxidants that may be used in development of functional foods and drugs effective against diseases caused by oxidative stress. Isolation, identification and possible synergism among various components may be the subject of interest for further studies.

### Hepatoprotective

Anti-hepatotoxic drug designing is a major thrust area seeking the attention of natural product researchers because synthetic formulations have serious side effects. *C. epithymum* is traditionally used as a liver tonic. *C. epithymum* whole plant extracted in methanol exhibited appreciably high hepatoprotective effect against CCl<sub>4</sub> induced hepatotoxicity in albino rats. Elevated serum aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total bilirubin have confirmed hepatic damage after CCl<sub>4</sub> administration. *C. epithymum* prevented the toxic effect in both anticipatory and curative models, which may be due to the presence of various bioactive moieties, including phenolics, flavonoids, and alkaloids (185).

Many investigators have studied the curative effect of *C. reflexa* against liver damage induced by cisplatin, paracetamol, carbon tetrachloride, ethanol, isoniazid, and rifampicin. Various biochemical measurements were observed including ALT, AST, ALP, and total bilirubin before and after the administration of *C. reflexa* extract. It improved liver function by significantly reducing the serum ALT, AST, and ALP levels in affected rats comparable to standard. Histopathological examination of liver section supports the results (32, 242, 243).

Ethanol extract of *C. australis* also appeared as liver protector against acetaminophen intoxication in an animal model. Two groups of rats were intoxicated on day eight after receiving doses of *C. australis* seed and stem extract separately for seven days. In untreated rats, severe periportal hepatic necrosis, considerably raised serum liver damage markers, noticeably augmented lipid peroxidation and suppressed liver antioxidant enzymes activities were witnessed. Comparative evaluation of seed and stem extract proves that stem is a more potent hepatoprotective counterpart than seed (70).

Seeds of *C. chinensis* are commonly employed to nourish and improve hepatic disorders in China and various other Asian countries. Oxidative stress can stimulate the development of acetaminophen-induced hepatotoxicity. Liver protecting and antioxidant activities of ethanolic and aqueous extracts of *C. chinensis* on acetaminophen-induced hepatotoxicity in rats. Ethanolic extract showed a significant hepatoprotective effect at an oral dose of 125 and 250 mg/kg confirmed by the measurement of various

parameters and observation of liver histopathology. Comparatively same doses of the aqueous extract were found ineffective rather; it resulted in further hepatic deterioration (33). *C. chinensis* nanoparticles were found more effective in this regard (198, 239). Thus, from the above findings it can be observed that many *Cuscuta* species are promising hepatoprotective agents supporting the claims of traditional healers. Further investigations on chemical components are needed to pinpoint the findings.

### Antidiabetic

Diabetes mellitus is becoming a growing threat for a vast population in almost all countries of the world due to a sluggish lifestyle leading to reduced physical activity and increase in obesity (244). Methanolic and chloroform extracts of *C. reflexa* whole plant exhibited significant hypoglycemic activity at doses of 50, 100, and 200 mg/kg body weight. Oral glucose tolerance test was used to estimate the effect in glucose-loaded Long Evans rats (50). Administration of methanolic extract *C. reflexa* to glucose-loaded mice led to notable reductions in blood glucose and improved metabolic alterations, thereby justifying its traditional folkloric claims (89, 245).

Antidiabetic activity of *C. chinensis* was evaluated in dexamethasone-induced insulin-resistant human liver carcinoma (HepG2) cells (246). *C. chinensis* polysaccharides can reduce blood sugar level in type-2 diabetes. Efficacy was tested on alloxan-induced diabetes in a mice model. Orally administered doses of 300 and 600 mg/kg remarkably decreased the elevated fasting blood glucose (247-248). In a similar study, oral administration of 200 and 400 mg/kg polysaccharides significantly lessened blood glucose along with glycosylate serum protein (249). A Chinese herbal prescription, Zhujing pill, having more than 50 % *C. chinensis* protected retina of diabetic rats, possibly through its antioxidation and anti-inflammatory effects (250). Recently mechanism of hypoglycemic activity of *C. chinensis* on type 1 diabetic disease was investigated using a rat model. Daily administration of *C. chinensis* extract returned fasting serum insulin and fasting blood glucose to normal value by upregulating the gene expression of hepatic and pancreas genes (251). It is crucial to continue the exploration of hypoglycaemic effect of more plants as these are blessed with similar chemical profile.

### Anti-inflammatory

Inflammatory reactions play a decisive role in different phases of pathogenesis of cancer. So, there may be an assumption that anti-inflammatory drugs can induce apoptosis in cancerous cells and may be equally beneficial as preventive measure and therapy (252). Aqueous and alcoholic extracts of stem of *C. reflexa* and its ethyl acetate fraction showed remarkable anti-inflammatory activity in *in vitro* and *in vivo* tests. Inflammation was induced by various chemicals like histamine and lipopolysaccharide. It was observed that extracts inhibited inflammatory responses that can be related to the presence of flavonoids, phenols, and polyphenols in this plant (43-44, 253). *C. reflexa* significantly suppressed inflammation by reducing

edema volume up to 80 % in rats as compared to standard 96.36 % (254).

*C. campestris* markedly inhibited carrageenan-induced edema in rats by oral pretreatment with 100 mg/kg extract (47). *C. chinensis*, by suppressing the inflammatory responses showed the potential for treatment of brain inflammation (255). Moreover,  $\lambda$ -carrageenan-induced paw edema treatment by using the methanolic extract of *C. chinensis* seed in mice, also confirmed its anti-inflammatory effect (256). *C. pedicellata* and *C. arvensis* were found effective against inflammation (168, 257). Further studies must be conducted to clarify the mechanism and to figure out the active principle behind the activity.

### Antibacterial, antifungal, and antiviral

Continuous and urgent exploration is required for new antimicrobial agents with new compositions and diverse mechanisms of action to overcome antimicrobial modifications (9). Methanolic extract of *C. reflexa* was found significantly active against a broad spectrum of bacterial species including *S. aureus*, *P. aeruginosa*, *S. dysenteriae*, *S. boydii*, and *E. coli* with impressive zone of inhibition (27, 258-260).

*Xanthomonas campestris* (XC) is a widely spread infectious agent causing a huge loss in food crops with visible symptoms and leave shedding. Aqueous decoction and infusion extract of *C. pedicellata* were evaluated for antibacterial activity against diverse pathogens of XC using *in vitro* well diffusion method. Inhibition zone diameter was observed from 1.0 to 5.0 cm (74). The methanolic extract also showed promising high antimicrobial activity (168). *C. australis* is another species having notable antibacterial effect. The 50 % methanolic extract was fractionated by hexane, ethyl acetate, and butanol with various polarities. All fractions were tested against fungal, yeast and various Gram-positive and Gram-negative bacteria. All extracts except n-hexane were found effective against different species (261). Additionally, methanolic extract of *C. epithymum* was also significantly active against *Bordetella bronchiseptica* demonstrating zone of inhibition from 10–14 mm (262). *C. europaea* was active against *Staphylococcus aureus* even higher than standard drug Amoxicillin. These results lead toward the concept that this plant can be used as a safer option against this microbe (263). Recently essential oils and carotenoids separated from *C. mitraeformis* were found antibacterial (78).

In addition to many other species of genus *Cuscuta*, *C. racemosa* offers flavonoids as chief metabolites. Slightly positive antimicrobial activity of this plant was observed against *S. aureus* using dilution in a liquid medium method. Minimum inhibiting concentration was 2.0 mg/ml. Phenolic compounds are documented as antimicrobial substances. So, the activity can be ascribed to the flavonoids and tannins in the plant (52).

Several secondary metabolites like flavans, flavones, and quinic acid derivatives have been found active against HIV infection. Crude aqueous extracts of *C. reflexa* exhibited anti-HIV activity. Virus inhibition may be attributed to the combinatory effects of nine closely related compounds (49). An antiviral protein with significantly high inhibiting property was isolated

from the aqueous extract of *C. reflexa* (219). Methanolic extract of *C. campestris* showed weak anti-HIV activity (264). A number of species have been found effective against microbes. It is recommended that further studies with isolated components instead of extracts may be more useful to identify the active compounds.

### Antitumor effect

Some species of the genus *Cuscuta* afford alkaloids with indolic nuclei that are considered potential antitumor substances. *C. chinensis* is a popular antitumor prescription in the Unani medicine system. Oral administration of the plant extract at a dose of 1 g/kg noticeably delayed the appearance and growth of skin papilloma and reduced the chances of carcinoma (30). Anticancer activity of *C. chinensis* has been evaluated by several pharmacological studies using a variety of cell lines. Results prove that it can act as an integrative approach to encounter ever-growing disease management (22, 31, 265-267).

*In vivo* anticancer potential of *C. reflexa* was determined by using murine models. Alcoholic extract and its chloroform fraction were found more potent. It showed highest toxicity against human breast cancer cell lines. Similarly, chloroform part of extract of alcohol showed considerable tumor growth inhibition, which reveals that these extracts interfere in cell proliferation to inhibit cancer (15). It can induce apoptosis in Hep3B cells (253). Phenolic components isolated from *C. reflexa* were also assessed in HCT116 colorectal cells amongst which 1-O-p-hydroxycinnamoylglucose could show considerable anticancer activity (10).

The seed extract of *C. kotschyana* induced apoptosis in breast cancer cell line (MCF7) (77). As the major active phytoconstituents of *C. kotschyana* are flavonols, quercetin, and kaempferol (231) and quercetin has been found to reduce cell viability of quite a lot of cancer cell lines *in vitro* (268-269). Therefore, these facts are consistent with results that the exposure of MCF7 cells to *C. kotschyana* considerably reduced viability (77).

*C. campestris* also has anticancer agents (270). Detection and evaluation of phytochemicals suggested that eugenol epoxide, lutein epoxide, and lupeol epoxide formed the most active fractions and exhibited the cytotoxic effects against breast cancer cells (271). In a recent effort, efficacy of a Korean herbal formula Ga Gam Nai Go Hyan containing *C. japonica* against benign prostatic hyperplasia was evaluated. This herbal prescription significantly decreases prostate weight by regulating inflammatory responses and apoptosis (92). There is need to develop new technologies such as nanoparticles to improve the therapeutic effect of compounds isolated from these plants. Further efforts may be used to design sustained and targeted drug release systems to improve avoiding side effects.

### Immunological effects

Ethanol extract of *C. chinensis* showed considerable adjuvant potentials towards cellular and humoral immune responses in mice models and can be used as vaccine adjuvants. Extract enhanced specific antibodies (IgG, IgG1, and IgG2b) to a noticeably high level by affecting Th1 and Th2 cell functions (272). Dendritic cells play a key role in regulating immune responses



and are a major target to develop immune modulators. n-butanol and methanol extracts exhibited the immunosuppressive effect on dendritic cells. Kaempferol was identified as the main flavonoid of methanol fraction. Results suggest that kaempferol has potential to treat chronic inflammatory and autoimmune diseases (273). Furthermore, aqueous extract of *C. chinensis* also improved the immune responses (274). *C. chinensis* can protect against tertiary butyl hydroperoxide induced murine osteoblastic MC3T3-E1 cell injury. Aqueous extract of seeds protected cells in a dose-dependent manner by modulating the oxidative stress-induced apoptosis probably owing to its antioxidant potential (240). *C. australis* may act as an immunopotentiator for mammals by increasing the percentage of phagocytosis (275). *C. australis* hyperoside can decrease T or B lymphocyte proliferation and phagocytic activity of the peritoneal M and mediate immune regulation (276).

### Effect on the neuronal system

*C. chinensis* can act as a neuroactive agent and improves memory by inducing cell differentiation. Glycoside of the plant induced neuronal differentiation in rat pheochromocytoma PC12 cells (277). In another experiment, *C. chinensis* improved memory and inhibited acetylcholinesterase activity in scopolamine-induced dymnesia mice (278). Oral administration of its aqueous extract recovered the ischemia-induced lethal damage of neurons and prevented learning disability (51). A traditional Chinese formula Wu-Zi-Yan-Zong containing *C. chinensis* suppresses neuroinflammatory responses and can act as an effective therapeutic agent to prevent and treat neuroinflammatory defects (279).

### Anti-aging activities

*C. chinensis* is an important antiaging prescription of the Chinese herbal medicinal system. Various experimental efforts have been employed to test the certainty of the claim. Polysaccharides of *C. chinensis* can exhibit anti-aging effects by scavenging free radicals and opposing lipid peroxidation (280). Ethanolic extract of *C. chinensis* significantly suppressed the non-enzymatic glycosylation of D-galactose-induced rat aging model (281). Various research reports obviously show that it can regulate immune responses, prolong cell cycle, positively affect body metabolism, improve physiology of internal body organs, and stress management, which proves its anti-aging effects (282).

### Antihypertensive

Ethanolic extract of *C. reflexa* decreased arterial blood pressure and heartbeat rate in Pentothal anesthetized rats. Experimental data indicated that it is a non-specific depressant on all the isolated tissues tested (283). In the course of experiments, ethyl acetate fraction of *C. japonica* exhibited distinctive angiotensin-converting enzyme (ACE) inhibition at a dose of 400 mg/ml. Four caffeoylquinic acid derivatives were isolated from the active fraction having inhibitory effects on ACE activity. Presence of these metabolites, at least in part is responsible for the antihypertensive activity extract (229).

### Anti-osteoporotic activity

*C. chinensis* effectively boasted tissue regeneration

of damaged bones by promoting the formation of osteoblasts from their precursor cells (284). It has been demonstrated in an experimental report that aqueous extract of *C. chinensis* significantly stimulated the differentiation and proliferation of osteoblasts in rat bone cells, but the osteoclasts activities were inhibited (285-286). Antagonistically antiosteoporotic effect of *C. chinensis* was also observed. Five flavonoids were isolated from which kaempferol and hyperoside were found osteogenic in nature (22).

### Renoprotective effects

Aqueous and alcoholic extract of *C. reflexa* exhibited substantial diuretic activity in Wister rats. Total urine volume and Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup> concentration was estimated after a dose of 300 mg/kg extract. There was a marked rise in Na<sup>+</sup> and K<sup>+</sup> excretion (45). *C. chinensis* has been used as a kidney tonic since ancient times. Effect of seed extract on renal function parameters in the rat model having ischemia/reperfusion-induced acute renal failure was studied. Results indicate that *C. chinensis* extract ameliorates renal functions and regulates urine concentration (287).

### Effect on the reproductive system

*C. reflexa* has an antifertility effect. Methanolic extract arrested the normal estrus cycle and decreased ovarian and uterus weight in adult female mice. Flavonoids are reported as antifertility agents, and *C. reflexa* is rich in flavonoids, so results can be attributed to the presence of such compounds (42).

*C. chinensis* extract, and its isolations can improve reproductive systems of both males and females. Ethanolic extract of *C. chinensis* induces a relaxing effect on cavernous penile tissue and may improve erectile dysfunction conditions (288). Many formulations of *C. chinensis* with other herbal prescriptions enhanced penile erection, improved erectile dysfunction, infantile uterus, and motility of sperm (154, 289-291). An herbal formula, KH-204 containing *C. chinensis*, ameliorates erectile dysfunction by its antioxidant and lipid profile improving property (292). Effect of various flavonoids from *C. chinensis* on sex hormones, and prevention of induced and threatened abortion were evaluated by measuring different parameters in a mice model (293-297).

### Anti-mutagenic activity

Mutations elicit an innate metabolic defect in regular cellular systems and lead to morbidity and mortality in mutated organisms. Therefore, exploration for novel bioactive phytochemicals to encounter promutagenic and carcinogenic effects is a subject of keen interest (298). Preliminary evaluation of methanolic extract of *C. chinensis* suppressed 90 % of mutagenic effect against Trp-P-1 in the Ames test, suggesting it as a potential antimutagenic agent (299).

Mutagenic and antimutagenic effects of *C. reflexa* were also studied by the Ames test against well-known positive mutagens including 2-aminofluorine, 4-nitro-o-phenylenediamine, and sodium azide in *Salmonella typhimurium* (TA 98 and TA 100) bacterial strains. The extract revealed noteworthy antimutagenic activity against 4-nitro-o-phenylenediamine and sodium azide

for *S. typhimurium* strains (122).

### Cardiovascular activities

The aging process is accompanied by so many diseases like diabetes, cancer, dementia, and cardiovascular diseases. Heart diseases, leading causes of mortality are due to cardiomyocyte apoptosis which play a key role in myocardial damage and heart failure (300-302). In an experiment, effect of polysaccharide of *C. chinensis* was investigated on D-galactose induced apoptosis of cardiomyocytes in an aging rate model. Apoptosis parameter evaluation indicated that polysaccharide extract decreased the apoptosis of cardiomyocytes (303). *C. chinensis* extract can increase coronary blood flow and decrease myocardial oxygen consumption (304).

### CNS depressant activities and anti-depressant activities

Central nervous system (CNS) disorders comprise 12 % of deaths worldwide and are still a hugely challenging endeavor for health care systems. Plenty of *Convolvulaceae* species, including *Cuscuta* members, are used to treat CNS related diseases traditionally and might be used as alternatives (184).

*C. campestris* affects the CNS action and decreases motor activity of mice sited on a rotarod. Various tests applied indicated the CNS-depressant activity of the extract, which probably seems due to an anesthetizing effect (8, 47). In another experimental trial, methanolic extract of *C. reflexa* served as a good anxiolytic agent in mice at a dose of 400 mg/kg (305).

*C. chinensis* methanolic extract considerably reduced immobility times estimated by FST forced swimming test, which reveals its antidepressant activity (306). While its aqueous extract shows CNS-depressant activity in mice by reducing motor activity and the tonic/clonic phases of electrically-induced seizures in rats (157). Recently a Chinese herbal medicine, Tiansi liquid, containing *C. chinensis* was evaluated for its antidepressant activity, and possible mechanism of action was predicted by *in silico* study (307). Capsules of *C. planiflora* (500 mg) prepared by a pharmacist were found effective for major depression patients. In a study period of eight weeks depression was measured before and after by Beck Depression Inventory and Hamilton Depression Inventory (308).

### Effect on melanin production

*C. chinensis* can promote melanogenesis of amelanotic melanocytes and improved the tyrosinase activities (247-248). Furthermore, it significantly enhanced skin melanin and tyrosinase production. It also positively affected vitiligo treatment in guinea pigs (309). Moreover, there is another report on melanogenesis effect of *C. chinensis* seeds aqueous and ethanolic extracts both *in vitro* and *in vivo*. The aqueous extract showed inhibitory effect on tyrosinase, while the ethanolic extract displayed the opposite effect in tyrosinase activity (160). In a similar study aqueous and ethanolic extracts of *C. chinensis* seeds significantly influenced the melanogenesis by regulating the activity of tyrosinase (310). Consumption of the *C. chinensis* extract with milk reduced the melatonin synthesis and thus ameliorated

the elimination of melasma (311).

*C. japonica* has an inhibitory effect on mushroom tyrosinase activity (312). It can also be used to improve hyperpigmentation. It was ascertained by the treatment of alpha-melanocyte-stimulating hormone-induced melanogenesis with aqueous extract in mouse melanoma cells (69).

### Anti hair fall and anthelmintic activities

Hair loss is a feared side effect of chemotherapy and creates a psychologically distressing condition among millions of men and women due to the deprivation of their major esthetic display feature. Plants as hair growth promoters have found their use in almost all traditional medicinal systems. *C. reflexa* extract is useful in the treatment of alopecia by promoting hair growth (40, 313). Methanolic extract of *C. chinensis* was used as an anthelmintic drug against *Dactylogyrus intermedius* in goldfish (314).

### Analgesic and psychopharmacological

*C. campestris* has analgesic properties. The whole plant grown on *Nerium indicum* was studied. Acetic acid induced writhing test and heat conduction method were used to study the described activity in an animal model. A dose of 400 mg/kg methanolic extract gave significant results as compared to standard Diclofenac sodium (46). In a similar experiment, protecting response against p-benzoquinone-induced writhing was studied by giving a dose of 100 mg/kg to mice, which suggested the analgesic activity of the extract (47). *C. chinensis* also has a pain-relieving ability which was examined by using acetic acid-induced writhing response and formalin-induced paw licking method (256).

Petroleum ether extract of *C. reflexa* noticeably decreased the spontaneous activity and behavior profile of Swiss albino mice. Steroids, the major constituents of the extract may be responsible for such changes (39). *C. japonica* treatment improved the cognitive function of mice in a dose-dependent manner. Novel object recognition and passive avoidance test proved that it might improve learning and memory (315).

### Antipyretic and antiulcer

Antipyretics agents lessened the body temperature in fever. Efficacy of *C. reflexa* as an antipyretic agent was confirmed in yeast induced pyrexia in rats. Aqueous and ethanolic extracts were both found active and started rectal temperature decline after three hours of dose. A dose of 400 mg/kg weight reduced the elevated temperature approximately 83.8 % (ethanolic) and 79 % (aqueous) as compared to the standard drug (96.5 %, Paracetamol) after six hours of treatment (48). *C. campestris* markedly lowered the body temperature of hyperthermic and normothermic mice (47).

Lyophilized raw extract of *C. racemosa* possesses antiulcer activity, which was ascertained by a test showing 44.22 % rate of activity, and 37.05 % rate of cure against acute and sub-chronic models of ulcers, respectively (52).

### Anticonvulsant and anti-obesity

*C. epithymum* have effective anticonvulsant constituents and delayed the onset of seizure (316).

Methanolic extract of *C. reflexa* stem demonstrated preventive effects against convulsion created by chemical agents in mice. Catecholamines levels augmented considerably. After a six-week treatment,  $\gamma$ -aminobutyric acid (GABA) involved in seizure activity was noticeably increased in the brains of mice (317). Ethanolic extract of *C. reflexa* significantly reduced convulsions by delaying onset and duration of seizures in an albino mice model. A dose of 400 mg/kg showed maximum delay in pentylenetetrazole induced convulsions (238).

*C. pedicellata* is widely used for management of obesity. Ethanolic extract of *C. pedicellata* has significantly reduced the bodyweight along with serum lipid profile in high-fat diet-fed rats (26). Recently, polyphenols are reported to possess anti-obesity activity (318).

### Cytotoxicity, insecticidal, antiarthritic, and wound healing activity

The ethanolic extract *C. reflexa*, parasitizing *Nerium oleander*, exhibited promising cytotoxic activity (208). Lectin-like glycoproteins isolated from *C. europaea* demonstrated the cytotoxic effects of LLP and LLP on C127 and B-16 cells (319). Various extracts of the plant have larvicidal potential against mosquitoes (320). *C. reflexa* protects against arthritis and nephrotoxicity. A dose at 600 mg/kg considerably reduced paw edema and joint swelling up to 71.22 % (321). Aqueous and ethanolic extracts of *C. reflexa* stem at 200 mg/kg and 400 mg/kg were able to heal wounds in a rat model (322).

### Conclusion

*Cuscuta* genus is a rich and diverse source of many valuable chemical components. It is loaded with flavonoids, alkaloids, lignans, polysaccharides, steroids, volatile oils, and resin glycosides. Medicinal importance of its various species is part of prehistoric texts. Traditionally it is considered a miracle genus equipped with broad spectrum of remedial values. Decoctions, extracts, paste, powder, juice, and infusions of different parts of the plants are important herbal prescriptions in traditional medicinal systems.

A lot of experimentation has been employed to verify its phytotherapy as claimed by traditional healers and local inhabitants. *C. reflexa*, *C. chinensis*, *C. pedicellata*, *C. approximate*, *C. monogyna*, *C. campestris*, and *C. mitraeformis* have shown impressive antioxidant activity. *C. chinensis*, *C. australis*, *C. reflexa*, and *C. epithymum* are significantly hepatoprotective in nature. Some species of *Cuscuta* including *C. reflexa*, *C. chinensis*, *C. campestris*, *C. japonica*, and *C. kotschyana* have been reported potentially antitumor against various cancer cell lines. Moreover crude extracts and compounds from the various parts possessed antibacterial, antiosteoporotic, anti-inflammatory, antihypertensive, analgesic, anti hair fall, analgesic, and antiestrogenic properties.

Rich and unrivaled medicinal history demands verification with modern scientific methodologies. Only a few of the species are thoroughly investigated up till now, especially *C. reflexa* and *C. chinensis* out of nearly 170, while the rest of the members are partially or fully undiscovered in terms of phytochemistry and pharmacology. Most of the efforts are limited to *in*

*vitro* and *in vivo* animal models or cell line level. Very few clinical studies are reported in humans. Although a good deal of secondary metabolites with multitudinous pharmacological attributes have been isolated, identified, and characterized but most of the pharmacological investigations are extract-based. Further studies must be conducted to clarify the mechanism and to figure out the active principle behind the activity to use these compounds as leads and template in development of new drugs.

### References

- Sermakkani M, Thangapandian V. GC-MS analysis of *Cassia italica* leaf methanol extract. *Asian J Pharm Clin Res* 2012; 5:90-94.
- Gulfraz M, Sadiq A, Tariq H, Imran M, Qureshi R, Zeenat A. Phytochemical analysis and antibacterial activity of *Eruca sativa* seed. *Pak J Bot* 2011; 43:1351-1359.
- Ramasamy S, Manoharan AC. Antibacterial effect of volatile components of selected medicinal plants against human pathogens. *Asian J Microbiol Biotechnol Environ Sci* 2004; 6:209-210.
- Hoareau L, DaSilva EJ. Medicinal plants: a re-emerging health aid. *Electron J Biotechnol* 1999; 2:3-4.
- Gurib-Fakim A. Medicinal plants: traditions of yesterday and drugs of tomorrow. *Mol Aspects Med* 2006; 27:1-93.
- Rates SM. Plants as source of drugs. *Toxicol* 2001; 39:603-613.
- Balandrin MF, Klocke JA. Medicinal, aromatic, and industrial materials from plants. In *Medicinal and Aromatic Plants I*. Berlin. Heidelberg: Springer 1988. p.3-36.
- Prajapati ND, Purohit SS. *Agro S Colour Atlas of Medicinal Plants*. Agrobios (India); 2003.
- Rojas R, Bustamante B, Bauer J, Fernández I, Albán J, Lock O. Antimicrobial activity of selected Peruvian medicinal plants. *J Ethnopharmacol* 2003; 88:199-204.
- Riaz M, Bilal A, Ali MS, Fatima I, Faisal A, Sherkheli MA, et al. Natural products from *Cuscuta reflexa* Roxb. with antiproliferative activities in HCT116 colorectal cell lines. *Nat Prod Res* 2017; 315:583-587.
- Kaul K, Jaitak V, Kaul VK. Review on pharmaceutical properties and conservation measures of *Potentilla fulgens* Wall. ex Hook-a medicinal endangered herb of higher Himalaya. *Indian J Nat Prod Resour* 2011; 2:298-306.
- Benkeblia N. Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and garlic (*Allium sativum*). *Food Sci Technol* 2004; 37:263-268.
- Anjum F, Bukhari SA, Shahid M, Anwar S, Afzal M and Akhter N. Comparative evaluation of antioxidant potential of parasitic plant collected from different hosts. *J Food Process Technol* 2013; 4:1-6.
- Jafari E, Bahmanzadegan A, Ghanbarian G, Rowshan V. Antioxidant activity and total phenolic content from aerial parts of three *Cuscuta* species. *Anal Chem Lett* 2015; 5:377-384.
- Bhagat M, Arora JS, Saxena AK. *In vitro* and *in vivo* antiproliferative potential of *Cuscuta reflexa* Roxb. *J Pharm Res* 2013; 6:690-695.
- Rao VS, Dasaradhan P, Krishnaiah KS. Antifertility effect of some indigenous plants. *Indian J Med Res* 1979; 70:517-520.
- Costa-Lotufo LV, Khan MT, Ather A, Wilke DV, Jimenez PC, Pessoa C, et al. Studies of the anticancer potential of plants used in bangladeshi folk medicine. *J Ethnopharmacol* 2005; 99:21-30.

18. Begum HA, Hamayun M, Zaman K, Hussain A, Ruaf M. Phytochemical evaluation of ethnobotanically selected medicinal plants of mardan, pakistan. *J Adv Bot Zool* 2015; 3:1-5.
19. Qureshi R, Bhatti GR. Ethnobotany of plants used by the thari people of nara desert, pakistan. *Fitoterapia* 2008; 79:468-473.
20. Sharma H, Kumar A. Ethnobotanical studies on medicinal plants of rajasthan (india): a review. *J Med Plants Res* 2011; 5:1107-1112.
21. Malhotra SP, Dutta BK, Gupta R, Gaur YD. Medicinal plants of the indian arid zone. *J Agric Tradit Bot Appl* 1966; 13:247-288.
22. Yang L, Chen Q, Wang F, Zhang G. Antiosteoporotic compounds from seeds of *Cuscuta chinensis*. *J ethnopharmacol* 2011; 135:553-560.
23. Schmelzer GH, Gurib-Fakim A. (2013). Plant resources of tropical Africa 11 (2): medicinal plants 2. Plant resources of tropical Africa 11: Medicinal Plants 2 P. 101-105
24. Sharma L, Khandelwal S. Weeds of rajasthan and their ethno-botanical importance. *Stud Ethno-Med* 2010; 4:75-79.
25. Jang IM. Treatise on asian herbal medicines. Seoul: Haksul-pyunsu-kwan in Research Institute of Natural Products of Seoul National University. 2003.
26. Zekry SH, Abo-elmatty DM, Zayed RA, Radwan MM, ElSohly MA, Hassanean HA, et al. Effect of metabolites isolated from *Cuscuta pedicellata* on high fat diet-fed rats. *Med Chem Res* 2015; 24:1964-1973.
27. Raza MA, Mukhtar F, Danish M. *Cuscuta reflexa* and *Carthamus Oxyacantha*: potent sources of alternative and complimentary drug. SpringerPlus. 2015; 4:76-82.
28. Inamdar FB, Oswal RJ, Chorage TV, Garje K. *In vitro* antimicrobial activity of *Cuscuta reflexa* ROXB. *Int Res J Pharm* 2011; 2:214-216.
29. Kalita D, Saikia J. Ethnomedicinal, Antibacterial and antifungal potentiality of *Centella asiatica*, *Nerium indicum* and *Cuscuta reflexa* -widely used in tiwa tribe of morigaon district of assam, india. *Int J Phytomed* 2012; 4:380-385.
30. Nisa M, Akbar S, Tariq M, Hussain Z. Effect of *Cuscuta chinensis* water extract on 7, 12-dimethylbenz [a] anthracene-induced skin papillomas and carcinomas in mice. *J Ethnopharmacol* 1986; 18:21-31.
31. Umehara K, Nemoto K, Ohkubo T, Miyase T, Degawa M, Noguchi H. Isolation of a new 15-membered macrocyclic glycolipid lactone, Cuscutic Resinoid a from the seeds of *Cuscuta chinensis*: a stimulator of breast cancer cell proliferation. *Planta Med* 2004; 70:299-304.
32. Balakrishnan BR, Sangameswaran B, Bhaskar VH. Effect of methanol extract of *Cuscuta reflexa* aerial parts on hepatotoxicity induced by antitubercular drugs in rats. *Int J Appl Res Nat Prod* 2010; 3:18-22.
33. Yen FL, Wu TH, Lin LT, Lin CC. Hepatoprotective and antioxidant effects of *Cuscuta chinensis* against acetaminophen-induced hepatotoxicity in rats. *J Ethnopharmacol* 2007; 111:123-128.
34. Borole SP, Oswal RJ, Antre RV, Kshirsagar SS, Bagul YR. Evaluation of anti-epileptic activity of *Cuscuta reflexa* Roxb. *Res J Pharm Biol Chem Sci* 2011; 2:657-663.
35. Amol P, Vikas P, Kundan C, Vijay P, Rajesh C. *In vitro* free radicals scavenging activity of stems of *Cuscuta reflexa*. *J Pharm Res* 2009; 2:58-61.
36. Bao X, Wang Z, Fang J, Li X. Structural features of an immunostimulating and antioxidant acidic polysaccharide from the seeds of *Cuscuta chinensis*. *Planta Med* 2002; 68:237-243.
37. Noureen S, Noreen S, Ghumman SA, Batool F, Arshad M, Noreen F, et al. Seeds of giant dodder (*Cuscuta reflexa*) as a function of extract procedure and solvent nature. *Not Bot Hort Agrobot Cluj* 2018; 46:653-662.
38. Anis E, Anis I, Ahmed S, Mustafa G, Malik A, Afza N, et al.  $\alpha$ -glucosidase inhibitory constituents from *Cuscuta reflexa*. *Chem Pharm Bull* 2002; 50:112-114.
39. Pal DI, Panda CH, Sinhababu SA, Dutta AR, Bhattacharya SH. Evaluation of psychopharmacol effects of petroleum ether extract of *Cuscuta reflexa* Roxb stem in mice. *Acta Pol Pharm* 2003; 60:481-486.
40. Pandit S, Chauhan NS, Dixit VK. Effect of *Cuscuta reflexa* Roxb on androgen-induced alopecia. *J Cosmet Dermatol* 2008; 7:199-204.
41. Roy RK, Thakur M, Dixit VK. Development and evaluation of polyherbal formulation for hair growth-promoting activity. *J Cosmet Dermatol* 2007; 6:108-112.
42. Gupta M, Mazumder UK, Pal DK, Bhattacharya S. Anti-steroidogenic activity of methanolic extract of *Cuscuta reflexa* Roxb. stem and *Corchorus olitorius* Linn. seed in mouse ovary. *Indian J Exp Biol* 2003; 41:641-644.
43. Katiyar NS, Rao NV, Gangwar AK. Evaluation of anti-inflammatory activity of stem extracts of *Cuscuta reflexa* Roxb in rats. *Int J Res Pharm Biomed Sci* 2012; 3:1805-1808.
44. Udavant PB, Satyanarayana SV, Upasani CD. Preliminary screening of *Cuscuta reflexa* stems for anti-inflammatory and cytotoxic activity. *Asian Pac J Trop Biom* 2012; 2:1303-1307.
45. Sharma S, Hullatti KK, Prasanna SM, Kuppast IJ, Sharma P. Comparative study of *Cuscuta reflexa* and *Cassytha filiformis* for diuretic activity. *Pharmacogn Res* 2009; 1:327-330.
46. Ghule RS, Venkatanarayan R, Thakare SP, Jain H, Ghule PR. Analgesic activity of *Cuscuta campestris* Yuncker a parasitic plant grown on *Nerium indicum* Mill. *J Adv Pharm Technol Res* 2011; 1:45-51.
47. Agha AM, Sattar EA, Galal A. Pharmacological study of *Cuscuta campestris* Yuncker. *Phytother Res* 1996; 10:117-120.
48. Bhattacharya S, Roy B. Preliminary investigation on antipyretic activity of *Cuscuta reflexa* in rats. *J Adv Pharm Technol Res* 2010; 1:83-87.
49. Mahmood N, Piacente S, Burke A, Khan AL, Pizza C. Constituents of *Cuscuta reflexa* are anti-HIV agents. *Antivir Chem Chemother* 1997; 8:70-74.
50. Rahmatullah M, Sultan S, Toma T, Lucky S, Chowdhury M, Haque W, et al. Effect of *Cuscuta reflexa* stem and *Calotropis procera* leaf extracts on glucose tolerance in glucose-induced hyperglycemic rats and mice. *Afri J Tradit Complementary Altern Med* 2010; 7:109-112.
51. Chung TW, Koo BS, Choi EG, Kim MG, Lee IS, Kim CH. Neuroprotective effect of a chuk-me-sun-dan on neurons from ischemic damage and neuronal cell toxicity. *Neurochem res* 2006; 31:1-9.
52. Ferraz HO, Silva MG, Kato ETM, Barros S, Bacchi EM. Antiulcer and antioxidant activities and acute toxicity of extracts of *Cuscuta racemosa* Mart (Convolvulaceae). *Lat Am Jo Pharm* 2011; 30:1090-1097.
53. Teware K. Pytochemical extraction and TLC estimation of extract of *Cuscuta reflexa*. *World J Pharm Pharm Sci* 2016; 5:378-384.
54. Kuijt J. The biology of parasitic flowering plants. University of California Press, Berkeley; 1969.
55. Liao GI, Chen MY, Kuoh CS. *Cuscuta* L. (Convolvulaceae) in Taiwan. *Taiwania* 2000; 45:226-34.
56. Parker C, Riches CR. Parasitic weeds of the world: biology and control. CAB International; 1993.
57. Yuncker TG. The genus *Cuscuta*. *Mem Torrey Bot Club*

- 1932; 18:109-331.
58. Chrtek J, Osbornová J. Notes on the synanthropic plants of Egypt 3. *Grammica campestris* and other species of family *Cuscutaceae*. *Folia Geobot Phytotax* 1991; 26:287-314.
59. Cronquist A, Takhtadzhan AL. An integrated system of classification of flowering plants. Columbia University Press; 1981.
60. Dahlgren G. The last Dahlgrenogram. System of Classification of the Dicotyledons. The Davis and Hedge Festschrift. 1989. p. 249-260.
61. Dawson JH, Musselman LJ, Wolswinkel PI, Dörr I. Biology and control of *Cuscuta*. *Rev Weed Sci* 1994; 6:265-317.
62. Fang R, Musselman L, Plitmann U, *Cuscuta* In P. Raven and C.Y. Wu (eds.) *Flora Chin* 1995; 16:322-325.
63. Gwo-Ing LI, Ming-Yih CH, Chang-Sheng KU. Pollen morphology of *Cuscuta* (Convolvulaceae) in Taiwan. *Bot Bull Acad Sinica* 2005; 46:75-81.
64. Hadač E, Chrtek J. Notes on the taxonomy of *Cuscutaceae*. *Folia Geobot* 1970; 5:443-445.
65. Täckholm V, Boulos L. Supplementary notes to Student's flora of Egypt. Cairo Univ. Herbarium; 1974.
66. Takhtajan A. Flowering Plants: Origin and Dispersal, Oliver and Boyd, Edinburgh; 1969.
67. Patel S, Sharma V, Chauhan NS, Dixit VK. An updated review on the parasitic herb of *Cuscuta reflexa* Roxb. *Jo Chin Integr Med* 2012; 10:249-255.
68. Donnapee S, Li J, Yang X, Ge AH, Donkor PO, Gao XM, Chang YX. *Cuscuta chinensis* Lam.: a systematic review on ethnopharmacology, phytochemistry and pharmacology of an important traditional herbal medicine. *J ethnopharmacol* 2014; 157:292-308.
69. Jang JY, Kim HN, Kim YR, Choi YH, Kim BW, Shin HK, et al. Aqueous fraction from *Cuscuta japonica* seed suppresses melanin synthesis through inhibition of the p38 mitogen-activated protein kinase signaling pathway in B16F10 cells. *J ethnopharmacol* 2012; 141:338-344.
70. Folarin RO, Omirinde JO, Bejide R, Isola TO, Usende LI, Basiru A. Comparative hepatoprotective activity of ethanolic extracts of *Cuscuta australis* against acetaminophen intoxication in wistar rats. *Int Sch Res Notices* 2014; 2014:1-6.
71. Dangwal LR, Rana CS, Sharma A. Ethno-medicinal plants from transitional zone of Nanda evi Biosphere Reserve, District Chamoli, Uttarakhand. *India* 2011; 2:116-120.
72. Haq F. The ethno botanical uses of medicinal plants of Allai Valley, Western Himalaya Pakistan. *Int J Plant Res* 2012; 2:21-34.
73. Meena AK, Rao MM. Folk herbal medicines used by the Meena community in Rajasthan. *Asian J Tradit Med* 2010; 5:19-31.
74. Ali A, Haider MS, Hanif S, Akhtar N. Assessment of the antibacterial activity of *Cuscuta pedicellata* Ledeb. *Afri J Biotechnol* 2014; 13:430-433.
75. Lakhdari W, Dehliz A, Acheuk F, Mlik R, Hammi H, Doumandji-mitiche B, et al. Ethnobotanical study of some plants used in traditional medicine in the region of Oued Righ (Algerian Sahara). *J Med Plants Stud* 2016; 4:6-10.
76. Njoroge GN, Bussmann RW. Traditional management of ear, nose and throat (ENT) diseases in Central Kenya. *J Ethnobiol Ethnomed* 2006; 2:54-63.
77. Sepehr MF, Jameie SB, Hajjafari B. The *Cuscuta kotschyana* effects on breast cancer cells line MCF7. *J Med Plants Res* 2011; 5:6344-6351.
78. Villa N, Pacheco Y, Rubio E, Cruz R, Lozoya E. Essential oil composition, carotenoid profile, antioxidant and antimicrobial activities of the parasitic plant *Cuscuta mitraeformis*. *Bol latinoam Caribe plantas med aromát* 2017; 16:463-470.
79. Weimann C, Heinrich M. Indigenous medicinal plants in Mexico: the example of the Nahua (Sierra de Zongolica). *Bot Acta* 1997; 110:62-72.
80. Ballabh B, Chaurasia OP, Ahmed Z, Singh SB. Traditional medicinal plants of cold desert Ladakh—used against kidney and urinary disorders. *J Ethnopharmacol* 2008; 118:331-339.
81. Holm LG, Holm L, Holm E, Pancho JV, Herberger JP. *World weeds: natural histories and distribution*. John Wiley & Sons; 1997.
82. Furuhashi T, Furuhashi K, Weckwerth W. The parasitic mechanism of the holostemparasitic plant *Cuscuta*. *J Plant Interact* 2011; 6:207-219.
83. Kaiser B, Vogg G, Fürst UB, Albert M. Parasitic plants of the genus *Cuscuta* and their interaction with susceptible and resistant host plants. *Front Plant Sci* 2015; 6:45-54.
84. Shen HW, Ye W, Hong L, Huang H, Wang Z, Deng X, et al. Progress in parasitic plant biology: host selection and nutrient transfer. *Plant Biol* 2006; 8:175-185.
85. Kelly CK, Horning K. Acquisition order and resource value in *Cuscuta attenuata*. *Proc Nat Acad Sci* 1999; 96:13219-13222.
86. Nwokocha MI, Aigbokhan EI. Host range and preference of *Cuscuta campestris* (Yunck). among common weeds in Benin city, Nigeria. *Niger J Bot* 2013; 26:1-29.
87. Prather LA. Biology of *Cuscuta Attenuata* Waterfall. *Proc Oklahoma Acad Sci* 1990; 73:7-13.
88. Diggs GM, Lipscomb BL, O'Kennon RJ, Mahler WF, Shinnors LH. Shinnors' and Mahler's illustrated flora of North Central Texas. *Bot Res Inst Texas* 1999.
89. Akter MH, Akter MH, Rahmatullah M. Synergistic antihyperglycemic activity of *Coccinia grandis* leaves and *Cuscuta reflexa* stems. *J Pharm Pharm Sci* 2016; 5:236-243.
90. Vijikumar S. *Cuscuta reflexa* Roxb.—a wonderful miracle plant in ethnomedicine. *Indian J Nat Sci* 2011; 11:676-683.
91. Mavlonov GT, Ubaidullaeva KA, Kadryaeva GV, Kuznetsova NN. Cytotoxic components of *Cuscuta*. *Chem Nat Compd* 2008; 44:409-410.
92. Shin SJ, Lee KH, Chung KS, Cheon SY, An HJ. The traditional Korean herbal medicine Ga-Gam-Nai-Go-Hyan suppresses testosterone-induced benign prostatic hyperplasia by regulating inflammatory responses and apoptosis. *Exp Ther Med* 2017; 13:1025-1031.
93. Talha J, Priyanka M, Akanksha A. Hypertension and herbal plants. *Int Res J Pharm* 2011; 2:26-30.
94. Kuo CS, Liao GI. Flower initiation and development in *Cuscuta australis* R. Br.(Convolvulaceae). *Taiwania* 1993; 38:99-108.
95. Quattrocchi U. *CRC world dictionary of plant names: common names, scientific names, eponyms, synonyms, and etymology*: CRC Press. 2000.
96. Weinberg T, Lalazar A, Rubin B. Effects of bleaching herbicides on field dodder (*Cuscuta campestris*). *Weed sci* 2003; 51:663-670.
97. Joshi SK, Sanjay G. *Cuscuta europaea* Linn. (Dodder plant): an emerging threat to plant diversity of Valley of Flowers. *Curr Sci* 2003; 84:1285-1286.
98. Papuc C, Crivineanu M, Nicorescu V, Predescu C. Reactive oxygen species scavenging activity and hepatoprotective effects of a polyphenolic extract obtained from *Cuscuta Europaea*. *Rev Chim (Bucharest)* 2012; 9:869-873.
99. Jafari EF, Assadi MO, Ghanbarian GA. A revision of *Cuscutaceae* family in Iran. *Iran J Bot* 2016; 22:23-29.
100. Costea M, Stefanović S. Evolutionary history and taxonomy of the *Cuscuta umbellata* complex (Convolvulaceae): evidence

- of extensive hybridization from discordant nuclear and plastid phylogenies. *Taxon* 2010; 59:1783-1800.
101. Hashem A, Patabendige D, Roberts C. Biology and management of red dodder-a new threat to the grains industry. In 15th Australian Weeds Conference, Papers and Proceedings, Adelaide, South Australia, 24-28 September 2006: Managing weeds in a changing climate Weed Management Society of South Australia. 2006 p. 163-166.
  102. Orr GL, Haidar MA, Orr DA. Smallseed dodder (*Cuscuta planiflora*) gravitropism in red light and in red plus far-red. *Weed sci* 1996; 44:795-796.
  103. Farah AF, Al-Abdulsalam MA. Effect of field dodder (*Cuscuta campestris* Yuncker) on some legume crops. *Sci J King Faisal Univ (Basic Appl Sci)* 2004; 5:103-113.
  104. Peng WH, Chen YW, Lee MS, Chang WT, Tsai JC, Lin YC, et al. Hepatoprotective effect of *Cuscuta campestris* Yunck. whole plant on carbon tetrachloride induced chronic liver injury in mice. *Int J Mol Sci* 2016; 17:2056-2067.
  105. Youssef SA. Medicinal and non-medicinal uses of some plants found in the middle region of Saudi Arabia. *J Med Plants Res* 2013; 7: 2501-2517.
  106. Hillman FH. Dodder In Relation To Farm Seeds. US Department of Agriculture. 1907.
  107. Mukhtar I, Atiq M, Hanan A, Iqbal Z. Antifungal activity of *Cuscuta reflexa* Roxb. *Pakistan J Phytopathol* 2012; 24:163-166.
  108. Quattrocchi U. CRC world dictionary of medicinal and poisonous plants: common names, scientific names, eponyms, synonyms, and etymology: CRC Press. 2012.
  109. Shahid M, Rao NK. New records of three Convolvulaceae species to the flora of the United Arab Emirates. *J New Biol Sci* 2016; 5:114-121.
  110. Doyle GJ. *Cuscuta epithimum* (L.) L. (Convolvulaceae), its hosts and associated vegetation in a limestone pavement habitat in the Burren lowlands in county Clare (H9), Western Ireland. In *Biology and Environment: Proceedings of the Royal Irish Academy*; 1993. p.61-67.
  111. Hussain F, Leghari IH, Naveed S. Vegetation in sindh: an analytical and literary study. *Karoonjhar* 2015; 7:11-28.
  112. Piwowarczyk R, Guzikowski S, Góralski G, Denysenko-Bennett M, Kwolek D, Joachimiak AJ. First report of dodder (*Cuscuta epithimum*) parasitizing hemiparasitic species of santalaceae (thesium) and orobanchaceae (euphrasia, melampyrum, odontites, orthantha, and rhinanthus) in Poland. *Plant Dis* 2018; 102:456-460.
  113. Shimi P, Rezapanah MR. A study of *Smicronyx robustus* faust (Curculionidae) as a biological control agent of eastern dodder (*Cuscuta monogyna* Vahl.) *Iran J Agric Sci* 1995; 1:43-51.
  114. Anac E, Kaya I, Tepe I. Determination of alfalfa dodder (*Cuscuta approximata* Bab.) damage on alfalfa (*Medicago sativa* L.) grown in Van, Turkey. In *Proceedings of Joint Workshop of the EWRS Working Groups Weed Management in Arid and Semi-arid Climate and Weed Management Systems in Vegetables* 2011. p. 4-8.
  115. Tepe I, Celebi SZ, Kaya I, Ozkan RY. Control of smoothseed alfalfa dodder (*Cuscuta approximata*) in alfalfa (*Medicago sativa*). *Int J Agric Biol* 2017; 19:199-203.
  116. Bhadrecha P, Kumar V, Kumar M. Medicinal plant growing under sub-optimal conditions in trans-himalaya region at high altitude. *Def Life Sci J* 2017; 2:37-45.
  117. Bibi T, Ahmad M, Tareen RB, Tareen NM, Jabeen R, Rehman SU, et al. Ethnobotany of medicinal plants in district Mastung of Balochistan province-Pakistan. *J Ethnopharm* 2014; 157:79-89.
  118. Petrovska BB. Historical review of medicinal plants' usage. *Pharmacogn Rev* 2012; 6:1-5.
  119. Ogbulie JN, Ogueke CC, Okorundu S. Antibacterial properties of *A. cordifolia*, *M. flurum*, *U. chamae*, *B. pinnatum*, *C. albidum* and *A. ciliata* on some hospital isolates. *Niger J Microbiol* 2004; 18:249-255.
  120. Chopra RN, Nayar L, Chopra IC. Glossary of Indian medicinal plants. New Delhi. C SIR. 1956.
  121. Chopra R, Chopra I Handa K, Kapur L. Indigenous drugs of India UN Dhur and Sons. Pvt. Ltd., Calcutta. 1958. p. 358.
  122. Dokuparthi SK, Banerjee N, Kumar A, Singamaneni V, Giri AK, Mukhopadhyay S. Phytochemical investigation and evaluation of antimutagenic activity of the extract of *Cuscuta reflexa* Roxb by Ames Test. *Int J Pharm Sci Res* 2014; 5:3430-3434.
  123. Saini P, Mithal R, Menghani E. A parasitic medicinal plant *Cuscuta reflexa* : an overview. *Int J Sci Eng Res* 2015; 6:951-959.
  124. Singh S, Sharma A. Studies on ethnomedicinal Plant of Baghicha Jashpur Chattisgarh. *J Sci Lett* 2017; 2:48-55.
  125. Basak S, Banerjee A, Manna CK. Role of some ethno medicines used by the Santal tribal people, of the district Bankura, WB, India, for abortifacient purposes. *J Med Plants Stud* 2016; 4:125-129.
  126. Singh RS, Shahi SK. Diversity of medicinal plants of Ratanpur region of Bilaspur district (Chhattisgarh). *J Med Plants* 2017; 5:276-281.
  127. Singh S. Ethnobotanical study of some climbers of Parsa district forest of Nepal. *J Med Plants* 2016; 4:6-10.
  128. Mohapatra SS, Sarma J, Roy RK, Panigrahi S, Ganguly S. Ethnomedicinal plants used in balasore district of Odisha: a comprehensive report. *Int J Cur Microbiol App Sci* 2018; 7:1959-1963.
  129. Kirtikar KR, Basum BD. *Indian medicinal plants. Vol 1. Delhi: Periodical Experts Book Agency; 1984.*
  130. Darias V, Bravo L, Rabanal R, Mateo CS, Luis RG, Perez AH. New contribution to the ethnopharmacological study of the Canary Islands. *J Ethnopharmacol* 1989 1; 25:77-92.
  131. Chowdhury M, Das AP. Folk medicines used by the Rabha tribe in Coochbehar district of West Bengal: a preliminary report. *Adv Ethnobot* 2007:289-296.
  132. Rai Y, Kumar D. Survey on medicinal climbers in meerut district, Uttar Pradesh, India. *Imperial J Interdisciplinary Res* 2016; 2:603-610.
  133. Patel JN, Patel NK. Study of parasite hosts of the genus *Cuscuta* and its traditional uses in Planpur Taluka, Gujarat, India. *Ethnobot Leaf* 2010; 14:126-135.
  134. Dutta ML. Plants used as ethnomedicine by the *Thengal Kacharies* of Assam, India. *Asian J Plant Sci Res* 2017; 7:7-8.
  135. Khalid M, Bilal M, Hassani D, Zaman S, Huang D. Characterization of ethno-medicinal plant resources of karamar valley Swabi, Pakistan. *J Radiat Res Appl Sci* 2017; 10:152-163.
  136. Khattak NS, Nouroz F, Rahman IU, Noreen S. Ethno veterinary uses of medicinal plants of district Karak, Pakistan. *J ethnopharmacol* 2015; 171:273-279.
  137. Kumar S, Singh BS, Singh RB. Ethnomedicinal plants uses to cure different human diseases by rural and tribal peoples of Hathras district of Uttar Pradesh. *J Pharmacogn Phytochem* 2017; 6:346-348.
  138. Azam MN, Mannan MA, Ahmed MN. Medicinal plants used by the traditional medical practitioners of Barendra and Shamatat (Rajshahi & Khulna Division) region in Bangladesh for treatment of cardiovascular disorders. *J Med Plants* 2014; 2:9-14.

139. Khanday ZH, Singh S. Ethnomedicinal Plants used for curing various skin diseases in Shopian district of Jammu and Kashmir. *J Phytology* 2017; 9:5-6.
140. Senthilkumar S, Vijayakumari K. A review-pharmacology of medicinal plants. *Int J Unvers Pharm Bio Sci* 2016; 5:37-59.
141. Shahidullah M, Al-Mujahidee M, Uddin SN, Hossan MS, Hanif A, Bari S, et al. Medicinal plants of the Santal tribe residing in Rajshahi district, Bangladesh. *Am Eur J Sustain Agric* 2009; 3:220-226.
142. Singh EA, Kamble SY, Bipinraj NK, Jagtap SD. Medicinal plants used by the Thakar tribes of Raigad district, Maharashtra for the treatment of snake-bite and scorpion-bite. *Int J Phytother Res* 2012; 2:26-35.
143. Hossan MS, Hanif A, Khan M, Bari S, Jahan R, Rahmatullah M. Ethnobotanical survey of the Tripura tribe of Bangladesh. *Am Eur J Sustain Agric* 2009; 3:253-261.
144. Patel H, Patel N. Sacred and medicinal plant diversity of patan sacres grove of Patan District (NG). *Life Sci Leaf* 2017; 92:50-60.
145. Siwakoti M, Siwakoti S. Ethnomedicinal uses of plants among the satar tribe of Nepal. *J Econ Taxon Bot* 2000; 24:323-333.
146. Saheb TS, Rao BR, Venkateswarlu M, Swamulu M. Medicinal plants used for jaundice by the tribal people of nallamalais in Andhra Pradesh. *J Pharmacogn Phytochem* 2018; 7:528-531.
147. Divakara BN, Prasad S. Ethnomedicinal importance of invasive alien flora of latehar and hazaribagh districts: Jharkhand. *Indian For* 2015; 141:1172-1175.
148. Mahmud MR, Parvin A, Anny IP, Akter F, Tarannom SR, Moury SI, et al. Home remedies of village people in six villages of Dinajpur and Rangpur districts, Bangladesh. *World J Pharm Pharm Sci* 2015; 4:63-73.
149. Rahmatullah M, Khatun Z, Hasan A, Parvin W, Moniruzzaman M, Khatun A, et al. Survey and scientific evaluation of medicinal plants used by the Pahan and Teli tribal communities of Natore district, Bangladesh. *Afr J Tradit Complementary Altern Med* 2012; 9:366-373.
150. Seliya AR, Patel NK. Ethnomedicinal uses of climbers from Saraswati river region of Patan district, North Gujarat. *Ethnobot leaf* 2009; 13:865-872.
151. Qureshi R, Bhatti GR, Memon RA. Ethnomedicinal uses of herbs from northern part of Nara desert, Pakistan. *Pak J Bot* 2010; 42:839-851.
152. Patil JU, Biradar SD. Folkloric medicinal plants of Hingoli district, Maharashtra. 2011; 2:97-101.
153. Van Sam H, Baas P, Kebler PJ. Traditional medicinal plants in Ben En national park, Vietnam. *Blumea Biodivers Evol Biogeogr Plants* 2008; 53:569-601.
154. Sohn DW, Kim HY, Kim SD, Lee EJ, Kim HS, Kim JK, et al. Elevation of intracavernous pressure and NO-cGMP activity by a new herbal formula in penile tissues of spontaneous hypertensive male rats. *J ethnopharmacol* 2008; 120:176-180.
155. Deepakkumar R, Sabari E, Karthick M, Raysad KS. Traditionally used ethno-medicinal plants of the Kurumba communities surrounded in Thalimalai hills, Namakkal district, Tamil Nadu. *South Indian J Biol Sci* 2017; 3:15-26.
156. Patil SJ, Patil HM. Ethnomedicinal herbal recipes from satpura hill ranges of shirpur tahsil, dhule, maharashtra. *India Res J Recent Sci* 2012; 1:333-336.
157. Akbar S, Nisa M, Tariq M. CNS depressant activity of *Cuscuta chinensis* Lam. *Int J Crude Drug Res* 1985; 23:91-94.
158. Fahmy GM. Qatar biodiversity newsletter. *Ostrich* 2008; 2:1-5.
159. Rizk AM, El-Ghazaly GA. Medicinal and poisonous plants of Qatar. University of Qatar; 1995.
160. Wang TJ, An J, Chen XH, Deng QD, Yang L. Assessment of *Cuscuta chinensis* seeds' effect on melanogenesis: comparison of water and ethanol fractions *in vitro* and *in vivo*. *J Ethnopharmacol* 2014; 154:240-248.
161. Shubhangi P, Patil DA. Herbal haircare as revealed by people in Jalgaon district, Maharashtra, India. *J Exp Sci* 2012; 3:32-34.
162. Ghayoumi A, Mashayekhi A. Scleroderma treatment in iranian traditional medicine: a case report. *Adv Herb Med* 2016; 2:1-4.
163. Tavili A, Farajollahi A, Pouzesh H, Bandak E. Treatment induced germination improvement in medicinal species of *Foeniculum vulgare* Miller and *Cuscuta epithymum* (L.) L. *Mod Appl Sci* 2010; 4:163-169.
164. Haq F, Ahmad H, Alam M. Traditional uses of medicinal plants of Nandiar Khuwarr catchment (District Battagram). *Pakistan J Med Plants Res* 2011; 5:39-48.
165. Dangwal LR, Sharma A, Rana CS. Ethnomedicinal plants of the Garhwal Himalaya used to cure various diseases: a case study. *N Y Sci J* 2010; 3:28-31.
166. Nedelcheva A, Pavlova D, Krasteva I, Nikolov S. Medicinal plants biodiversity and their resources of one serpentine site in the Rhodope MTS (Bulgaria). *Nat Montenegr* 2010; 9:373-387.
167. Uniyal B, Shiva V. Traditional knowledge on medicinal plants among rural women of the Garhwal Himalaya, Uttaranchal. *Indian J Tradit Knowl* 2005; 4:259-266.
168. Naz R, Ayub H, Nawaz S, Islam ZU, Yasmin T, Bano A, et al. Antimicrobial activity, toxicity and anti-inflammatory potential of methanolic extracts of four ethnomedicinal plant species from Punjab, Pakistan. *BMC Complement Altern Med* 2017; 17:302-315.
169. Nita RD, Haresh DL. Ethno-botanical survey of some medicinal plants in jatasankar region of girnar forest, gujarat, india. *Glob J Res Med Plants Indig Med* 2013; 2:830-841.
170. Paudel N, Adhikari DC, Das BD. Some medicinal plants uses in ethnical group from batnagar, eastern, Nepal. *Am Sci Res J Eng Tech Sci* 2018; 41:233-239.
171. Ahirwar RK. Diversity of ethnomedicinal plants in Boridand forest of district Korea, Chhattisgarh, India. *Am J Plant Sci* 2015; 6:413-425.
172. Yaseen G, Ahmad M, Potter D, Zafar M, Sultana S, Mir S. Ethnobotany of medicinal plants for livelihood and community health in deserts of Sindh-Pakistan. In *Plant and Human Health, Volume 1*. Springer, Cham. 2018. p. 767-792.
173. Kala CP. Ethnomedicinal botany of the Apatani in the Eastern Himalayan region of India. *J Ethnobiol Ethnomed* 2005; 1:1-8.
174. Akter MH, Akter MH, Prophan MT, Akter S, Akter N, Sultana J, et al. Documentation of plant-based remedies of a folk herbalist of Comilla district, Bangladesh. *World J Pharm Pharm Sci* 2017; 6:1-11.
175. Khatun MM, Rahma M. Medicinal plants used by the village Pania under Baghmara District, Bangladesh. *Discovery* 2018; 54:60-71.
176. Azam MN, Ahmed MN, Rahman MM, Rahmatullah M. Ethnomedicines used by the Oraon and Gor tribes of Sylhet district, Bangladesh. *Am-Eurasian J Sustain Agric* 2013; 7:391-402.
177. Verma N, Yadav RK. *Cuscuta reflexa*: a paracitic medicinal plant. *Plant Arch* 2018; 18:1938-1942.
178. Kumar S, Sharma SD, Kumar N. Ethnobotanical study of some common plants from district hamirpur of Himachal Pradesh (India). *Int J Adv Res* 2015; 3:492-496.

179. Shipa A, Koli S, Akter K, Shahriar SS, Rahmatullah M. Phytotherapeutic practices of a folk medicinal practitioner in Kishoreganj district, Bangladesh. *J Med Plants* 2018; 6:240-242.
180. Khan W, Khan SM, Ahmad H. Ethno-ecology, Human Health and Plants of the Thandiani Sub Forest Division, Abbottabad, KP, Pakistan. In *Plant and Human Health, Volume 1*. Springer, Cham. 2018. p. 547-567.
181. Chen GT, Lu Y, Yang M, Li JL, Fan BY. Medicinal uses, pharmacology, and phytochemistry of convolvulaceae plants with central nervous system efficacies: a systematic review. *Phytother Res* 2018; 32:823-864.
182. Ganapaty SE, Ramaiah MA, Yasaswini KA, Kumar CR. Determination of total phenolic, flavonoid, alkaloidal contents and in vitro screening for hepatoprotective activity of *Cuscuta epithymum* (L) whole plant against CCl<sub>4</sub> induced liver damage animal model. *Int J Pharm Pharm Sci* 2013; 5:738-742.
183. Sahranavard S, Ghafari S, Mosaddegh M. Medicinal plants used in Iranian traditional medicine to treat epilepsy. *Seizure* 2014; 23:328-332.
184. Farnsworth NR, Morris RW. Higher plants-the sleeping giant of drug development. *Am J Pharm Sci Support Public Health* 1976; 148:46-52.
185. Dillard CJ, German JB. Phytochemicals: nutraceuticals and human health. *J Sci Food Agric* 2000; 80:1744-1756.
186. Ahmad M, Khan MA, Zafar M, Sultana S. Ethnomedicinal demography and ecological diversification of some important weeds from district attock Pakistan. *Pak J Weed Sci Res* 2006; 12:37-46.
187. Dhalwal K, Shinde VM, Mahadik KR, Namdeo AG. Rapid densitometric method for simultaneous analysis of umbelliferone, psoralen, and eugenol in herbal raw materials using HPTLC. *J Sep Sci* 2007; 30:2053-2058.
188. Mir MA, Sawhney SS, Jassal MM. Qualitative and quantitative analysis of phytochemicals of *Taraxacum officinale*. *Wudpecker J Pharm Pharmacol* 2013; 2:1-5.
189. Almodaifer S, Alsibaie N, Alhoumendani G, Alammari G, Kavita MS. Role of phytochemicals in health and nutrition. *BAO J Nutr* 2017; 3:28-34.
190. Savithamma N, Rao ML, Sahrulatha D. Screening of medicinal plants for secondary metabolites. *Middle East J Sci Res* 2011; 8:579-584.
191. Yahara S, Domoto H, Sugimura C, Nohara T, Niiho Y, Nakajima Y, et al. An alkaloid and two lignans from *Cuscuta chinensis*. *Phytochem* 1994; 37:1755-1757.
192. Garcia MR, Erazo GS, Pena RC. Flavonoids and alkaloids from *Cuscuta* (Cuscutaceae). *Biochem Syst Ecol* 1995; 23:571-572.
193. Miyahara K, Du XM, Watamab M, Sugimura C, Yahara S, Nohara T. Resin glycosides. XXIII. Two novel acylated trisaccharides related to resin glycoside from the seeds of *Cuscuta chinensis*. *Chem Pharm bull* 1996; 44:481-485.
194. Du XM, Kohinata K, Kawasaki T, Guo YT, Miyahara K. Components of the ether-insoluble resin glycoside-like fraction from *Cuscuta chinensis*. *Phytochem* 1998; 48:843-850.
195. Yen FL, Wu TH, Lin LT, Cham TM, Lin CC. Concordance between antioxidant activities and flavonol contents in different extracts and fractions of *Cuscuta chinensis*. *Food Chem* 2008; 108:455-462.
196. He XH, Yang WZ, Meng AH, He WN, Guo DA, Ye M. Two new lignan glycosides from the seeds of *Cuscuta chinensis*. *J Asian Nat Prod Res* 2010; 12:934-939.
197. Fan BY, Luo JG, Gu YC, Kong LY. Unusual ether-type resin glycoside dimers from the seeds of *Cuscuta chinensis*. *Tetrahedron* 2014; 70:2003-2014.
198. Wang J, Tan D, Wei G, Guo Y, Chen C, Zhu H, et al. Studies on the Chemical Constituents of *Cuscuta chinensis*. *Chem Nat Compd* 2016; 52:1133-1136.
199. Ibrahim M, Rehman K, Hussain I, Farooq T, Ali B, Majeed I, et al. Ethnopharmacological investigations of phytochemical constituents isolated from the genus *Cuscuta*. *Crit Rev Eukaryot Gene Expr* 2017; 27:113-150.
200. Löffler C, Czygan FC, Proksch P. Phenolic constituents as taxonomic markers in the genus *Cuscuta* (Cuscutaceae). *Biochem Syst Ecol* 1997; 25:297-303.
201. Wink M, Witte L. Quinolizidine alkaloids in *Genista acanthoclada* and its holoparasite, *Cuscuta palaestina*. *J Chem Ecol* 1993; 19:441-448.
202. Ye M, Li Y, Yan Y, Liu H, Ji X. Determination of flavonoids in *Semen Cuscutae* by RP-HPLC. *J Pharm Biomed Anal* 2002; 28:621-628.
203. Siddiqui MS, Memon AA, Memon S, Baloch SG. *Cuscuta reflexa* as a rich source of bioactive phenolic compounds. *J Herbs Spices Med Plants* 2017; 23:157-168.
204. Ramya B, Natrajan E, Vijaykumar S, John Vasanth MS, Muthuramsanjivani VK. Isolation and characterization of bioactive metabolites in *Cuscuta reflexa* Roxb. *Indian J Nat Sci* 2010; 1:134-139.
205. Uddin SJ, Shilpi JA, Middleton M, Byres M, Shoeb M, Nahar L, et al. Swarnalin and cis-swarnalin, two new tetrahydrofuran derivatives with free radical scavenging activity, from the aerial parts of *Cuscuta reflexa*. *Nat Prod Res* 2007; 21:663-668.
206. Tripathi VJ, Yadav SB, Upadhyay AK. A new flavanone, reflexin, from *Cuscuta reflexa* and its selective sensing of nitric oxide. *Appl Biochem Biotechnol* 2005; 127:63-67.
207. Chatterjee DP, Sahu RK. Chemical characterization of the flavonoid constituents of *Cuscuta reflexa*. *UK J Pharm Bio Sci* 2014; 2:13-16.
208. Awasthi LP. The purification and nature of an antiviral protein from *Cuscuta reflexa* plants. *Arch Virol* 1981; 70:215-223.
209. Shekarchi M, Kondori BM, Hajimehdipoor H, Abdi L, Naseri M, Pourfarzib M, et al. Finger printing and quantitative analysis of *Cuscuta chinensis* flavonoid contents from different hosts by RP-HPLC. *Food Nutr Sci* 2014; 5:914-922.
210. Zhan W, Zhisheng H. Studies on the chemical constituents of the seed of chinese dodder (*Cuscuta chinensis*). *Chin Tradit Herb drugs* 1998; 9:115-117.
211. Ye M, Yan Y, Guo DA. Characterization of phenolic compounds in the Chinese herbal drug Tu-Si-Zi by liquid chromatography coupled to electrospray ionization mass spectrometry. *Rapid Commun Mass Spectrom* 2005; 19:1469-1484.
212. Tsai YC, Lai WC, Du YC, Wu SF, El-Shazly M, Lee CL, et al. Lignan and flavonoid phytoestrogens from the seeds of *Cuscuta chinensis*. *J Nat Prod* 2012; 75:1424-1431.
213. Du XM, Sun NY, Nishi M, Kawasaki T, Guo YT, Miyahara K. Components of the ether-insoluble resin glycoside fraction from the seed of *Cuscuta australis*. *J Nat Prod* 1999; 62:722-725.
214. Ferraz HO, Silva MG, Carvalho R, Suffredini IB, Kato E, Arakaki F, et al. Phytochemical study and evaluation of the antimicrobial activity and cytotoxicity of *Cuscuta racemosa*. *Rev Bras Farm* 2011; 21:41-46.
215. Shailajan S, Joshi H. Optimized separation and quantification of pharmacologically active markers quercetin, kaempferol,  $\beta$ -sitosterol and lupeol from *Cuscuta reflexa* Roxb. *J Pharm Res* 2011; 4:1851-1853.
216. Versiani MA, Kanwal A, Faizi S, Farooq AD. Cytotoxic cardiac glycoside from the parasitic plant *Cuscuta reflexa*.



- Chem Nat Compd 2017; 53:915-922.
217. Jahan IA, Akbar PN, Enayetullah M, Ahmmad N, Nuruddin M, Ahmed MR. Elemental and fatty acid content of four medicinal plants: *Kaempferia rotunda*, *Cuscuta reflexa*, *Centella asiatica* and *Asparagus racemosus*. *European J Med Plants* 2015;1-10.
  218. Bais N, Kakkar A. Comparative phytochemical analysis of *Cuscuta reflexa* parasite grown on *Cassia fistula* and *Ficus benghlensis* by GC-MS. *Int J Pharm Pharm Sci* 2013; 5:350-355.
  219. Rath D, Panigrahi SK, Kar DM, Maharana L. Identification of bioactive constituents from different fractions of stems of *Cuscuta reflexa* Roxb. Using GC-MS. *Nat Prod Res* 2017; 32:1977-1981.
  220. Mukherjee R, Bordoloi J, Goswami A, Goswami BC. Carotenoids of dodder (*Cuscuta reflexa*) grown on hedge, *Clerodendrum demryi*. *Adv Nat Appl Sci* 2008; 2:99-103.
  221. Ye M, Yan Y, Ni X, Qiao L. Studies on the chemical constituents of the herba of *Cuscuta chinensis*. *J Chinese Med Mat* 2001; 24:339-341.
  222. Cheng PP, Shi J, Du P, Liu DH, Cao X, Wen X. Fatty acid in the *Cuscuta chinensis* lam by capillary gas chromatography. *Acad Periodical Farm Prod Process* 2013; 8:116-118.
  223. Kwon Y, Chang B, Kim C. Antioxidative constituents from the seeds of *Cuscuta chinensis*. *Nat Prod Sci* 2000; 6:135-138.
  224. Xiang SX, He ZS, Ye Y. Furofuran lignans from *Cuscuta chinensis*. *Chin J Chem* 2001; 19:282-285.
  225. Lin Q, Jia LY, Sun QS. Chemical constituents of the seeds of *Cuscuta chinensis* Lam.[J]. *J Shenyang Pharm Univ* 2009; 12:1-10.
  226. Oh H, Kang DG, Lee S, Lee HS. Angiotensin converting enzyme inhibitors from *Cuscuta japonica* Choisy. *J Ethnopharmacol* 2002; 83:105-108.
  227. Baccarini A, Bertossi F, Bagni N. Carotenoid pigments in the stem of *Cuscuta australis*. *Phytochemistry* 1965; 4:349-351.
  228. Hongzhu G, Jiashi L. Study on constituents of the seed from *Cuscuta Australis*. *J Beijing Univ Tradit Chin Med* 2000; 23:20-23.
  229. Guo H, Li J. Study on flavonoids of *Cuscuta australis* R. Br. *China J Chin Materia Med* 1997; 22:38-39.
  230. Sousa AL, Sales QS, Braz-Filho R, de Oliveira RR. Lignans and flavonoids isolated from *Cuscuta racemosa* Mart. & Humb (Convolvulaceae) by droplet counter-current chromatography. *J Liq Chromatogr R T* 2012; 35:2294-2303.
  231. Bacchi EM. Flavonoids from *Cuscuta racemosa*. *Planta Medi* 1993; 59:605-606.
  232. Abdallah WE, Elsayed WM, Abdelshafeek KA. Chemical constituents and radical scavenging activity of *Cuscuta pedicellata* seed extracts. *Int J ChemTech Res* 2016; 9:580-587.
  233. Szymańska R, Kruk J. Tocopherol content and isomers' composition in selected plant species. *Plant Physiol Biochem* 2008; 46:29-33.
  234. Prior RL, Cao G. Antioxidant phytochemicals in fruits and vegetables: diet and health implications. *Hort Sci* 2000; 35:588-592.
  235. Rice-Evans C. Flavonoids and isoflavones: absorption, metabolism, and bioactivity. *Free Radic Biol Med* 2004; 7:827-828.
  236. Chanda S, Dave R, Kaneria M, Nagani K. Seaweeds: A Novel, Untapped Source of Drugs From Sea to Combat Infectious Diseases. *Current research, Technology And Education Topics In Applied Microbiology Microbial Biotechnology* 2010. p. 473-480.
  237. Tanruean K, Poolprasert P, Kumla J, Suwannarach N, Lumyong S. Bioactive compounds content and their biological properties of acetone extract of *Cuscuta reflexa* Roxb. grown on various host plants. *Nat Prod Res* 2017; 3:1-4.
  238. Hussain SA, Farheen S, Sultana T, Tabassum A, Hussain SI, Khan R. Evaluation of anticovulsant and anioxidant activity of selected medicinal plants. *World J Pharm Pharm Sci* 2017; 6:1899-1914.
  239. Yen FL, Wu TH, Lin LT, Cham TM, Lin CC. Nanoparticles formulation of *Cuscuta chinensis* prevents acetaminophen-induced hepatotoxicity in rats. *Food Chem toxicol* 2008; 46:1771-1777.
  240. Gao JM, Li R, Zhang L, Jia LL, Ying XX, Dou DQ, et al. *Cuscuta chinensis* seeds water extraction protecting murine osteoblastic MC3T3-E1 cells against tertiary butyl hydroperoxide induced injury. *J Ethnopharmacol* 2013; 148:587-595.
  241. Zhen GH, Jiang B, Bao YM, Li DX, An LJ. The protective effect of flavonoids from *Cuscuta chinensis* in PC12 cells from damage induced by H<sub>2</sub>O<sub>2</sub>. *J Chin Med Mater* 2006; 29:1051-1055.
  242. Amaresh P, Seemanchala R, Debashis P, Arpan M, Bijan G, Kumar BN. hepatoprotective activity of whole part of the plant *Cuscuta reflexa* Roxb.(Convolvulaceae) in chloroform, ethanol and paracetamol induced hepatotoxic rat models. *Int J Pharm Clin Res* 2014; 6:127-132.
  243. Taghizadieh M, Issabeagloo E, Valiloo MR, Afshari F, Asadi J. Hepatoprotective and antioxidant activity of ethanolic extract of aerial parts of *Cuscuta reflexa* Roxb. on liver damage due to cisplatin in rats. *Baltica* 2014; 27:274-279.
  244. Shaw JE, Sicree RA, Zimmet PZ. Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Res Clin Pract* 2010; 87:4-14.
  245. Rath D, Kar DM, Panigrahi SK, Maharana L. Antidiabetic effects of *Cuscuta reflexa* Roxb. in streptozotocin induced diabetic rats. *J Ethnopharmacol* 2016; 192:442-449.
  246. Ma JZ, Yang LX, Shen XL, Qin JH, Deng LL, Ahmed S, et al. Effects of traditional Chinese medicinal plants on anti-insulin resistance bioactivity of DXMS-induced insulin resistant HepG2 cells. *Nat Prod Bioprospect* 2014; 4:197-206.
  247. Li DZ, Peng DY, Zhang R, Xu XX. Effects of *Cuscuta chinensis* polysaccharide on diabetic mice by alloxan. *Anhui Med Pharm J* 2008; 12:900-911.
  248. Li XJ, You HY, Yang J, Liu BM, You G, Song Y. Aqueous extracts of *Cuscuta chinensis* Lam induces differentiat ion of amelanotic melanocytes of human hair follicles. *Chin J Dermatovenereol* 2008; 22:13-15.
  249. Xu XX, Li DZ, Peng DY, Zhang R. Effects of *Cuscuta chinensis* Polysaccharide on Glucose-lipid Metabolism in Diabetic Rats. *Chin J Exp Tradit Med Formula* 2011; 17:232-234.
  250. Lei X, He J, Ren C, Zhou Y, Chen X, Dou J. Protective effects of the Chinese herbal medicine prescription Zhujing pill on retina of streptozotocin-induced diabetic rats. *Biomed Pharmacother* 2018; 98:643-650.
  251. Al-Sultany, Fadia H, Al-Saadi AH, Al-Husainy IM. Evaluated the Up-regulation in gene expression of hepatic insulin gene and hepatic insulin receptor gene in type 1 diabetic rats treated with *Cuscuta chinesis* Lam. *J Babylon Univ* 2018; 26:75-93.
  252. Trinchieri G. Cancer and inflammation: an old intuition with rapidly evolving new concepts. *Annu Rev Immunol* 2012; 30:677-706.
  253. Suresh V, Sruthi V, Padmaja B, Asha VV. *In vitro* anti-inflammatory and anti-cancer activities of *Cuscuta reflexa* Roxb. *J Ethnopharmacol* 2011; 134:872-877.
  254. Katiyar NS, Singh AP, Gangwar AK, Rao NV. Evaluation of carrageenan induced antiinflammatory activity of stem extracts of *Cuscuta reflexa* (Roxb) in rats. *Int J Res Pharm Chem*

- 2015; 5:322-326.
255. Kang SY, Jung HW, Lee MY, Lee HW, Chae SW, Park YK. Effect of the semen extract of *Cuscuta chinensis* on inflammatory responses in LPS-stimulated BV-2 microglia. *Chin J Nat Med* 2014; 12:573-581.
256. Liao JC, Chang WT, Lee MS, Chiu YJ, Chao WK, Lin YC, et al. Antinociceptive and anti-inflammatory activities of *Cuscuta chinensis* seeds in mice. *The Am J Chin Med* 2014; 42:223-242.
257. Koca U, Küpeli-Akkol E, Sekeroglu N. Evaluation of *in vivo* and *in vitro* biological activities of different extracts of *Cuscuta arvensis*. *Nat Prod Commun* 2011; 6:1433-1436.
258. Islam R, Rahman MS, Rahman SM. GC-MS analysis and antibacterial activity of *Cuscuta reflexa* against bacterial pathogens. *Asian Pac J Trop Dis* 2015; 5:399-403.
259. Pal DK, Mandal M, Senthilkumar GP, Padihari A. Antibacterial activity of *Cuscuta reflexa* stem and *Corchorus olerius* seed. *Fitoterapia* 2006; 77:589-591.
260. Bibi Y, Naeem J, Zahara K, Arshad M, Qayyum A. *In Vitro* antimicrobial assessment of selected plant extracts from pakistan. *Iran J Sci Technol A* 2018; 42:267-272.
261. Okiei W, Ogunlesi M, Ademoye MA. An assessment of the antimicrobial properties of extracts of various polarities from *Chasmanthera dependens*, *Emilia coccinea* and *Cuscuta australis*, herbal medications for eye diseases. *J Appl Sci* 2009; 9:4076-4080.
262. Bonjar S. Evaluation of antibacterial properties of some medicinal plants used in iran. *J Ethnopharmacol* 2004; 94:301-305.
263. Abdullah JA, Hammadi AA, Hakem R, Hatf Z, Hussein N. Study effect of plant extraction for *Cuscuta europaea* (Dodder) against two species of bacteria *Staphylococcus aureus* and *Escherichia coli*. *J Contemp Med Sci* 2016; 2:133-137.
264. Etedali P, Behbahani M, Rahiminejad RM, Rad SJ. Effect of crude extracts and fractions of *Cuscuta campestris* and two different hosts on peripheral blood mononuclear cells and HIV replication. *Int J Biosci* 2014; 4:83-89.
265. Ahmed HM, Yeh JY, Tang YC, Cheng WT, Ou BR. Molecular screening of chinese medicinal plants for progestogenic and anti-progestogenic activity. *J Biosci* 2014; 39:453-461.
266. Alaoui-Jamali M, editor. *Alternative and complementary therapies for cancer: Integrative approaches and discovery of conventional drugs*. Springer Science Business Media; New York, USA, 2010. p.63.
267. Zeraati F, Zamani A, Goodarzi MT, Hashjin SM, Razzaghi K. *In vitro* cytotoxic effects of *Cuscuta chinensis* whole extract on human acute lymphoblastic leukemia cell line. *Iran J Med Sci* 2015; 35:310-314.
268. Choi EJ, Kim GH, Kim T. Equol induced the apoptosis via cell cycle arrest in MDA-MB-453 but not in MCF-7 cells. *Faseb J* 2008; 22:265-265.
269. Magee PJ, Raschke M, Steiner C, Duffin JG, Pool-Zobel BL, Jokela T, et al. Equol: a comparison of the effects of the racemic compound with that of the purified S-enantiomer on the growth, invasion, and DNA integrity of breast and prostate cells *in vitro*. *Nutr Cancer* 2006; 54:232-242.
270. Selvi EK, Turumtay H, Demir A, Turumtay EA. Phytochemical profiling and evaluation of the hepatoprotective effect of *Cuscuta campestris* by high-performance liquid chromatography with diode array detection. *Anal Lett* 2018; 51:1464-1478.
271. Behbahani M. Evaluation of *in vitro* anticancer activity of *Ocimum basilicum*, *Alhagi maurorum*, *Calendula officinalis* and their parasite *Cuscuta campestris*. *PloS one* 2014; 9:1-13.
272. Pan HJ, Sun HX, Pan YJ. Adjuvant effect of ethanol extract of *semen cuscutae* on the immune responses to ovalbumin in mice. *J Ethnopharmacol* 2005; 99:99-103.
273. Lin MK, Yu YL, Chen KC, Chang WT, Lee MS, Yang MJ, et al. Kaempferol from *semen cuscutae* attenuates the immune function of dendritic cells. *Immunobiology* 2011; 216:1103-1109.
274. Lin HB, Lin JQ, Lu N, Yi XY. Comparative study on immune enhancement effects of four kinds of dodder seeds in shandong province. *J Chin Integr Med* 2003; 1:51-53.
275. Xiao J, Cui F, Ning T, Zhao W. Effects of alcohol extract from *Polygonatum odoratum* and *Cuscuta australis* on immunological function of mice injured by burns. *Chin J Chin Mater Med* 1990; 15:557-559.
276. Gu LG, Ye M, Yan YN, Jia L, Zhao JQ. Study of *Cuscuta australis* hyperoside on immunological function of mice *in vivo* and *in vitro*. *Chin J Tradit Chin Med Inf* 2001; 8:42-44.
277. Jian-Hui L, Bo J, Yong-Ming B, Li-Jia A. Effect of *Cuscuta chinensis* glycoside on the neuronal differentiation of rat pheochromocytoma PC12 cells. *Int J Dev Neurosci* 2003; 21:277-281.
278. Liu ZY, Yang YG, Zheng B. Effect of improving memory and inhibiting acetylcholinesterase activity by invigorating-qi and warming-yang recipe. *Chin J Integr Tradit West Med* 1993; 13:675-676.
279. Yu Q, Song FJ, Chen JF, Dong X, Jiang Y, Zeng KW, et al. antineuroinflammatory effects of modified wu-zi-yan-zong prescription in  $\beta$ -amyloid-stimulated BV2 microglia via the NF- $\kappa$ B and ERK/p38 MAPK signaling pathways. *J Evid Based Complementary Altern Med* 2017; 2017:1-10.
280. Cai XG, Xu AX, Ge B, Gao X, Yang SH. Effects of a polysaccharide from CCL on inhibiting oxygen free radical threshold of senile mice model. *Acta Acad Med Mil Tertiae* 2005; 27:1326-1328.
281. Li CS, Deng HB, Li DD, Li ZH. Advances and challenges in screening traditional chinese anti-aging materia medica. *Chin J Integr Med* 2013; 19:243-252.
282. Yang FY, Huang J. "Tai Ping Sheng Hui Fang" in the anti-aging effects medical research. *J Guiyang Coll of Tradit Chin Med* 1998; 2:7-8.
283. Gilani AU, Aftab K. Pharmacological actions of *Cuscuta reflexa*. *Int J Pharma* 1992; 30:296-302.
284. Yao CH, Tsai CC, Chen YS, Chang CJ, Liu BS, Lin CC, et al. Fabrication and evaluation of a new composite composed of tricalcium phosphate, gelatin and Chi-Li-Saan as a bone substitute. *Am J Chin Med* 2002; 30:471-482.
285. Yang HM, Shin HK, Kang YH, Kim JK. *Cuscuta chinensis* extract promotes osteoblast differentiation and mineralization in human osteoblast-like MG-63 cells. *J Med Food* 2009; 12:85-92.
286. Yang M, Sun J, Lu Z, Chen G, Guan S, Liu X, et al. Phytochemical analysis of traditional chinese med using liquid chromatography coupled with mass spectrometry. *J Chromatogr A* 2009; 1216:2045-2062.
287. Shin S, Lee YJ, Kim EJ, Lee AS, Kang DG, Lee HS. Effect of *Cuscuta chinensis* on renal function in ischemia/reperfusion-induced acute renal failure rats. *Am J Chin Med* 2011; 39:889-902.
288. Sun K, Zhao C, Chen XF, Kim HK, Choi BR, Huang YR, et al. *Ex vivo* relaxation effect of *Cuscuta chinensis* extract on rabbit corpus cavernosum. *Asian J Androl* 2013; 15:134-137.
289. Peng SJ, Lu RK, Yu LH. Effect of semen cucutae, rhizoma curculiginis, radix morindae, officinalis on human spermatozoa's motility and membrane function *in vitro*. *Chin J West Med* 1997; 17:145-147.
290. Shah GR, Chaudhari MV, Patankar SB, Pensalwar SV, Sabale VP, Sonawane NA. Evaluation of a multi-herb supplement for

- erectile dysfunction: a randomized double-blind, placebo-controlled study. *BMC Complementary Altern Med* 2012; 12:155-163.
291. Linmao Y. Integrating chinese and western medicine to treat infantile uterus. *Herb J Tradit Chin Med* 1992; 14:40-54.
292. Jang H, Bae WJ, Kim SJ, Cho HJ, Yuk SM, Han DS, et al. The herbal formula KH-204 is protective against erectile dysfunction by minimizing oxidative stress and improving lipid profiles in a rat model of erectile dysfunction induced by hypercholesterolaemia. *BMC Complementary Altern Med* 2017; 17:129-140.
293. Yang J, Wang Y, Bao Y, Guo J. The total flavones from semen *cuscutae* reverse the reduction of testosterone level and the expression of androgen receptor gene in kidney-yang deficient mice. *J Ethnopharmacol* 2008; 119:166-171.
294. Wang J, Wang M, Ou Y, Wu Q. Effects of flavonoids from semen *cuscutae* on changes of beta-EP in hypothalamuses and FSH and LH in anterior pituitaries in female rats exposed to psychologic stress. *J Chin Med Mater* 2002; 25:886-888.
295. Ma HX, You ZL, Wang RG. Effect of total flavones from *Cuscuta chinensis* on expression of Th type-1/Th type-2 cytokines, serum P and PR in abortion rats model. *J Chin Med Mater* 2008; 31:1201-1204.
296. Ma HX, You ZL, Wang XY. Effect of total flavones from *Cuscuta chinensis* on expression of Fas/FasL, PCNA and HB-EGF in SD rats model with bromocriptine-induced abortion. *J Chin Med Mater* 2008; 31:1706-1709.
297. Zhu JF, She YC, Zhou CH. Experimental and clinical studies on the effect of Shou Tai Wan and additives on threatened abortion. *J Integr Tradit Western Med* 1987; 7:407-409.
298. Aqil F, Zahin M, Ahmad I. Antimutagenic activity of methanolic extracts of four ayurvedic medicinal plants. *Indian J Exp Biol* 2008; 46:668-672.
299. Nakahara K, Trakoontivakorn G, Alzoreky NS, Ono H, Onishi-Kameyama M, Yoshida M. Antimutagenicity of some edible Thai plants, and a bioactive carbazole alkaloid, mahanine, isolated from *Micromelum minutum*. *J Agric Food Chem* 2002; 50:4796-4802.
300. Zweier JL, Talukder MH. The role of oxidants and free radicals in reperfusion injury. *Cardiovasc Res* 2006; 70:181-190.
301. Veinot JP, Gattinger DA, Fliss H. Early apoptosis in human myocardial infarcts. *Hu Pathol* 1997; 28:485-492.
302. Rabkin SW. Apoptosis in human acute myocardial infarction: the rationale for clinical trials of apoptosis inhibition in acute myocardial infarction. *Sch Res Exch* 2009; 2009:1-10.
303. Sun SL, Guo L, Ren YC, Wang B, Li RH, Qi YS, et al. Anti-apoptosis effect of polysaccharide isolated from the seeds of *Cuscuta chinensis* lam on cardiomyocytes in aging rats. *Mol Biol Rep* 2014; 41:6117-6124.
304. Zhongrong L, Pengtie L, Tiejun F, Yuanqiao J, Ruozhu W. The effect of three extraction technique of Chinese dodder seed on cardiovascular activity. *Nat Prod Res Develop* 2004; 16:532-533.
305. Thomas S, Shrikumar S, Velmurugan C, Kumar BA. Evaluation of anxiolytic effect of whole plant of *Cuscuta reflexa*. *World J Pharm Sci* 2015; 4:1245-1253.
306. Mokhtarifar N, Sharif B, Naderi N, Mosaddegh M, Faizi M. Evaluation of anti-depressant effects of *Cuscuta chinensis* in experimental models. *Res Pharm Sci* 2012; 7:826-827.
307. Cheng D, Murtaza G, Ma S, Li L, Li X, Tian F, et al. In silico prediction of the anti-depression mechanism of a herbal formula (tiansi liquid) containing *Morinda officinalis* and *Cuscuta chinensis*. *Molecules* 2017; 22:1614-1630.
308. Firoozabadi A, Zarshenas MM, Salehi A, Jahanbin S, Mohagheghzadeh A. Effectiveness of *Cuscuta planiflora* ten and *Nepeta menthoides* Boiss & Buhse in major depression: a triple-blind randomized controlled trial study. *J Evi based Complementary Altern Med* 2015; 20:94-97.
309. Shen L, Huang YY, Wang XN, Du J, Wang YY, Zhang DQ. Pharmacological effect of *cuscutae Semen* by external use on experimental vitiligo in guinea pigs. *Chin J Exp Tradit Med Formulae* 2012; 18:199-202.
310. Wang TJ, An J, Chen XH, Deng QD, Yang L. Assessment of *Cuscuta chinensis* seeds: effect on melanogenesis: comparison of water and ethanol fractions *in vitro* and *in vivo*. *J Ethnopharmacol* 2014; 154:240-248.
311. Mojtabae M, Mokaberinejad R, Hamzeloo-Moghadam M, Nasab MR, Adhami S, Farshi S, et al. The effect of the traditional medicine product "Milk-Cuscuta" on skin hyperpigmentation in patients with melasma. *Middle East J Family Med* 2018; 7:204-211.
312. Suk KD, Lee SJ, Bae JM. Inhibitory effects of *Cuscuta japonica* extract and *C. australis* extract on mushroom tyrosinase activity. *Korean J Pharma* 2004; 35:380-383.
313. Patel S, Sharma V, Chauhan NS, Dixit VK. A study on the extracts of *Cuscuta reflexa* Roxb in treatment of cyclophosphamide induced alopecia. *Daru J Pharm Sci* 2014; 22:27-34.
314. Huang AG, Yi YL, Ling F, Lu L, Zhang QZ, Wang GX. Screening of plant extracts for anthelmintic activity against *Dactylogyrus intermedius* (Monogenea) in goldfish (*Carassius auratus*). *Parasitol Res* 2013; 112:4065-4072.
315. Moon M, Jeong HU, Choi JG, Jeon SG, Song EJ, Hong SP, et al. Memory-enhancing effects of *Cuscuta japonica* Choisy via enhancement of adult hippocampal neurogenesis in mice. *Behav Brain Res* 2016; 311:173-182.
316. Mehrabani M, Modirian E, Ebrahimabadi AR, Vafazadeh J. Study of the effects of hydro-methanol extracts of *Lavandula vera* DC and *Cuscuta epithimum* Murr on the seizure induced by pentylentetranzol in mice. *J Kerman Univ Med Sci* 2014; 14:25-32.
317. Gupta MA, Mazumder UK, Pal D, Bhattacharya S, Chakrabarty SU. Studies on brain biogenic amines in methanolic extract of *Cuscuta reflexa* Roxb and *Corchorus olitorius* Linn seed treated mice. *Acta Pol Pharma* 2003; 60:207-210.
318. Ohta Y, Sami M, Kanda T, Saito K, Osada K, Kato H. Gene expression analysis of the anti-obesity effect by apple polyphenols in rats fed a high fat diet or a normal diet. *J Oleo Sci* 2006; 55:305-314.
319. Kakhrova KA, Khashimova ZS, Terenteva EO. Studies on cytotoxicity and antioxidant activities of lectin-like proteins from phytoparasites (*Cuscuta eurospaea*). *Asian Pharm Pharmacol* 2018; 4:265-270.
320. Bhan S, Mohan L, Srivastava CN. Efficacy of *Cuscuta reflexa* extract and its synergistic activity with Temephos against mosquito larvae. *Int J Mosquito Res* 2015; 2:34-41.
321. Alamgeer, Niazi SG, Uttra AM, Qaiser MN, Ahsan H. Appraisal of anti-arthritis and nephroprotective potential of *Cuscuta reflexa*. *Pharm Biol* 2017; 55:792-798.
322. Gautam T, Thakur V, Gupta V. Phytochemical Screening and Wound Healing Potential of *Cuscuta reflexa* Roxb. *Int J Pharm Life Sci* 2018; 9:21-21.
323. Sakib MH, Hossain MS, Hossain MS, Al Mahmood A, Sarkar MY, Rahman S, Shill LK. *In-vitro* cytotoxicity and antioxidant property evaluation from methanolic extract of *Cuscuta Reflexa* flowers. *Asian J Med Biol Res* 2015; 1:285-291.
324. Manirujjaman M, Suchana S, Collet T, Nawshin LN, Chowdhury MA. Antimicrobial effects of ethanolic extracts

from *Cuscuta reflexa* Roxb (Convolvulaceae). Int J Pharmacogn Phytochem Res 2016; 8:930-932.

325. Praseeja RJ, Sreejith PS, Asha VV. Studies on the apoptosis inducing and cell cycle regulatory effect of *Cuscuta reflexa* Roxb chloroform extract on human hepatocellular carcinoma cell line, Hep 3B. Int J Appl Res Nat Prod 2015; 8:37-47.

326. Roohina Ali S, Haque S, Versiani MA, Faizi S, Farooq

AD. Cytotoxicity and chromosomal aberrations induced by methanolic extract of *Cuscuta reflexa* and its pure compounds on meristematic cells of *Allium* species. Pak J Pharm sci 2017; 30:521-529.

327. Mala FA, Sofi MA. Evaluation of antihistaminic Activity of herbal drug isolated from *Cuscuta reflexa* Roxb. Ann Plant Sci 2017; 6:1807-1810.