Iranian Journal of Basic Medical Sciences

ijbms.mums.ac.ir

Effect of myricetin on the gene expressions of NOX3, TGF- β 1, prestin, and HSP-70 and anti-oxidant activity in the cochlea of noise-exposed rats

Maryam Bahaloo¹, Mohammad Ebrahim Rezvani², Ehsan Farashahi Yazd³, Fatemeh Zare Mehrjerdi², Mohammad Hossein Davari⁴, Ali Roohbakhsh⁵, Abolfazl Mollasadeghi¹, Haniyeh Nikkhah³, Maryam Vafaei ³, Amir Houshang Mehrparvar ^{1*}

¹ Industrial Diseases Research Center, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

² Department of Physiology, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

³ Stem Cell Biology Research Center, Yazd Reproductive Sciences Institute, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

⁴ Department of Occupational Medicine, Shahid Sadoughi University of Medical Sciences, Yazd, Iran
⁵ Pharmaceutical Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran

ARTICLEINFO	ABSTRACT	
<i>Article type:</i> Original article	Objective(s): Noise-induced hearing loss is one of the most common occupational diseases in industrialized countries and can be affected by various environmental and genetic factors. This study	
<i>Article history:</i> Received: Jun 9, 2019 Accepted: Nov 26, 2019	was designed to examine the effect of myricetin in preventing this disorder. <i>Materials and Methods:</i> Twenty-one Wistar rats were randomly divided into five groups: Non-exposed, noise exposure only, noise exposure with vehicle, noise exposure with myricetin 5 mg/Kg, and noise exposure with myricetin 10 mg/kg.	
<i>Keywords:</i> Anti-oxidants Gene expression Myricetin NADPH oxidase Noise-induced hearing loss Superoxide dismutase	noise exposure with myricetin 10 mg/kg. All animals were sacrificed after last noise exposure. The left cochlea was dissected from each rat. It was used for mRNA expression analysis (NOX3, TGF-β1, prestin, and HSP-70). Blood samples were collected to assess superoxide dismutase (SOD) activity, 1, 1 diphenyl picrylhydrazyl (DPPH), and malondialdehyde (MDA) measurements. <i>Results:</i> Real time-PCR assay revealed that noise decreased NOX3 and increased TGF-β1, prestin, and HSP-70 gene expressions. Administration of myricetin at the dose of 5 mg/kg, but not at 10 mg/kg, significantly reversed these changes. Noise also increased MDA levels and decreased SOD and DPPH scavenging activities. Myricetin at the doses of 5 and 10 mg/kg also reversed these changes. <i>Conclusion:</i> The findings of this study showed that myricetin at the dose of 5 mg/Kg was able to reverse noise-induced abnormalities in gene expression and oxidant/anti-oxidant balance. It is a possibility that myricetin via enhancement of anti-oxidant activity induced these effects.	

Please cite this article as:

Bahaloo M, Rezvani MH, Farashahi Yazd E, Zare Mehrjerdi F, Davari MH, Roohbakhsh A, Mollasadeghi A, Nikkhah H, Vafaei M, Mehrparvar AH. Effect of myricetin on the gene expressions of NOX3, TGF-β1, prestin, and HSP-70 and anti-oxidant activity in the cochlea of noise-exposed rats. Iran J Basic Med Sci 2020; 23:594-599. doi: 10.22038/IJBMS.2020.41007.9693

Introduction

Exposure to noise intensities higher than the permissible level, is among the most common factors with damaging effects on hearing (1). Ears cannot tolerate frequent or prolonged exposure to noise, and exposure to high levels of noise can damage sensory hair cells in the cochlea, inducing noise-induced hearing loss (NIHL) (2). NIHL has been reported as one of the most common occupational diseases in industrialized countries.

Different mechanisms, such as mechanical stress, oxidative stress, and free radical production have been proposed as the cause of NIHL. Recently, in order to find a practical method to prevent NIHL, scientists have paid attention to the role of genetic susceptibilities and application of different chemicals, especially antioxidants, for the prevention of NIHL (3).

Malondialdehyde (MDA) is the final product of lipid peroxidation which is used for the evaluation of oxidative stress (4). Noise exposure as an inducer of oxidative stress, increases MDA levels (5). To counteract these conditions, the enzymatic defense system will be activated. One of these defending enzymes is

superoxide dismutase (SOD), which protects cells from the harmful effects of superoxide anions (6). Another factor for assessment of anti-oxidant activity of natural compounds is 1,1 diphenyl picrylhydrazyl (DPPH) (7).

IJ MS

Among genes that are considered to be related to NIHL, NOX3 gene is one of the most important genes. This gene is one isomere of the NADPH (nicotinamide adenine dinocoids phosphate) oxidase enzyme family which includes seven enzymes producing superoxides: NOX1, NOX2, NOX3, NOX4, NOX5, DOUX1, and DOUX2 (8). It was reported that expression of NOX3 was reduced after noise exposure. So, it has been suggested as an endogenous protective factor against reactive oxygen species (ROS) generation, as a mechanism of NIHL (9).

Transforming growth factor $\beta 1$ (TGF- $\beta 1$) is a multifunctional peptide with unclear effects on the cochlea. Over-expression of TGF- β 1 in the inner ear has been proposed as a factor of susceptibility to otosclerosis and NIHL (10).

Noise exposure can increase the expression of prestin mRNA in rats and guinea pigs (11), which may damage proteins and decrease the electromotility of outer

*Corresponding author: Amir Houshang Mehrparvar. Occupational medicine department, Shahid Rahnemoun hospital, Farrokhi ave., Yazd, Iran. Tel/Fax: +98-3537247111: Email: ah.mehrparvar@ssu.ac.ir

hair cells (OHC) (12). Heat shock proteins (HSP) are a group of proteins that protect inner ear against acoustic trauma and ototoxic agents (13).

Considering that one of the mechanisms of NIHL occurrence is oxidative stress and increased free radical production, the use of anti-oxidants by reducing ROS will be an effective step in the prevention of NIHL (14). Resveratrol (15), vitamins E and C, and magnesium (16), N-acetyl cysteine (17), statins (18), and ascorbic acid (19) are examples of these anti-oxidants used in previous studies.

Myricetin belongs to the polyphenols family and is found in berries, grapes, tea, fruits, and some medicinal plants (20). It is a potent anti-inflammatory and anti-oxidant agent with preventive effects on neurodegenerative and inflammatory diseases (21).

This study was designed to examine the effect of myricetin on NOX3, TGF- β 1, prestin, and HSP-70 genes expression and SOD and DPPH scavenging activities in rats as factors that may prevent NIHL.

Materials and Methods

Animals

The protocol used in this study was approved by the ethics committee of Shahid Sadoughi University of Medical Sciences. We tried to use the minimum number of animals and minimize their suffering during the study. Animals included 2 month-old male Wistar rats (weighing 200–250 g at study onset) obtained from animal house of Shahid Sadoughi University of Medical Sciences. Animals were housed in standard plexi-glass cages with controlled temperature (22–23 °C) and humidity (60±5%), under a 12 hr light/dark cycle with food and water available.

Twenty-one Wistar rats were randomly divided into the following groups:

1. Control group: Non-exposed (n=3);

2. Noise group: Noise exposure only (n=6);

3. Noise+Veh group Noise exposure with vehicle (n=4);

4. Noise+Myrc5 group: Noise exposure with myricetin (Sigma-Aldrich GmbH) 5 mg/kg, (n=4);

5. Noise+Myrc10 group: Noise exposure with myricetin

10 mg/kg (n=4).

Myricetin and vehicle were administered from 3 days before exposure to noise and continued for 13 days. Myricetin was injected IP 1 hr before exposure to noise.

Noise exposure

Animals were exposed to a 10 KHz octave band noise with the intensity of 100 dB sound pressure level (SPL) for 1 hr each day for 10 consecutive days (22) in a plexiglass box. Noise was generated by an audiometer (OB 929, Madsen, Denmark). Sound level was measured using a sound level meter (TES-1351 digital sound level meter, Taiwan).

Tissue preparation

All Animals were sacrificed after the last noise exposure. The left cochlea was dissected from each rat and used to determine the gene expression levels and assessment of oxidant biomarkers.

Gene expression assessment

In order to determine the effect of different doses of myricetin on noise-exposed cochlea, mRNA expressions of NOX3, TGF-B1, prestin, and HSP-70 genes in the cochlea were assessed by semi-quantitative real-time polymerase chain reaction (PCR) (Qiagen, Germany). Total RNA was isolated using the RNX Plus Kit (Cinnaclon, Iran). cDNA was synthesized using a cDNA synthesis kit (Thermo Fisher Scientific, Lithuania). Real time PCR products were detected using SYBR Green and were normalized by β -Actin expression. The PCR cycling conditions included one cycle with 95 °C for 10 min (denaturation I) then 40 cycles in the following steps: 95 °C for 15 sec (denaturation II), 60 °C for 30 sec (annealing), and 72 °C for 30 sec (extension). Primers were designed using the Beacon designer software package (Premier Biosoft) and synthesized by Integrated DNA Technologies (BVBA, Leuven, Belgium). The primer sequences are presented in Table 1.

Biochemical analysis

Blood samples were collected for measuring the

Table 1. The primer sequences of different genes for real-time PCR analysis

Gene	Primers sequence $(5 \longrightarrow 3)$	Size (bp)	Annealing °C
Rat NOX3 (9)	F: ATCTTTATCCAGTGCCCATCC	<u>129</u>	<u>59</u>
	R: CTTCAGTAACGCCTCTGTCCA		
Rat TGFβ1 (23)	F: GTCAACTGTGGAGCAACACG	<u>116</u>	<u>61</u>
	R: CGTCAAAAGACAGCCACTCA		
Rat Prestin (24)	F: CACAGAGTCCGAGCTACACAGTC	<u>159</u>	<u>60</u>
	R: TCAGTGCGCTGCTGTACAAG		
Rat HSP-70 (25)	F: CTCGTGCGTGGGCGTGTTCC	285	<u>61</u>
	R: TTTCCCTTGTAGTTCACCTG		
Ratβ-Actin (23)	F: CTATCGGCAATGAGCGGTTCC	<u>146</u>	<u>61</u>
	R: TGTGTTGGCATAGAGGTCTTTACG		

The relative gene expression was measured by the $2-\Delta\Delta Ct$ method and then normalized to the expression levels of endogenous β -Actin gene

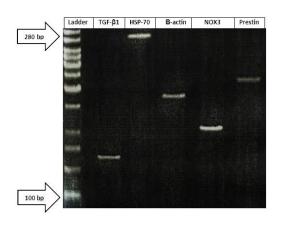


Figure 1. The polymerase chain reaction (PCR) reactions for non-template control were demonstrated using the agarose gel electrophoresis (10bp DNA ladder and 8% agarose gel). As shown, the individual amplifications matched the predicted sizes

serum levels of MDA and DPPH and activity of SOD. The samples were centrifuged at 3500 rpm for 15 min to separate the serum, and the serum was then stored at -80 $^{\circ}$ C until the time of analysis. SOD activity and blood concentration of MDA were measured with a commercial assay kit (Zell Bio, Germany) according to the manufacturer's instructions.

Anti-oxidant activity assay for DPPH

To determine the ability of myricetin to scavenge DPPH radicals, freshly prepared methanol DPPH (0.1 mM/ml) solution was added to different concentrations of myricetin. After half an hour, the absorbance ability was recorded at 517 nm. Results were expressed as percentage of inhibition (Inhibition (%) = [(A Control – A Extract) / A Control] ×100)

A blank represents the measured absorbance of the sample containing control reaction (all reagents without the test compound). A sample represents the measured absorbance of the test compound (26).

Statistical analysis

The data were expressed as mean±SEM (standard error of mean) and analyzed using graphPad prism 7.0 (GraphPad, Inc., USA) statistical software. Kruskal-Wallis test was used to determine the significant differences between groups and was followed by Tukey's test for *post-hoc* comparisons. *P*-value<0.05 was considered as statistical significance.

Results

Real time-PCR assay revealed that NOX3 expression was significantly decreased in noise and vehicle groups; however in Myrc 5 mg group, it was significantly increased. Expressions of TGF- β 1, prestin, and HSP-70 genes were significantly increased in the noise group and was decreased in Myrc groups, but the difference was significant only for Myrc 5 mg group. Figure 1 compares the effect of myricetin on NOX3, TGF β 1, prestin, and HSP-70 gene expression in different study groups.

Figure 2 compares the SOD activity, DDPH scavenging activity, and MDA level in different groups.

Figure 3 compare the plasma MDA level, SOD activity, and DPPH scavenging activity in different groups.

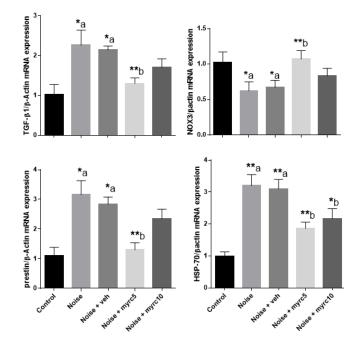


Figure 2. Effect of myricetin on relative gene expression of TGF- β 1, NOX3, prestin, and HSP-70. Beta actin was used for housekeeping gene. One-way ANOVA were used for statistical comparisons. Data was shown as means±SEM. **P*<0.05 and ***P*<0.01. "a" and "b" when compared to Control or Noise group respectively

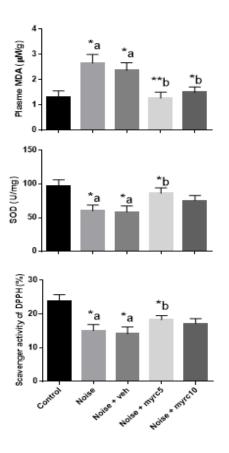


Figure 3. Effect of myricetin on plasma Malondialdehyde (MDA) level, superoxide dismutase (SOD) activity, and 1,1 diphenyl picrylhydrazyl (DPPH) scavenging activity in different groups. One-way ANOVA was used for statistical comparisons. Data was shown as means±SEM. **P*<0.05 and ***P*<0.01. "a" and "b" when compared to Control or Noise group, respectively

Discussion

NIHL is a complex disease that is affected by personal, environmental and genetic backgrounds. Workers with similar exposure to noise may show different levels of hearing damage, which proposes a genetic predisposing role (27).

Exposure to noise induces production of ROS and may lead to hearing loss (28). Present evidence suggests that anti-oxidants can prevent NIHL (29). Myricetin is an anti-oxidant and anti-inflammatory flavonoid that has ability to scavenge ROS and shows anti-oxidant activity (21). In the current study, we assessed the anti-oxidant effects of myricetin on prevention of NIHL.

Previous studies have suggested that the NADPH family can be the source of ROS production and induction of apoptosis in various cells (30, 31). NOX3 is expressed in the inner ear (32).

Our results showed that NOX3 gene expression was reduced following exposure to noise, and administration of myricetin enhanced its expression. In agreement with the present findings, a study by Vlajkovic et al. evaluated the role of the NADPH oxidase family and reported that during prolonged exposure to noise, the expression levels of NOX1 and NOX2 genes increased, but NOX3 gene expression decreased. This may be a putative mechanism against oxidative stress after exposure to noise in the cochlea (9). In a study that completely evaluated the role of NOX3 in the development of NIHL, it was shown that NOX3 deficiency or absence in the inner ear increased susceptibility to NIHL (8). However, during cisplatin-induced ototoxicity, NOX3 has been reported as an inducing factor for other genes that ultimately lead to cochlear damage (31, 33).

It has been reported that cochlear injury due to neomycin (34) and noise (35) elevated TGF- β 1 activation in the cochlea, so inhibition of TFG- β 1 may prevent cochlear damage due to exposure to noise or ototoxic agents. Losartan as a blocker of TFG- β 1 signaling prevented axonal sprouting in the cochlea after hearing loss (36). Similarly, in the present study, myricetin decreased TGF- β 1 expression. However, there are controversial reports regarding the effect of TGF- β 1 inhibition on the prevention of NIHL. Murillo *et al.* showed that TGF- β 1 inhibitor peptides, if used before exposure to noise, improved hearing threshold and degenerative changes due to noise exposure in a dose-dependent manner (10), but single dose of these chemicals did not prevent NIHL (35).

Prestin is a protein located in OHC lateral membrane and presents in the inner ear (11, 12). It acts as a motor protein and is necessary for OHC electromotility (37). Expression of prestin increases after exposure to noise and returns to its basal level after a few weeks, which may damage proteins and lead to cochlear damage (12). In the present study, exposure to noise increased the expression of the prestin gene, which was reversed in the myricetin-treated groups. It has been proposed as a mechanism to prevent NIHL (38).

Environmental stresses such as hyperthermia, ischemia, and exposure to noise can induce HSP-70 (39, 40). Administration of HSP-70 may protect inner hair cells from severe ototoxic injury (41). This study showed that noise significantly increased the expression of HSP-

70, but myricetin decreased its expression. This finding shows that anti-oxidant effect of myricetin decreased the oxidant stress and eventually the need for expression of HSP-70, which is a cytoprotective agent in the cochlea.

During oxidative stress, SOD activity is increased (42). SOD is an anti-oxidant enzyme in the mitochondria. It plays an important role in preventing noise-induced damage to the cochlea by ROS inhibition (43). It has been reported that lack of SOD makes young mice susceptible to NIHL (44). Exposure to noise in tile workers reduced SOD levels as well (45). In the current study, anti-oxidant effects of myricetin increased the activity of SOD in the Myrc 5 mg group. Other antioxidants, such as D-methionine (46), vitamin E (47), and Rehmannia Glutionosa, a traditional medicine (48), also increased the activity of erythrocyte SOD. In addition, myricetin increased the levels of anti-oxidant enzymes such as SOD and prevented the effects of 7.12 dimethyl benzanthracene (a breast cancer agent) on mammary tissue in female rats (49).

A study by Yildirim *et al.* showed that noise exposure in tile workers increased MDA levels due to oxidative stress in the cochlea (45). In a similar study, exposure to intense noise increased lipid peroxidation and MDA levels in the blood and administration of vitamin E decreased the MDA level (47). Similar to these studies, the present results showed that noise increased the blood level of MDA in rats, which was decreased in myrectin-treated animals.

DPPH assay is a common method for assessing free radical scavenging capacity in natural compounds. In DPPH assay, a natural compound transfers electron or hydrogen atoms and changes DPPH activity (50). According to this mechanism, we assessed DPPH free radical scavenging capacity of myricetin. DPPH free radical scavenging capacity decreased in noise group that was reversed by administration of low dose of myricetin. Scavenging effects of APP3 (a mushroom with anti-oxidant effects) was positively correlated with concentration and it inhibited the generation of hydroxyl radicals (50).

This study showed that administration of 5 mg/ Kg myricetin had a more favorable effect on NIHL prevention in genetic and biochemical levels than higher doses of this substance, which is probably due to systemic toxicity in higher doses. Similarly, N-acetyl cysteine also prevented NIHL at relatively low doses and resulted in systemic toxicity at higher doses (51).

Conclusion

The findings of this study showed that noise exposure increased the expression of TGF- β 1, prestin, and HSP-70 genes and MDA levels and decreased the expression of NOX3, SOD activity, and DPPH scavenging activity in rats, but administration of 5 mg/Kg myricetin in noise-exposed rats significantly reversed these effects. Therefore, myricetin at dose of 5 mg/Kg can probably be used as an anti-oxidant in prevention of NIHL.

Acknowledgment

The results presented in this paper were part of a PhD student thesis, funded by Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

Limitations

This study had some limitations: Due to monetary constraints, we couldn't perform Western blot, which is a more accurate method.

References

1. Carlsson PI, Van Laer L, Borg E, Bondeson ML, Thys M, Fransen E, *et al.* The influence of genetic variation in oxidative stress genes on human noise susceptibility. Hear Res 2005; 202:87-96.

2. Liberman MC. Noise-induced and age-related hearing loss: new perspectives and potential therapies. F1000Research 2017; 6:927-945.

3. Park J, Jou I, Park SM. Attenuation of noise-induced hearing loss using methylene blue. Cell death & dis 2014; 5:e1200.

4. Xiong M, Lai H, Yang C, Huang W, Wang J, Fu X, *et al.* Comparison of the protective effects of *Radix astragali*, α -lipoic acid, and vitamin E on acute acoustic trauma. Clin Med Insights Ear Nose Throat 2012; 29:25-31.

5. Dereköy FS, Dündar Y, Aslan R, Cangal AJEaoo-r-l. Influence of noise exposure on anti-oxidant system and TEOAEs in rabbits. Eur Arch Otorhinolaryngol 2001; 258:518-522.

6. Kasperczyk S, Dobrakowski M, Kasperczyk A, Machnik G, Birkner E. Effect of N-acetylcysteine administration on the expression and activities of anti-oxidant enzymes and the malondialdehyde level in the blood of lead-exposed workers. Environ Toxicol Pharmacol 2014; 37:638-647.

7. de Pinedo AT, Peñalver P, Pérez-Victoria I, Rondón D, Morales JCJFC. Synthesis of new phenolic fatty acid esters and their evaluation as lipophilic anti-oxidants in an oil matrix. Food Chem 2007; 105:657-665.

8. Lavinsky J, Crow AL, Pan C, Wang J, Aaron KA, Ho MK, *et al.* Genome-wide association study identifies nox3 as a critical gene for susceptibility to noise-induced hearing loss. PLoS Genet 2015; 11:e1005094.

9. Vlajkovic SM, Lin SCy, Wong ACY, Wackrow B, Thorne PRJHr. Noise-induced changes in expression levels of NADPH oxidases in the cochlea. Hear Res 2013; 304:145-152.

10. Murillo-Cuesta S, Rodríguez-de la Rosa L, Contreras J, Celaya AM, Camarero G, Rivera T, *et al.* Transforming growth factor β 1 inhibition protects from noise-induced hearing loss. Front Aging Neurosci 2015; 7:32-55.

11. Mazurek B, Haupt H, Amarjargal N, Yarin YM, Machulik A, Gross JJHr. Up-regulation of prestin mRNA expression in the organs of Corti of guinea pigs and rats following unilateral impulse noise exposure. Hear Res 2007; 231:73-83.

12. Chen G-DJHr. Prestin gene expression in the rat cochlea following intense noise exposure. Hear Res 2006; 222:54-61.

13. Taleb M, Brandon CS, Lee F-S, Harris KC, Dillmann WH, Cunningham LLJCS, *et al.* Hsp70 inhibits aminoglycoside-induced hearing loss and cochlear hair cell death. Cell Stress Chaperones 2009; 14:427-437.

14. Adelman C, Freeman S, Paz Z, Sohmer H. Salicylic acid injection before noise exposure reduces permanent threshold shift. Audiol Neurootol 2008; 13:266-272.

15. Seidman MD, Tang W, Bai VU, Ahmad N, Jiang H, Media J, *et al.* Resveratrol decreases noise-induced cyclooxygenase-2 expression in the rat cochlea. Otolaryngol Head Neck Surg 2013; 148:827-833.

16. Le Prell C, Johnson A-C, Lindblad A-C, Skjönsberg Å, Ulfendahl M, Guire K, *et al.* Increased vitamin plasma levels in Swedish military personnel treated with nutrients prior to

automatic weapon training. Noise Health 2011; 13:432-443. 17. Wu HP, Hsu CJ, Cheng TJ, Guo YL. N-acetylcysteine attenuates noise-induced permanent hearing loss in diabetic rats. Hear Res 2010; 267:71-77.

18. Jahani L, Mehrparvar A, Esmailidehaj M, Rezvani M, Moghbelolhossein B, Razmjooei Z. The effect of atorvastatin on preventing noise-induced hearing loss: an experimental study. Int J Occup Environ Med 2016; 7:615-621.

19. Loukzadeh Z, Hakimi A, Esmailidehaj M, Mehrparvar AH. Effect of Ascorbic Acid on Noise Induced Hearing Loss in Rats. Iran J Otorhinolaryngol 2015; 27:267-272.

20. Hou W, Hu S, Su Z, Wang Q, Meng G, Guo T, *et al*. Myricetin attenuates LPS-induced inflammation in RAW 264.7 macrophages and mouse models. Future Med Chem 2018; 10:2253-2264.

21. Domitrović R, Rashed K, Cvijanović O, Vladimir-Knežević S, Škoda M, Višnić AJC-bi. Myricitrin exhibits anti-oxidant, antiinflammatory and antifibrotic activity in carbon tetrachlorideintoxicated mice. Chem Biol Interact. 2015; 230:21-29.

22. Fetoni AR, De Bartolo P, Eramo SLM, Rolesi R, Paciello F, Bergamini C, *et al.* Noise-induced hearing loss (NIHL) as a target of oxidative stress-mediated damage: cochlear and cortical responses after an increase in anti-oxidant defense. J Neurosci 2013; 33:4011-4023.

23. Tang M, Zhou F, Zhang W, Guo Z, Shang Y, Lu H, *et al*. The role of thrombospondin-1-mediated TGF- β 1 on collagen type III synthesis induced by high glucose. Mol Cell Biochem 2011; 346:49-56.

24. Mazurek B, Haupt H, Joachim R, Klapp BF, Stöver T, Szczepek AJJHr. Stress induces transient auditory hypersensitivity in rats. Hear Res 2010; 259:55-63.

25. Kumei Y, Morita S, Shimokawa H, Ohya Ki, Akiyama H, Hirano M, *et al.* Inhibition of HSP70 and a collagen-specific molecular chaperone (HSP47) expression in rat osteoblasts by microgravity. Ann N Y Acad Sci 2003; 1010:476-480.

26. Megateli S, Krea MJP, plants mbo. Enhancement of total phenolic and flavonoids extraction from *Rosmarinus officinalis* L. using electromagnetic induction heating (EMIH) process. Physiol Mol Biol Plants 2018; 24:889-897.

27. Zhang X, Liu Y, Zhang L, Yang Z, Shao Y, Jiang C, *et al*. Genetic variations in protocadherin 15 and their interactions with noise exposure associated with noise-induced hearing loss in Chinese population. Environ Res 2014; 135:247-252.

28. Van Campen LE, Murphy WJ, Franks JR, Mathias PI, Toraason MA. Oxidative DNA damage is associated with intense noise exposure in the rat. Hear Res 2002; 164:29-38.

29. Ohinata Y, Miller JM, Schacht JJBr. Protection from noiseinduced lipid peroxidation and hair cell loss in the cochlea. Brain Res 2003; 966:265-273.

30. Rybak LP, Mukherjea D, Jajoo S, Kaur T, Ramkumar VJC, sciences ml. siRNA-mediated knock-down of NOX3: therapy for hearing loss. Cell Mol Life Sci 2012; 69:2429-2434.

31. Mukherjea D, Jajoo S, Whitworth C, Bunch JR, Turner JG, Rybak LP, *et al.* Short interfering RNA against transient receptor potential vanilloid 1 attenuates cisplatin-induced hearing loss in the rat. J Neurosci 2008; 28:13056-13065.

32. Sirokmány G, Donkó Á, Geiszt MJTips. Nox/Duox family of NADPH oxidases: lessons from knockout mouse models. Trends Pharmacol Sci 2016; 37:318-327.

33. Mukherjea D, Jajoo S, Sheehan K, Kaur T, Sheth S, Bunch J, *et al.* NOX3 NADPH oxidase couples transient receptor potential vanilloid 1 to signal transducer and activator of transcription 1-mediated inflammation and hearing loss. Antioxid Redox Signal 2011; 14:999-1010.

34. Wissel K, Wefstaedt P, Miller JM, Lenarz T, Stöver TJN. Differential brain-derived neurotrophic factor and transforming growth factor-beta expression in the rat cochlea

following deafness. Neuroreport 2006; 17:1297-1301.

35. Sanz L, Murillo-Cuesta S, Cobo P, Cediel R, Contreras J, Rivera T, *et al*. Swept-sine noise-induced damage as a hearing loss model for preclinical assays. Front Aging Neurosci 2015; 7:7-28.

36. Mun SK, Han KH, Baek JT, Ahn SW, Cho HS, Chang MYJC, *et al.* Losartan prevents maladaptive auditory-somatosensory plasticity after hearing loss via transforming growth factor- β signaling suppression. Clin Exp Otorhinolaryngol 2019; 12:33-39.

37. Geleoc GS, Holt JRJTin. Auditory amplification: outer hair cells pres the issue. Trends Neurosci 2003; 26:115-117.

38. Xia A, Song Y, Wang R, Gao SS, Clifton W, Raphael P, *et al.* Prestin regulation and function in residual outer hair cells after noise-induced hearing loss. PLoS One 2013; 8:e82602.

39. Gong T-W, Fairfield DA, Fullarton L, Dolan DF, Altschuler RA, Kohrman DC, *et al.* Induction of heat shock proteins by hyperthermia and noise overstimulation in Hsf1–/– mice. J Assoc Res Otolaryngol 2012; 13:29-37.

40. Du Y, Liu XH, Zhu HC, Wang L, Wang ZS, Ning JZ, Xiao CC. Hydrogen sulfide treatment protects against renal ischemiareperfusion injury via induction of heat shock proteins in rats. Iran J Basic Med Sci 2019; 22:99-112.

41. Takada Y, Takada T, Lee MY, Swiderski DL, Kabara LL, Dolan DF, *et al.* Ototoxicity-induced loss of hearing and inner hair cells is attenuated by HSP70 gene transfer. Mol Ther Methods Clin Dev 2015; 2:15019-15034.

42. Samson J, Wiktorek-Smagur A, Politanski P, Rajkowska E, Pawlaczyk-Luszczynska M, Dudarewicz A, *et al.* Noise-induced time-dependent changes in oxidative stress in the mouse cochlea and attenuation by D-methionine. Neuroscience 2008; 152:146-150.

43. Tuerdi A, Kinoshita M, Kamogashira T, Fujimoto C, Iwasaki S, Shimizu T, *et al*. Manganese superoxide dismutase influences

the extent of noise-induced hearing loss in mice. Neurosci Lett 2017; 642:123-128.

44. McFadden SL, Ohlemiller KK, Ding D, Shero M, Salvi RJ. The influence of superoxide dismutase and glutathione peroxidase deficiencies on noise induced hearing loss in mice. Noise Health 2001; 3:49-64.

45. Yildirim I, Kilinc M, Okur E, Tolun FI, Kilic MA, Kurutas EB, *et al.* The effects of noise on hearing and oxidative stress in textile workers. Ind Health 2007; 45:743-749.

46. Rewerska A, Pawelczyk M, Rajkowska E, Politanski P, Sliwinska-Kowalska M. Evaluating D-methionine dose to attenuate oxidative stress-mediated hearing loss following overexposure to noise. Eur Arch Otorhinolaryngol 2013; 270:1513-1520.

47. Kapoor N, Mani KV, Shyam R, Sharma RK, Singh AP, Selvamurthy WJN, *et al.* Effect of vitamin E supplementation on carbogen-induced amelioration of noise induced hearing loss in man. Noise Health 2011; 13:452-458.

48. Jayakumar J, Nirmala P, Kumar BP, Kumar APJSAjoc. Evaluation of protective effect of myricetin, a bioflavonoid in dimethyl benzanthracene-induced breast cancer in female Wistar rats. South Asian J Cancer 2014; 3:107-111.

49. Yu HH, Kim YH, Jung SY, Shin MK, Park RK, So HS, *et al.* Rehmannia glutinosa activates intracellular anti-oxidant enzyme systems in mouse auditory cells. Am J Chin Med 2006; 34:1083-1093.

50. Chen Y, Xue Y. Purification, chemical characterization and anti-oxidant activities of a novel polysaccharide from *Auricularia polytricha*. Int J Biol Macromol 2018; 120:1087-1092.

51. Lorito G, Giordano P, Prosser S, Martini A, Hatzopoulos S. Noise-induced hearing loss: a study on the pharmacological protection in the Sprague Dawley rat with N-acetyl-cysteine. Acta Otorhinolaryngol Ital 2006; 26:133-139.