

## Anti-Aging Effects of Some Selected Iranian Folk Medicinal Herbs-Biochemical Evidences

Azadeh Mohammadirad<sup>1</sup>, Fatemeh Aghamohammadali-Sarraf<sup>2</sup>, Simin Badiei<sup>2</sup>, Zakie Faraji<sup>2</sup>, Reza Hajiaghaee<sup>3</sup>, Maryam Baeri<sup>1</sup>, Mahdi Gholami<sup>1</sup>, Mohammad Abdollahi<sup>1\*</sup>

<sup>1</sup> Department of Toxicology and Pharmacology, Faculty of Pharmacy, and Pharmaceutical Sciences Research Center, Tehran University of Medical Sciences (TUMS), Tehran, Iran

<sup>2</sup> Pharmaceutical Sciences Branch, Islamic Azad University, Tehran, Iran

<sup>3</sup> Pharmacognosy & Pharmaceutics Department of Medicinal Plants Research Center, Institute of Medicinal Plants, ACECR, Karaj, Iran

### ARTICLE INFO

*Article type:*  
Original article

*Article history:*  
Received: May 25, 2013  
Accepted: Aug 26, 2013

*Keywords:*  
Aging  
D-galactose  
Herbal  
Mouse  
Oxidative stress

### ABSTRACT

**Objective(s):** In the current study, the effects of selected folk medicinal herbs were evaluated in D-galactose-induced aging in male mice.

**Materials and Methods:** Male BALB/c mice were randomly divided into 12 groups composing sham, control, and treated groups. Aging was induced by administration of D-galactose (500 mg/kg/day for 6 weeks). A positive control group was assigned that received vitamin E (200 mg/kg/day). The extract of herbs was prepared, lyophilized, and used in this study. The herbs were administered by gavage for 4 weeks to D-galactose-aged animals at the selected doses (mg/kg/day) as follows: *Zingiber officinale* (250), *Glycyrrhiza glabra* (150), *Rosmarinus officinalis* (300), *Peganum harmala* (50), *Aloe vera* (150), *Satureja hortensis* (200), *Teucrium scordium* (200), *Hypericum perforatum* (135) and *Silybum marianum* (150). One group of animals was assigned as sham and not given D-galactose.

**Results:** At the end of treatment, pro-inflammatory markers including tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukine-1 $\beta$  (IL- $\beta$ ), interleukine-6 (IL-6), NF-kappaB (NF-kb), total antioxidant power (TAP), thiobarbituric acid reactive substances (TBARS) as lipid peroxidation (LPO) marker and male sex hormones i.e. testosterone and dehydroepiandrosterone-sulfate (DHEA-S) were measured in the blood.

**Conclusion:** These data for the first time indicate significant anti-aging potential of examined herbs. Results showed that D-galactose induces a significant oxidative stress and promotes proinflammatory cascade of aging while all herbs more or less recovered these changes. Among 9 herbal extracts, *Silybum marianum* showed the best effect in restoring aging changes.

► Please cite this paper as:

Mohammadirad A, Aghamohammadali-Sarraf F, Badiei S, Faraji Z, Hajiaghaee R, Baeri M, Gholami M, Abdollahi M. Anti Aging Effects of Some Selected Iranian Folk Medicinal Herbs —Biochemical Evidences. Iran J Basic Med Sci; 2013; 16:1170-1180.

### Introduction

Aging as a complex of natural circumstance is exhibited by an augmentation in the chance of illness and finally death. Although some theories have been proposed as the mechanisms of aging but the one relating aging and cellular oxidative stress have received more supports. Therefore, it can be said that reduced sex hormones and augmented quantity of oxidative stress parameters or inflammatory cytokines are main biochemical manifestations of aging (1-2). In accord with this theory, the production of reactive oxygen species (ROS) and/or free radicals can injure cells and tissues paralleled by malfunction of many systems. The eventual consequence of these actions is aging and finally premature cell death (3). During aging process, various pro-inflammatory molecules are generated

to strengthen inflammation cascade associated with different age-related pathologies (4).

One of the problems in testing anti-aging compounds is lack of suitable animal models. Although several models have been used so far but among them, typical mouse D-galactose-induced model of aging is the best one that gives closer results to clinical studies. D-galactose is a sugar that at higher levels converts to aldose and hydroperoxide during the catalysis of galactose oxidase, culminated in the generation of free radicals (6). These modifications are substantially similar to the normal aging process demonstrated as neurological deterioration, diminished activity of antioxidant enzymes, and miserable immune responses (7-8).

Many scientists and pharmaceutical companies try to develop a drug to reduce speed of human aging

\*Corresponding author: Mohammad Abdollahi. Division of Toxicology, Department of Toxicology and Pharmacology, Faculty of Pharmacy and Pharmaceutical Sciences Research Center, Tehran University of Medical Sciences, Keshavarz Bulvd., Tehran, Iran. email: mohammad.abdollahi@utoronto.ca

but no effective drug has been discovered yet. In the last decade the importance of folk medicine and herbal medicines have been revisited that resulted in developing many effective drugs for many human diseases. For instance, in the recent years, efficacy of herbal medicines in diseases like inflammatory bowel diseases (9-10), obesity (11), diabetes (12), pancreatitis (13), osteoporosis (14), hyperlipidemia (15), and so on has been proved. Our recent systematic review specified anti-aging herbs and their characteristics in different clinical or experimental models (16). Most of anti-aging herbs have antioxidant components and reduces free radicals which are by-product of abnormal body metabolism in the elderly.

We recently proved anti-aging potential of naturally-based drugs like IMOD and Angipars which have strong antioxidant power (2). On the basis of our systematic review, among various species we could select nine herbs with the strongest antioxidant effects such as *Z. officinale*, *G. glabra*, *R. officinalis*, *P. harmala*, *A. vera*, *S. hortensis*, *T. scordium*, *H. perforatum* and *S. marianum* to test in D-galactose-induced model of mouse aging.

## Materials and Methods

### Chemicals

Thiobarbituric acid (TBA), trichloroacetic acid (TCA), n-butanol, hexadecyltrimethyl ammonium bromide (HETAB), tri (2-pyridyl)-s-triazine (TPTZ), HCl, malondialdehyde (MDA), ferric chloride ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ ), D-galactose, and vitamin E (Trolox) were purchased from Merck (Germany). Rat specific tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukine-1 $\beta$  (IL- $\beta$ ), interleukine-6 (IL-6), NF-kappa B (NF- $\kappa$ b) ELISA kits were purchased from BenderMed Systems (Austria). Testosterone and dehydroepiandrosterone ELISA kits were purchased from Dia Metra (Italy).

### Preparation of herbs, extraction, and lyophilization

Herbs were provided from the Research Institute of Medicinal Plants Karaj during June 2009 and were air-dried at room temperature. Samples were authenticated by a botanist (Y. Ajani), and voucher specimens were preserved in the central herbarium of medicinal plants (RIMP). The scientific names and tested parts of the herbal materials are detailed in Table 1. The dried plants powder (40 g) was

extracted using percolation method by methanol at room temperature. Solvents were completely removed by drying under reduced pressure at 40°C in a rotary evaporator. The samples were stored at 4°C until use. Specifically, the *A. vera* leaves (1000 g) were washed in a suitable bactericide (chlorhexidine). The filets were grounded to a liquid, and the pulp was removed by filtering. The resultant gel was then freeze dried.

### Animals

Male BALB/c mice (12 weeks old, 18–22 g) were provided from Tehran University of Medical Sciences (TUMS) animal house. The animals were housed in standard polypropylene cages with wired-net top in a controlled room (temperature 23±1°C, humidity 55±10%, 12 hr light–dark cycle) and were allowed free access to standard laboratory pellet diet and water during the experiments. All ethical issues on the use of animals were carefully considered and the study protocol was approved by TUMS review board with code number of 90-03-33-15668.

### Experimental design

Before starting the main study, a pilot was designed to set up aging model and to get proper doses of herbal materials. In the main study, 120 mice were randomly divided into 12 groups, each consisting of 10 animals. D-galactose was dissolved in a measured quantity of mice drinking water. D-galactose was given to 11 out of 12 groups of animals at 500 mg/kg D-galactose per 1 ml drinking water for 6 weeks by gavage (2, 17). The 12<sup>th</sup> group of animals was the sham group which was not given D-galactose. After 2 weeks, the 11 groups which had been given D-galactose were randomly divided into aging control group (500 mg/kg D-galactose per 1 ml drinking water, for 6 weeks), positive control group (500 mg/kg D-galactose per 1ml drinking water plus vitamin E 200 mg/kg/day by gavage for 4 weeks) and herb-treated groups including 9 groups that each received 500 mg/kg D-galactose per 1 ml drinking water plus *Z. officinale* (250 mg/kg/day), *G. glabra* (150 mg/kg/day), *R. officinalis* (300 mg/kg/day), *P. harmala* (50 mg/kg/day), *A. vera* (150 mg/kg/day), *S. hortensis* (200 mg/kg/day), *T. scordium* (200 mg/kg/day), *H. perforatum* (135

**Table 1.** The scientific names and tested parts of the plant materials

Scientific name	Tested parts	Extraction yield (mg/g)	Used Dose (mg/kg)	References
<i>Zingiber officinale</i>	Rhizome	140.57	250	18
<i>Glycyrrhiza glabra</i>	Root	129.52	150	19
<i>Rosmarinus officinalis</i>	Aerial parts	236.51	300	20
<i>Peganum harmala</i>	Seed	169.25	50	21
<i>Aloe vera</i>	Gel	4.87	150	22
<i>Satureja hortensis</i>	Aerial parts	134	200	23
<i>Teucrium scordium</i>	Aerial parts	205	200	24
<i>Hypericum perforatum</i>	Aerial parts	100.58	135	25
<i>Silybum marianum</i>	Seed	123.49	150	26

mg/kg/day) and *S. marianum* (150 mg/kg/day), respectively by gavage for 4 weeks (18-26).

Twenty-four hours after the last treatment, blood samples were taken of each animal under anesthesia through the tail vein. Serum samples were obtained by centrifuging the whole blood at  $1000 \times g$  at  $4^{\circ}\text{C}$  for 10 min and the supernatants were transferred into several microtubes for separate biochemical assays and maintained at  $-80^{\circ}\text{C}$  until the analyses were performed. Biochemical markers including TNF- $\alpha$ , IL- $\beta$ , IL-6, NF- $\kappa$ b, ferric reducing total antioxidant power (TAP), lipid peroxidation (LPO) and male sex hormones including testosterone and dehydroepiandrosterone-sulfate (DHEA-S) were measured in the serum.

#### Measurement of LPO

LPO was measured by the reaction of thiobarbituric acid (TBA) with lipid peroxides. Samples were mixed with TCA (20%) and the precipitate was dispersed in  $\text{H}_2\text{SO}_4$  (0.05 M). After addition of TBA (0.2% in sodium sulfate), the sample was heated for 30 min in a boiling water bath. Then, TBA reactive substances (TBARS) as LPO marker adducts were extracted by n-butanol and absorbance was measured at 532 nm as described in details in our previous work (27). Data were expressed as nM.

#### Measurement of TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and NF- $\kappa$ b

Quantitative detection of TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and NF- $\kappa$ b levels in serum were performed using an enzyme-linked immunosorbent assay rat specific ELISA kit according to each specific brochure. The absorbance of the final colored product was measured in 450 nm as the primary wave length and 620 nm as the reference wave length. TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and NF- $\kappa$ b levels were expressed as pg/mg.

#### Measurement of TAP

Serum TAP was evaluated by measuring the ability to reduce  $\text{Fe}^{3+}$  to  $\text{Fe}^{2+}$ . Interaction of TPTZ with  $\text{Fe}^{2+}$  results in formation of a blue color with a maximum absorbance at 593. The whole procedure has been described in our previous study (27). Data were expressed as mM.

#### Measurement of testosterone and DHEA-S

For determination of testosterone and DHEA-S, specific ELISA kits were used and the instruction of their brochure was followed. Testosterone and DHEA-S were expressed as ng/ml.

#### Statistical analysis

Results are expressed as mean  $\pm$  standard error of the mean (SEM). Data were analyzed by one-way ANOVA followed by Tukey post-hoc test for multiple comparisons to ensure the variances of the data are distributed properly. A *P*-value less than 0.05 were

considered significant. The Stats Direct version 2.7.9 was used.

## Results

A significant increase in TBARS (Figure 1,  $11.9 \pm 0.2$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ) and a significant decrease in TAP (Figure 2,  $218 \pm 8$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ) were observed when sham group was compared with D-galactose-received aged group. Figures 3-6 show the effects of aging on the levels of TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF- $\kappa$ B, respectively in comparison to sham ( $32 \pm 2.3$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.2 \pm 0.05$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $27 \pm 3.9$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $45.7 \pm 2.4$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ). As shown in Figures 7 and 8, testosterone and DHEA-S ( $0.6 \pm 0.05$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.2 \pm 0.2$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice was lower than that in the sham.

#### Effects of *Z. officinale* in aged mice

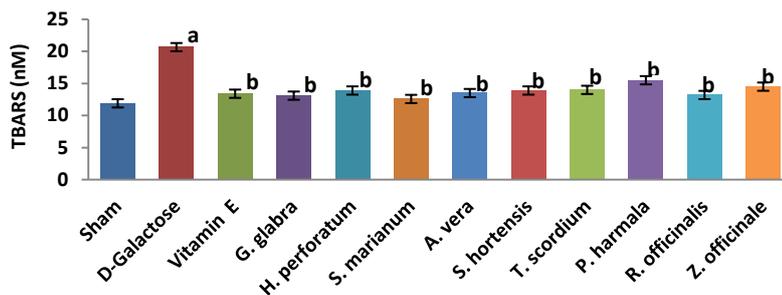
*Z. officinale* treatment recovered D-galactose-induced rats by reducing TBARS ( $14.5 \pm 1.6$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ), and increasing TAP ( $169 \pm 3.5$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ), (Figures 1, 2). Figures 3-6 show that administration of *Z. officinale* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF- $\kappa$ B ( $39 \pm 2.6$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.3 \pm 0.3$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $32.3 \pm 0.54$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $68.1 \pm 5.7$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *Z. officinale* increased testosterone and DHEA-S ( $0.48 \pm 0.04$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.28 \pm 0.17$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

#### Effects of *G. glabra* in aged mice

D-galactose-induced elevation of TBARS and reduction of TAP (Figures 1, 2) were significantly recovered following treatment with *G. glabra* ( $13.1 \pm 1.01$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ;  $203 \pm 17$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of *G. glabra* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$  and NF- $\kappa$ B ( $33 \pm 9.5$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.2 \pm 0.14$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $30.78 \pm 3.1$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $57.52 \pm 8.7$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *G. glabra* increased testosterone and DHEA-S levels ( $0.49 \pm 0.05$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.3 \pm 0.34$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

#### Effects of *R. officinalis* in aged mice

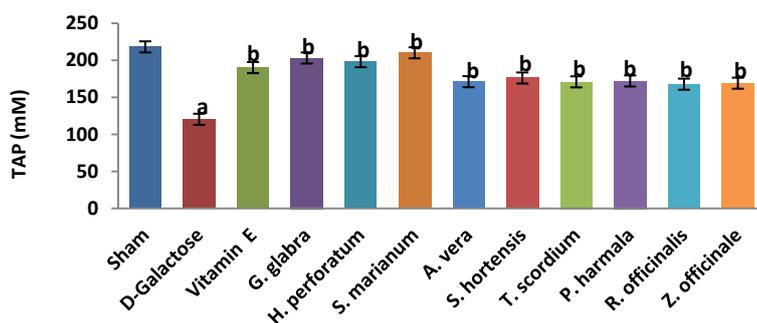
*R. officinalis* treatment recovered D-galactose-induced rats by reducing TBARS ( $13.2 \pm 0.63$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ), and increasing TAP ( $167.7 \pm 5.3$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ) (Figures 1, 2). Figures 3-6 show that administration of *R. officinalis* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF- $\kappa$ B ( $42 \pm 12$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.2 \pm 0.2$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $33.5 \pm 4.1$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $58.1 \pm 3.8$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *R. officinalis* increased testosterone, ( $0.46 \pm 0.09$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ) and DHEA-S ( $1.17 \pm 0.19$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.



**Figure 1.** Effects of herbs on serum thiobarbituric acid reactive substances (TBARS) as lipid peroxidation (LPO) marker in aged mice. Data are mean±SEM of ten animals.

<sup>a</sup>Significantly different between sham group and other groups at  $P<0.05$ .

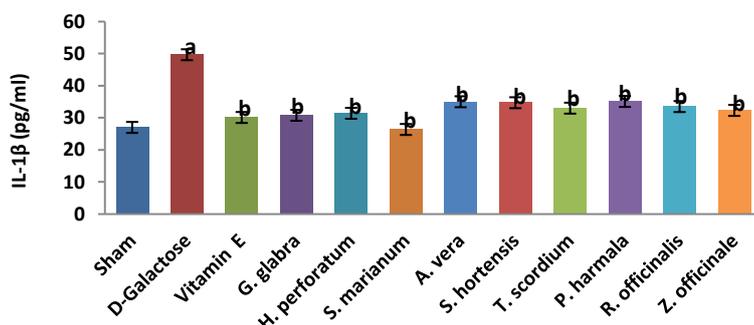
<sup>b</sup>Significantly different between D-galactose group and other groups at  $P<0.05$



**Figure 2.** Effects of herbs on serum total antioxidant power (TAP) of aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between sham group and other groups at  $P<0.05$

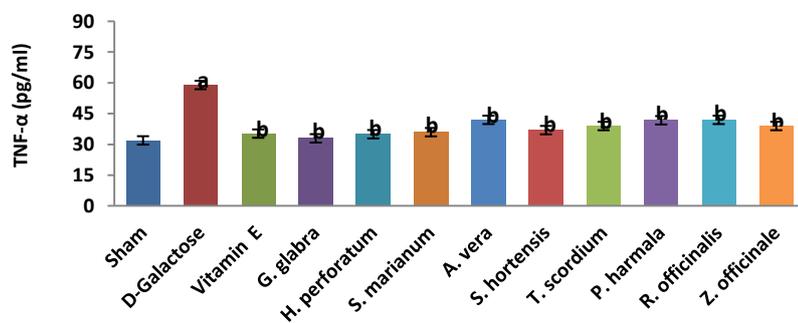
<sup>b</sup>Significantly different between D-galactose group and other groups at  $P<0.05$



**Figure 3.** Effects of herbs on serum interleukin-1 beta (IL-1β) in aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between Sham group and other groups at  $P<0.05$

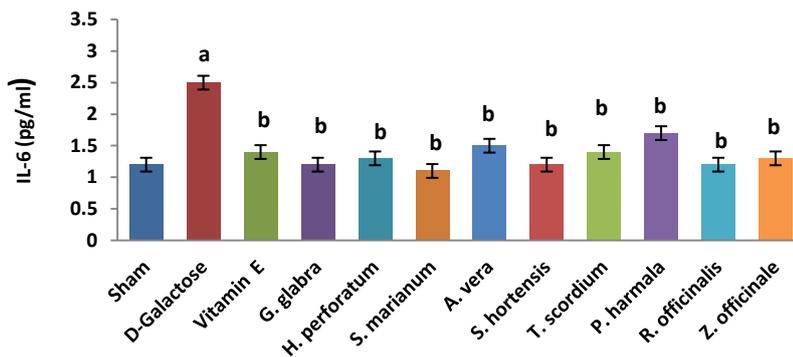
<sup>b</sup>Significantly different between D-Galactose group and other groups at  $P<0.05$



**Figure 4.** Effects of herbs on serum tumor necrosis factor-alpha (TNF-α) in aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between Sham group and other groups at  $P<0.05$

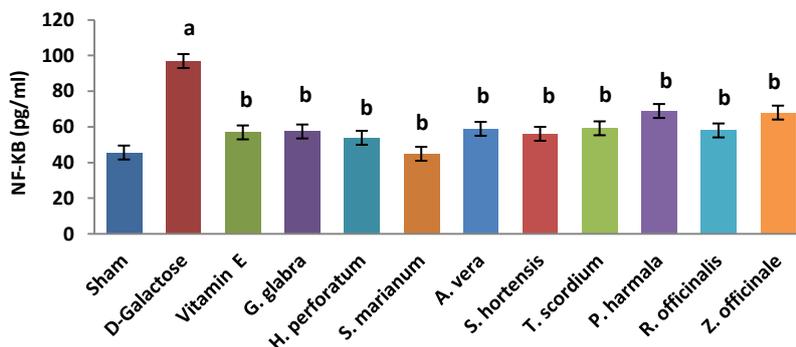
<sup>b</sup>Significantly different between D-Galactose group and other groups at  $P<0.05$ .



**Figure 5.** Effects of herbs on serum interleukin 6 (IL-6) in aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between Sham group and other groups at  $P<0.05$

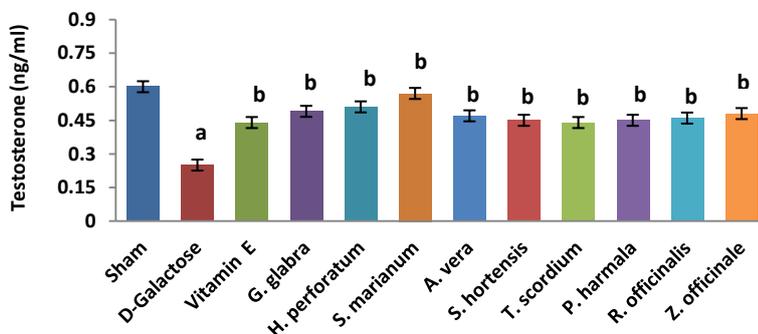
<sup>b</sup>Significantly different between D-Galactose group and other groups at  $P<0.05$



**Figure 6.** Effects of herbs on serum NF-kappaB (NF-kB) in aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between Sham group and other groups at  $P<0.05$

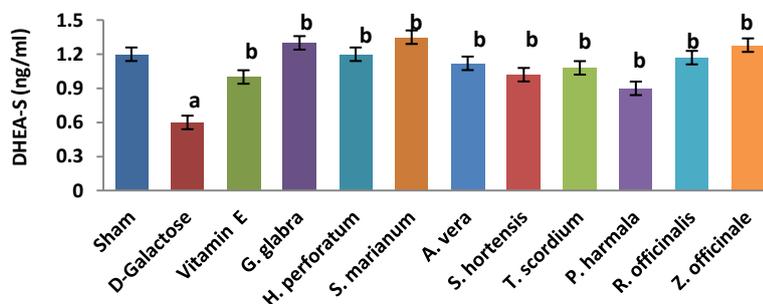
<sup>b</sup>Significantly different between D-Galactose group and other groups at  $P<0.05$



**Figure 7.** Effects of herbs on serum testosterone in aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between Sham group and other groups at  $P<0.05$

<sup>b</sup>Significantly different between D-Galactose group and other groups at  $P<0.05$



**Figure 8.** Effects of herb on serum dehydroepiandrosterone-sulfate (DHEA-S) in aged mice. Data are mean±SEM of ten animals

<sup>a</sup>Significantly different between Sham group and other groups at  $P<0.05$

<sup>b</sup>Significantly different between D-Galactose group and other groups at  $P<0.05$

*Effects of P. harmala in aged mice*

D-galactose-induced elevation of TBARS and reduction of TAP (Figure 1, 2) were significantly recovered following treatment with *P. harmala* ( $15.5 \pm 1.73$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ;  $172 \pm 13.9$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of *P. harmala* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF-kB ( $41.8 \pm 6.8$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.7 \pm 0.18$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $35.1 \pm 1.42$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $69 \pm 7.2$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *P. harmala* increased testosterone and DHEA-S ( $0.45 \pm 0.08$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $0.9 \pm 0.07$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

*Effects of A. vera in aged mice*

*A. vera* treatment recovered D-galactose-induced elevation of TBARS (Figure 1,  $13.5 \pm 1.7$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ), and improved reduction of TAP (Figure 2,  $171 \pm 4.03$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of *A. vera* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF-kB ( $42 \pm 6.94$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.5 \pm 0.09$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $35 \pm 0.77$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $59 \pm 14$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *A. vera* increased testosterone and DHEA-S levels ( $0.47 \pm 0.09$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.12 \pm 0.19$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

*Effects of S. hortensis in aged mice*

*S. hortensis* treatment recovered D-galactose-induced elevation of TBARS (Figure 1,  $13.9 \pm 2.2$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ), and increased TAP (Figure 2,  $176 \pm 14.16$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of *S. hortensis* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF-kB ( $37 \pm 9.9$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.2 \pm 0.05$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $34.7 \pm 1.8$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $56.2 \pm 13.9$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *S. hortensis* increased testosterone and DHEA-S ( $0.45 \pm 0.05$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.02 \pm 0.15$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

*Effects of T. scordium in aged mice*

D-galactose-induced elevation of TBARS and reduction of TAP (Figure 1, 2) were significantly recovered following treatment with *T. scordium* ( $14 \pm 0.76$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ;  $170.8 \pm 7.64$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of *T. scordium* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF-kB ( $39 \pm 10.26$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.4 \pm 0.28$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $33 \pm 1.1$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $59.3 \pm 4.42$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *T. scordium* increased testosterone and DHEA-S ( $0.44 \pm 0.05$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.08 \pm 0.24$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

*Effects of H. perforatum in aged mice*

*H. perforatum* treatment recovered D-galactose-induced rats by reducing TBARS ( $13.9 \pm 1.9$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ) and increasing TAP ( $198 \pm 23$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ) (Figures 1, 2). Figures 3-6 show that administration of *H. perforatum* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$  and NF-kB ( $35 \pm 6$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.3 \pm 0.17$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $31.4 \pm 0.51$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $53.98 \pm 2.7$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, *H. perforatum* increased testosterone and DHEA-S ( $0.51 \pm 0.06$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.2 \pm 0.18$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

*Effects of S. marianum in aged mice*

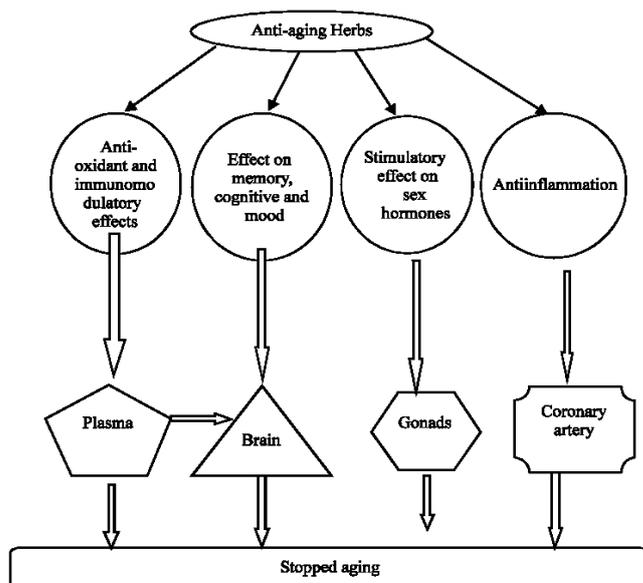
D-galactose-induced elevation of TBARS and reduction of TAP (Figure 1, 2) were significantly recovered following treatment with *S. marianum* ( $12.58 \pm 0.64$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ;  $210 \pm 12.14$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of *S. marianum* recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF-kB ( $36 \pm 5.4$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.1 \pm 0.09$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $26.36 \pm 1.1$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $45 \pm 4.2$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, the *S. marianum* recovered D-galactose-induced reduction of testosterone and DHEA-S ( $0.57 \pm 0.09$  vs.  $0.25 \pm 0.03$ ,  $P < 0.05$ ;  $1.35 \pm 0.22$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

*Effects of vitamin E in aged mice*

D-galactose-induced elevation of TBARS and reduction in TAP (Figure 1, 2) were significantly recovered following treatment with Vitamin E ( $13.4 \pm 0.83$  vs.  $20.66 \pm 0.88$ ,  $P < 0.05$ ;  $190 \pm 13.1$  vs.  $120 \pm 7.5$ ,  $P < 0.05$ ). Figures 3-6 show that administration of Vitamin E recovered D-galactose-induced increase in TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and NF-kB ( $35.5 \pm 2.12$  vs.  $59 \pm 15$ ,  $P < 0.05$ ;  $1.4 \pm 0.26$  vs.  $2.5 \pm 0.33$ ,  $P < 0.05$ ;  $30.1 \pm 2.2$  vs.  $49.66 \pm 3.4$ ,  $P < 0.05$ ;  $57 \pm 3.9$  vs.  $97 \pm 21.2$ ,  $P < 0.05$ ), respectively. As shown in Figures 7 and 8, vitamin E increased testosterone and DHEA-S ( $0.44 \pm 0.02$  vs.  $0.02 \pm 0.03$ ,  $P < 0.05$ ;  $1 \pm 0.16$  vs.  $0.6 \pm 0.08$ ,  $P < 0.05$ ) in aged mice.

**Discussion**

In this study, for the first time, we analyzed the anti-aging potentials of nine famous herbs in a well-setup animal aging model using chronic administration of D-galactose. Our results showed that production of free radicals is the principal reason of up-regulation of pro-inflammatory cytokines and the main determinant involved in the D-galactose-induced aging model. Furthermore, these herbs dramatically diminished oxidative stress and proinflammatory cytokines in the aged mice. Supporting the mechanism of action of these herbs and the theory of oxidative stress in aging, vitamin E was used as the standard and showed the similar effects in examined markers of aging.



**Figure 9.** Suggested mechanisms of action of herbs in reducing aging process. Adapted from corresponding author's previous paper published in open access source (16).

Interestingly, present results indicated improvement of testosterone and DHEA-S by herbs in the aged mice. Decline of steroid hormones with aging is already known and is believed a major contributor to elevation of pro-inflammatory markers (28).

Recent studies have shown the mechanisms of action of anti-aging herbs in reducing aging process that is divided into four categories including anti-oxidant, anti-inflammatory, effect on memory/cognition/mood, and the sex hormones (Figure 9). This indicates that most of anti-aging herbals have antioxidant components (16) and thus supports the present findings and hypothesis of this study.

*Ginger [Zingiber officinale Roscoe (Zingiberaceae)]* and supplements derived from ginger like zingerone, shogaols and gingerols posse the abilities for the treatment of chronic inflammation. The protective effects of *Z. officinale* in lessening macromolecular damage in aged mice were shown in this study. Besides, recent study has shown that ginger extracts owns antioxidant activity (29). It has been recently shown that pre-trial administration of this herb expedites conditioned inhibitory learning in adult rats (30). Also, it has been found that *Z. officinale* has possibly good effects on age-related execution shortages and defends against oxidative stress in old rats, suggesting this compound as a useful factor in treating age-related disturbances (31).

*G. glabra* (licorice extract) or licorice is the root of *G. glabra* from which a sweet flavor can be extracted. The results of this study showed that *G. glabra* has the protective effects in declining macromolecular damage in aged mice. It has been shown that *G. glabra* extract is the safest pigment-lightening agent with the fewest side effects (32). Additionally, *G. glabra* has anti-

inflammatory properties hypothetically helpful in diminishing skin ruddiness and postinflammatory hyperpigmentation. Interestingly, it appears to be more useful for the hyperpigmentation related to skin aging (33).

*R. officinalis* leaves possess a variety of bioactive agents, including antioxidants and anti-inflammatories (34). The most potent antioxidant constituents are polyphenolics such as carnosic acid and carnosol (35). The results of this study showed that *R. officinale* has the protective effects in decreasing macromolecular damage in aged mice during aging. In addition, *R. officinale* extract has shown free radical scavenging effect in the hippocampus (36). This is supported with a raising number of reports showing that natural extracts and phytochemicals have a constructive effect on brain aging through their action on ROS, specifically in the hippocampus (37).

*P. harmala* L. is known as Syrian rue, Wild rue and Harmal. The *P. harmala* has antibacterial, antifungal, antiviral, antioxidant, antidiabetic, antitumor, antileishmanial, insecticidal, cytotoxic, hepatoprotective, and antinociceptive effects (38-39). In this study *P. harmala* showed the protective effects in improving antioxidant, anti-inflammatory and male sex hormones that were affected in aged mice during aging. In fact, flavonoids as a powerful antioxidant isolated from *P. harmala*, can remove the lipid peroxide radicals (40). Also, *P. harmala* treatment appeared to be a versatile strategy to conserve testicular uprightness and function during aging in male rats (41).

The leaves of *A. vera* (*A. barbadensis*) (Fam. Liliaceae) are the source of aloe vera gel. *A. vera* gel is greatly used in cosmetics and toiletries for its moisturizing and regenerating action. Also, the leaf

of *A. vera* could assist cellular repairing, imbibition of foods, vitamins, minerals and vital nutrients (42). In this study *A. vera* showed a protective effect in improving antioxidant, anti-inflammatory and male sex hormones that were affected in aged mice. The anti-inflammatory property of *A. vera* has been documented in inflammation through suppression of free radicals and ROS (43). It has been shown that the life-long dietary supplementation of *A. vera* suppresses many age-related consequences in rats (44). Also, it has been suggested *A. vera* could suppress oxidative damage and age-related increases in hepatic cholesterol during life-long dietary (45).

*S. hortensis* L. is an annual culinary herb belonging to the family Labiatae. It is known as summer savory. Besides, this plant exhibited analgesic, antibacterial, antifungal, antioxidant and antihyperglycemic properties. In addition, the antigenotoxic effects of *S. hortensis* L. was shown on rat lymphocytes exposed to oxidative stress (46). The major constituents of the *S. hortensis* are carvacrol, gamma-terpinene, thymol and p-cymene (47). The investigation showed that carvacrol, thymol and flavonoids of *Satureja* spices are responsible to marked reduction of serum cholesterol in diabetic patients (48-50). It has been reported that age-related alterations of fatty acid composition in liver was accompanied by intake of savory essential oil through intensification of polyunsaturated fatty acids synthesis in mice liver and reduction of lipid peroxidation products (51). As a result, *S. hortensis* shows protective effects in improving antioxidant, anti-inflammatory and male sex hormones that were affected in aged mice during aging.

The genus *Teucrium* (Labiatae) comprises 12 species, which possess antioxidant, anti-spasmodic, antinociceptive and anti-inflammatory properties (52-54). According to the literature, *b*-caryophyllene and caryophyllene oxide were reported as the main sesquiterpenes in many *Teucrium* species. Investigation of the chemical constituents of the oil of *T. scordium* showed *b*-caryophyllene, caryophyllene oxide and also (*E*)-*b*-farnesene as the major components of *T. scordium* (55). In addition, *b*-caryophyllene is known as anti-inflammatory sesquiterpene (56); this effect may confirm the anti-inflammatory activity of this plant (55). Nevertheless, no reports are valid about *T. scordium* in relation to its possible oxidative stress inhibitory potential in aged individuals. As much as we know this is the first study that shows antioxidant and anti-inflammatory effects of *T. scordium* extract in the aged rats. Our results indicated that *T. scordium* decreases inflammatory mediators and increases anti-oxidative power and steroid hormones in aged mice. Thus, *T. scordium* could be included in the diet

as a nutritional supplement to increase the defenses of body against oxidative stress.

*H. perforatum* L. is responsible for pharmacological properties, antiseptics (57), anti-inflammatory (58), antitumoral activities (59, 60). Today, *H. perforatum* is known as one of the few economic plants that include great ingredients of hypericins, hyperforins, and flavonoids. The results of this study showed that *H. perforatum* owns the protective effects in decreasing macromolecular damage in aged mice during aging. St. John's wort is the dried tops or aerial parts of *H. perforatum* which gathered before or during flowering and is used in the therapy of anxiety related to aging (61).

Silymarin (SM), a flavonoids complex known as 'milk thistle' is extracted from the fruit of *Silybum marianum* (L.) Gaertn. (*Carduus marianus* L., Asteraceae). Interestingly, the present findings confirmed that *S. marianum* causes the best effects in improving antioxidant, anti-inflammatory and male sex hormones in aged mice. This effect is so important and should be considered as an advantage. This can be explained with current knowledge that among many medicinal plants, *S. marianum*, has been greatly used for centuries as a natural popular complementary medicine for the treatment of several diseases. The main indications for the use of silymarin are related to the hepatoprotection (62, 63). Also, its efficacy in inflammatory oxidative-mediated diseases like colitis has been confirmed (64, 65). It is noteworthy that several age-related brain and neurodegenerative diseases happen due to amplified oxidative stress. The search for compounds acting on the upgrading of cognitive performance and neuroprotection through antioxidant motion is now a great interest (66). Excitingly, it has been shown that use of *S. marianum* for prevention and treatment of neurodegenerative diseases and processes associated with aging improves physiological responses against the ROS in the neural cells (67).

## Conclusion

Taking collectively, the present results confirmed our hypothesis that the herbs with highest antioxidant power may reduce speed and rate of aging as evidenced by recovery of proinflammatory cytokines and sex hormones. Among tested herbs, *S. marianum* showed the best effect in improving all the D-galactose-induced aging effects. Since all of the selected and examined herbs are already found safe in human and there are good information from traditional medicine, therefore, they can be supplemented into the diet of elderly people to reduce speed of aging. Testing the mixture of these herbs together or with other anti-aging products is among the plans of future.

## Acknowledgment

This study was undertaken as a project and the results described in this paper were part of student thesis. Authors also thank assistance of National Elite Foundation and the INSF.

## References

1. Momtaz S, Abdollahi M. A comprehensive review of biochemical and molecular evidences from animal and human studies on the role of oxidative stress in aging: An epiphenomenon or the cause. *Asian J Anim Vet Adv* 2012; 7:1-19.
2. Ghanbari S, Yonessi M, Mohammadirad A, Gholami M, Baeeri M, Khorram-Khorshid HR, et al. Effects of IMOD and Angipars on mouse D-galactose-induced model of aging. *DARU J Pharm Sci* 2012; 20:68.
3. Zhang X, Zhang A, Jiang B, Bao Y, Wang J, An L. Further pharmacological evidence of the neuroprotective effect of catalpol from *Rehmannia glutinosa*. *Phytomedicine* 2008; 15:484-490.
4. Chung HY, Sung B, Jung KJ, Zou Y, Yu BP. The molecular inflammatory process in aging. *Antioxid Redox Signal* 2006; 8:572-581.
5. Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci* 2000; 908:244-254.
6. Song X, Bao M, Li D, Li YM. Advanced glycation in D-galactose induced mouse aging model. *Mech Ageing Dev* 1999; 108:239-251.
7. Wei H, Li L, Song Q, Ai H, Chu J, Li W. Behavioral study of the D-galactose induced aging model in C57BL/6 J mice. *Behav Brain Res* 2005; 157:245-251.
8. Xu Y, Wu T, Jin Y, Fu Z. Effects of age and jet lag on d-galactose induced aging process. *Biogerontology* 2009; 10:153-61.
9. Rahimi R, Abdollahi M. Herbal medicines for the management of irritable bowel syndrome: A comprehensive review. *World J Gastroenterol* 2012; 18:589-600.
10. Rahimi R, Mozaffari S, Abdollahi M. On the use of herbal medicines in management of inflammatory bowel diseases: A systematic review of animal and human studies. *Digestive Diseases and Sciences. Dig Dis Sci* 2009; 54:471-480.
11. Hasani-Ranjbar S, Nayebi N, Larijani B, Abdollahi M. A systematic review of the efficacy and safety of herbal medicines used in the treatment of obesity. *World J Gastroenterol* 2009; 15:3073-3085.
12. Rahimi R, Nikfar S, Larijani B, Abdollahi M. A review on the role of antioxidants in the management of diabetes and its complications. *Biomed Pharmacother* 2005; 59:365-373.
13. Mohseni-Salehi-Monfared SS, Larijani B, Abdollahi M. Islet transplantation and antioxidant management: A comprehensive review. *World J Gastroenterol* 2009; 15: 1153-1161.
14. Abdollahi M, Larijani B, Rahimi R, Salari P. Role of oxidative stress in osteoporosis. *Therapy* 2005; 2:787-796.
15. Hasani-Ranjbar S, Nayebi N, Moradi L, Mehri A, Larijani B, Abdollahi M. The efficacy and safety of herbal medicines used in the treatment of hyperlipidemia; a systematic review. *Curr Pharm Des* 2010; 16:2935-2947.
16. Hasani-Ranjbar S, Khosravi S, Nayebi N, Larijani B, Abdollahi M. A systematic review of the efficacy and safety of anti-aging herbs in animals and human. *Asian J Anim Vet Adv* 2012; 7:621-640.
17. Ho SC, Liu JH, Wu RY. Establishment of the mimetic aging effect in mice caused by D-galactose. *Biogerontology* 2003; 4:15-18.
18. Jagetia G, Baliga M, Venkatesh P. Ginger (*Zingiber officinale* rosc.), a dietary supplement, protects mice against radiation-induced lethality: Mechanism of action. *Cancer Biother Radiopharma* 2004; 19:422-435.
19. Sofiabadi M, Esmaeili MH, Haghdoost Yazdy H, Azhdari Zarmehri H. The prenatal consumption of aqueous extract of *Glycyrrhiza glabra*, improves memory retrieval in mice. *J Med Plants* 2011; 10:49-54.
20. Maistro EL, Mota SF, Lima EB, Bernardes BM, Goulart FC. Genotoxicity and mutagenicity of *rosmarinus officinalis* (labiateae) essential oil in mammalian cells in vivo. *Gene Mol Res* 2010; 9:2113-22.
21. Lamchouri F, Settaf A, Cherrah Y, Zemzami M, Lyoussi B, Zaid A, et al. Antitumour principles from *Peganum harmala* seeds. *Therapie* 1999; 54:753-758.
22. Tanaka M, Yamada M, Toida T, Iwatsuki K. Safety evaluation of supercritical carbon dioxide extract of *Aloe vera* gel. *J Food Sci* 2010; 77:T2-T9.
23. Hajhashemi V, Ghannadi A, Pezeshkian SK. Antinociceptive and anti-inflammatory effects of *Satureja hortensis* L. extracts and essential oil. *J Ethnopharmacol* 2002; 82:83-87.
24. Kundaković T, Milenković M, Topić A, Stanojković T, Juranić Z, Lakušić B. Cytotoxicity and antimicrobial activity of *teucrium scordium* L. (Lamiaceae) extracts. *Afr J Microbiol Res* 2011; 5:2950-2954.
25. Higuchi A, Yamada H, Yamada E, Jo N, Matsumura M. Hypericin inhibits pathological retinal neovascularization in a mouse model of oxygen-induced retinopathy. *Mol Vis* 2008; 14:249-254.
26. Chou CH, Chen YC, Hsu MC, Tsai WL, Chang CY, Chiu CH. Effect of silymarin on lipid and alcohol metabolism in mice following long-term alcohol consumption. *J Food Biochem* 2012; 36:369-377.
27. Astaneie F, Afshari M, Mojtahedi A, Mostafalou S, Zamani MJ, Larijani B, et al. Total antioxidant capacity and levels of epidermal growth factor and nitric oxide in blood and saliva of insulin-dependent diabetic patients. *Arch Med Res* 2005; 36:376-381.
28. Maggio M, Basaria S, Ceda GP, Ble A, Ling SM, Bandinelli S, et al. The relationship between testosterone and molecular markers of inflammation in older men. *J Endocrinol Invest* 2005; 28:116-119.
29. Ahmed RS, Seth V, Banerjee BD. Influence of dietary ginger (*Zingiber officinale* Rosc) on antioxidant defense system in rat: comparison with ascorbic acid. *Indian J Exp Biol* 2000; 38:604-606.
30. Topic B, Tani E, Tsiakitzis K, Kourounakis PN, Dere E, Hasenöhrl RU, et al. Enhanced maze performance and reduced oxidative stress by combined extracts of *zingiber officinale* and *ginkgo biloba* in the aged rat. *Neurobiol Aging* 2002; 23:135-143.

30. Topic B, Hasenöhrl RU, Häcker R, Huston JP. Enhanced conditioned inhibitory avoidance by a combined extract of *Zingiber officinale* and *Ginkgo biloba*. *Phytother Res* 2002; 16:312-315.
31. Draelos ZD. Skin lightening preparations and the hydroquinone controversy. *Dermatol Ther* 2007; 20:308-313.
32. Cronin H, Draelos ZD. Top 10 botanical ingredients in 2010 anti-aging creams. *J Cosmet Dermatol* 2010; 9:218-225.
33. Altinier G, Sosa S, Aquino RP, Mencherini T, Della Loggia R, Tubaro A. Characterization of topical antiinflammatory compounds in *Rosmarinus officinalis* L. *J Agric Food Chem* 2007; 55:1718-1723.
34. Ramírez P, Señoráns FJ, Ibañez E, Reglero G. Separation of rosemary antioxidant compounds by supercritical fluid chromatography on coated packed capillary columns. *J Chromatogr A* 2004; 1057:241-245.
35. Posadas SJ, Caz V, Largo C, De la Gándara B, Matallanas B, Reglero G, et al. Protective effect of supercritical fluid rosemary extract, *Rosmarinus officinalis*, on antioxidants of major organs of aged rats. *Exp Gerontol* 2009; 44:383-389.
36. Bastianetto S, Quirion R. Natural extracts as possible protective agents of brain aging. *Neurobiol Aging* 2002; 23:891-8897.
37. Hamden K, Carreau S, Ayadi F, Masmoudi H, El Feki A. Inhibitory effect of estrogens, phytoestrogens, and caloric restriction on oxidative stress and hepato-toxicity in aged rats. *Biomed Environ Sci* 2009; 22:381-387.
38. Monsef-Esfahani HR, Ghobadi A, Iranshahi M, Abdollahi M. Antinociceptive effects of *Peganum harmala* L. alkaloid on mouse formalin test. *J Pharm Pharm Sci* 2004; 7:70-75.
39. Berrougui H, Isabelle M, Cloutier M, Hmamouchi M, Khalil A. Protective effects of *Peganum harmala* L. extract, harmine and harmaline against human low-density lipoprotein oxidation. *J Pharm Pharmacol* 2006; 58:967-974.
40. Hamden K, Masmoudi H, Ellouz F, ElFeki A, Carreau S. Protective effects of *Peganum harmala* extracts on thiourea-induced diseases in adult male rat. *J Environ Biol* 2008; 29:73-77.
41. Asadi-Shahmirzadi A, Mozaffari S, Sanei Y, Baeeri M, Hajiaghahae R, Monsef-Esfahani HR, et al. Benefit of *Aloe vera* and *Matricaria recutita* mixture in rat irritable bowel syndrome: Combination of antioxidant and spasmolytic effects. *Chin J Integr Med* 2012; [Epub ahead of print]
42. Yoo EA, Kim SD, Lee WM, Park HJ, Kim SK, Cho JY, et al. Evaluation of antioxidant, antinociceptive, and anti-inflammatory activities of ethanol extracts from *Aloe saponaria* Haw. *Phytother Res* 2008; 22:1389-1395.
43. Ikeno Y, Hubbard GB, Lee S, Yu BP, Herlihy JT. The influence of long-term *Aloe vera* ingestion on age-related disease in male Fischer 344 rats. *Phytother Res* 2002; 16:712-218.
44. Lim BO, Seong NS, Choue RW, Kim JD, Lee HY, Kim SY, et al. Efficacy of dietary *aloe vera* supplementation on hepatic cholesterol and oxidative status in aged rats. *J Nutr Sci Vitaminol (Tokyo)* 2003; 49:292-296.
45. Mosaffa F, Behravan J, Karimi G, Iranshahi M. Antigenotoxic effects of *Satureja hortensis* L. on rat lymphocytes exposed to oxidative stress. *Arc Pharm Res* 2006; 29:159-164.
46. Momtaz S, Abdollahi M. An update on pharmacology of *Satureja* species; from antioxidant, antimicrobial, antidiabetes and anti-hyperlipidemic to reproductive stimulation. *Int J Pharmacol* 2010; 6:454-461.
47. Momtaz S, Abdollahi M. A systematic review of the biological activities of *Satureja* L. Species. *Pharmacologyonline* 2008; 2:34-54.
48. Vosough-Ghanbari S, Rahimi R, Kharabaf S, Zeinali S, Mohammadirad A, Amini A, et al. Effects of *Satureja khuzestanica* on serum glucose, lipids and markers of oxidative stress in patients with type 2 diabetes mellitus: A double-blind randomized controlled trial. *Evid Based Complement Alternat Med* 2010; 7:465-470.
49. Abdollahi M, Salehnia A, Mortazavi SH, Ebrahimi M, Shafiee A, Fouladian F, et al. Antioxidant, antidiabetic, antihyperlipidemic, reproduction stimulatory properties and safety of essential oil of *Satureja khuzestanica* in rat *in vivo*: a oxycopharmacological study. *Med Sci Monit* 2003; 9:BR331-BR335.
50. Misharina TA, Burlakova EB, Fatkullina LD, Terenina MB, Krikunova NI, Vorob'eva AK, et al. Changes in fatty acid composition in the brain and liver in aging mice of high cancer risk AKR strain and effect of savory essential oil administration on leukemic process. *Biomed Khim* 2011; 57:604-614.
51. Abdolghaffari AH, Baghaei A, Moayer F, Esmaily H, Baeeri M, Monsef-Esfahani HR, et al. On the benefit of *Teucrium* in murine colitis through improvement of toxic inflammatory mediators. *Hum Exp Toxicol* 2010; 29:287-295.
52. Hasani P, Yasa N, Vosough-Ghanbari S, Mohammadirad A, Dehghan G, Abdollahi M. *In vivo* antioxidant potential of *Teucrium polium*, as compared to alpha-tocopherol. *Acta Pharm* 2007; 57:123-129.
53. Abdollahi M, Karimpour H, Monsef-Esfahani HR. Antinociceptive effects of *Teucrium polium* L total extract and essential oil in mouse writhing test. *Pharmacol Res* 2003; 48:31-35.
54. Morteza-Semnani K, Saeedi M, Akbarzadeh M. Essential oil composition of *Teucrium scordium* L. *Acta Pharm* 2007; 57:499-504.
55. Tambe Y, Tsujiuchi H, Honda G, Ikeshiro Y, Tanaka S. Gastric cytoprotection of the non-steroidal anti-inflammatory sesquiterpene, beta-caryophyllene. *Planta Med* 1996; 62:469-470.
56. Reichling J, Weseler A, Saller R. A current review of the antimicrobial activity of *Hypericum perforatum* L. *Pharmacopsychiatry* 2001; 34:S116-S118.
57. Mozaffari S, Esmaily H, Rahimi R, Baeeri M, Sanei Y, Asadi-Shahmirzadi A, et al. Effects of *Hypericum perforatum* extract on rat irritable bowel syndrome. *Pharmacogn Mag* 2011; 7:213-223.
58. Martarelli D, Martarelli B, Pediconi D, Nabissi MI, Perfumi M, Pompei P. *Hypericum perforatum* methanolic extract inhibits growth of human prostatic carcinoma cell line orthotopically implanted in nude mice. *Cancer Lett* 2004; 210:27-33.

59. Rahimi R, Abdollahi M. An update on the ability of St. John's wort to affect the metabolism of other drugs. *Exp Opin Drug Met Toxicol* 2012; 8: 691-708.
60. Hänsgen KD, Vesper J, Ploch M. Multicenter double-blind study examining the antidepressant effectiveness of the hypericum extract LI 160. *J Geriatr Psychiatry Neurol* 1994; 7:S15-S18.
61. Das SK, Vasudevan DM. Protective effects of silymarin, a milk thistle (*Silybium marianum*) derivative on ethanol-induced oxidative stress in liver. *Indian J Biochem Biophys* 2006; 43:306-311.
62. Hasani-Ranjbar S, Larijani B, Abdollahi M. A systematic review of the potential herbal sources of future drugs effective in oxidant-related diseases. *Inflamm Allergy Drug Targets* 2009; 8:2-10.
63. Esmaily H, Hosseini-Tabatabaei A, Rahimian R, Khorasani R, Baeeri M, Barazesh-Morgani A, *et al.* On the benefits of silymarin in murine colitis by improving balance of destructive cytokines and reduction of toxic stress in the bowel cells. *Cent Eur J Biol* 2009; 4: 204-213.
64. Malihi F, Hosseini-Tabatabaei A, Esmaily H, Khorasani R, Baeeri M, Abdollahi M. Improvement of inflammatory and toxic stress biomarkers by silymarin in a murine model of type one diabetes mellitus. *Cent Eur J Biol* 2009; 4:369-380.
65. Metodiewa D, Kośka C. Reactive oxygen species and reactive nitrogen species: relevance to cyto(neuro)toxic events and neurologic disorders. An overview. *Neurotox Res* 2000; 1: 197-233.
66. Galhardi F, Mesquita K, Monserrat JM, Barros DM. Effect of silymarin on biochemical parameters of oxidative stress in aged and young rat brain. *Food Chem Toxicol* 2009; 47:2655-2660.