Iranian Journal of Basic Medical Sciences

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8-OH-DPAT (5-HT1A agonist) Attenuates 6-Hydroxydopamine-induced catalepsy and Modulates Inflammatory Cytokines in Rats

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ARTICLE INFO	ABSTRACT
<i>Article type:</i> Original article	Objective (<i>s</i>): Neuroinflammation in Parkinson disease (PD) is associated with glial cells activation and production of different inflammatory cytokines. In this study, we investigated the effect of chronic administration of 8-OH-DPAT on 6-OHDA-induced catalepsy and levels of inflammatory cytokines in cerebrospinal fluid (CSF). Materials and Methods: Catalepsy was induced by unilateral infusion of 6-OHDA (8 µg/2 µl/rat) into the central region of the sabstantia nigra pars compacta (SNc) being assessed by the bar-test, 5, 60, 120 and 180 min after intraperitoneal (IP) administration of 8-OH-DPAT (5-HT _{1A} receptor agonist; 0.25, 0.5 and 1mg/kg. IP for 10 days). CSF samples were collected on the tenth day of 8-OH-DPAT administration and analyzed by ELISA method to measure levels of TNF-α, IL-1β and IL-6. Results: Chronic injection of 8-OH-DPAT decreased catalepsy in a dose dependent manner when compared with the control group. The most anti-cataleptic effect was observed at the dose of 1 mg/kg of 8-OH-DPAT. Levels of TNF-α in CSF increased three weeks after 6-OHDA injection while there was a significant decrease in TNF-α level of parkinsonian animals treated with 8-OH-DPAT (1 mg/kg, IP for 10 days). IL-1β and IL-6 decreased and increased in parkinsonian rats and in 8-OH- DPAT treated parkinsonian rats, respectively. Conclusion: Our study indicated that chronic administration of 8-OH-DPAT improves catalepsy in 6- OHDA-induced animal model of PD and restores central concentration of inflammatory cytokines to the basal levels. 5-HT _{1A} receptor agonists can be suggested as potential adjuvant therapy in PD by modulation of cerebral inflammatory cytokines.
Article history: Received: Sep 9, 2012 Accepted: Jan 29, 2013	
<i>Keywords:</i> 8-OH-DPAT Catalepsy Chronic Cytokines Rat	

Please cite this paper as:

Sharifi H, Mohajjel Nayebi A, Farajnia S. 8-OH-DPAT (5-HT1A agonist) Attenuates 6-Hydroxydopamine-induced catalepsy and Modulates Inflammatory Cytokines in Rats. Iran J Basic Med Sci; 2013; 16: 1270-1275.

Introduction

Parkinson disease (PD) is the second most common and progressive neurodegenerative disease caused mainly by loss of dopaminergic neurons in the substantia nigra pars compacta (SNc). The major movement symptoms are rigidity, akinesia, tremor and postural abnormalities as well as cognitive disturbances (1).

The role of neuroinflammation in degeneration of nigrostriatal neurons is of interest to many investigators (2, 3). The first evidence for the role of inflammation in PD came from an observation by McGeer and colleagues on activated microglia and T cells in the post-mortem SNc of a patient suffering from PD (4). Epidemiological studies have shown that the incidence of idiopathic PD is lower in chronic users of anti-inflammatory drugs (5, 6). Neuroinflammation is regulated by many signal molecules including cytokines. They are multifunctional proteins and in the CNS, play a role in the normal development of the brain as well as in neuroimmuno-pathological processes following injury and neurodegeneration (7). Several studies have reported significant increase of pro-inflammatory cytokines such as IFN- γ , IL-1 β and TNF- α , being expressed by glial cells in the nigrostratial regions of patients with PD (4, 8-10). In general, proinflammatory cytokines such as TNF- α has neurotoxic effects, while IL-6 and IL-1 β , classical proinflammatory cytokines, have a dual effect. For instance, low concentrations of IL-6 protect neuronal cells from death, while larger concentrations are neurotoxic (11, 12).

Previous studies have shown that serotonergic system is involved in PD (13-15). Serotonergic projections originating from the dorsal raphe nuclei innervate all parts of the basal ganglia and play a role

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in the regulation of movements executed by the basal ganglia (14). In this context, role of 5-HT_{1A} receptors in motor impairments of PD is the center for attention (13, 16). Activation of these receptors could decrease serotonin release and subsequently improve motor function in 6-OHDA-lesioned rats (17, 18). Studies have shown that stimulation of the 5-HT_{1A} receptor attenuates anoxia-induced apoptosis in the neuronal HN2-5 (Hippocampal neuronderived cell line) cells (19). It seems that cytokines act as messengers between the immune system and the brain, exerting their effect on serotonergic system through several processes such as degradation of their precursor, tryptophan (20). The role of pro-inflammatory cytokines in the pathogenesis of PD has been shown in several studies (12, 21, 22), although the effect of chronic administration of 5-HT_{1A} receptor agonists on 6-OHDA-induced catalepsy and the role of cytokines such as TNF- α , IL-1 β and IL-6 has not been clearly studied yet. Thus, in this study we attempted to investigate the effect of chronic administration of 8-OH-DPAT on 6-OHDA-induced catalepsy and possible involvement of TNF- α , IL-1 β and IL-6.

Materials and Methods Chemicals

All chemicals were obtained from Sigma Chemical Co. (USA), except for ELISA kits, which were purchased from eBioscience Co. (Austria). All solutions were prepared freshly on the experimentation day. 8-OH-DPAT (a 5-HT_{1A} receptor agonist) and 6-OHDA were dissolved in physiological saline (0.9% NaCl) and 0.9% saline containing 0.2% (w/v) ascorbic acid, respectively. 6-OHDA was injected into the central region of the substantia nigra pars compacta (SNc) in a total volume of 2 μ l /rat with a constant injection rate of 0.2 μ l /min.

Animals

The experiments were carried out on male Wistar rats weighing 270-300 g. Animals were housed in standard polypropylene cages, four per cage, under a 12:12 hr light/dark schedule at an ambient temperature of $25 \pm 2^{\circ}$ C and were allowed food and water ad libitum. Animals were acclimated to the testing conditions for 2 days before the behavioral experiment was conducted. A11 procedures were carried out under the ethical guidelines of the Tabriz University of Medical Sciences. Twenty-four rats were divided into three groups: normal, sham-operated (receiving 2 µl vehicle) and 6-OHDA ($8 \mu g/2 \mu l/rat$)-injected.

6-OHDA-induced SNc lesion

Animals were anesthetized with an IP injection of ketamine (50 mg/kg) and xylazine (5 mg/kg). After being deeply anesthetized (loss of corneal and toe pad reflexes), rats were mounted in a Stoelting stereotaxic frame in the flat skull position. The scalp was shaved, swabbed with povidone-iodine 10%, and a central incision made to expose the skull. 6-OHDA was injected thorough a guide cannula (23 gauge stainless steel) implanted in the SNc. The coordinates for this site were based on the rat brain atlas (23) as follows: anteroposterior (AP): -5.0 mm from the bregma; mediolateral (ML): -2.1 mm from the midline and dorsoventral (DV): -7.7 from the skull.

Desipramine (25 mg/kg, IP) was injected 30 min before the intra-SNc injection of 6-OHDA to avoid destruction of noradrenergic neurons. Thereafter, 6-OHDA (8 µg per rat in 2 µl saline with 0.2% ascorbic acid) was infused with an infusion pump at a constant flow rate of 0.2 µl/min into the left SNc. At the end of the infusion, the injection tube was kept implanted for an additional 2 min and then was slowly retracted. Sham-operated animals were submitted to the same procedure but 2 µl vehicle (0.9% saline containing 0.2% (w/v) ascorbic acid) instead of 6-OHDA was infused into the SNc.

Catalepsy test

Catalepsy was measured using a standard bar test 21 days after 6-OHDA and 10 days after IP injection of 8-OH-DPAT. In this method, forepaws of rats were placed over a 9-cm-high standard wooden bar, and the duration of retention of rats in this imposed posture was considered as the bar test elapsed time. The end point of catalepsy was considered when both front paws were removed from the bar or when the animal moved its head in an exploratory manner. The cut-off time of the test was 600 sec. The test was carried out 5, 60, 120 and 180 min after drug administration on the 10th day. All observations were made between 9 am and 4 pm. After a three-week recovery period, only the rats being markedly immobilized in the bar test were subjected to further experimentation (parkinsonian rats). Afterwards, the parkinsonian rats were randomly divided into equal groups and received IP injections of 8-OHD-PAT (0.25, 0.5 and 1 mg/kg, IP) once daily (9 a.m.) for 10 days.

CSF sampling

CSF samples were collected on day 10 of 8-OH-DPAT administration. Animals were anesthetized by IP injections of ketamine (50 mg/kg) and xylazine (5 mg/kg) and mounted in a Stoelting stereotaxic frame. The skull was kept in 45 ° position and CSF was aspirated using a sterile 100 μ l syringe 23gauge needle. The CSF samples were kept at -70°C until being assessed by Enzyme-linked immunosorbent assays (ELISA) method.

Analysis of TNF- α , IL-1 β and IL-6 expression by ELISA

ELISA method was employed for determination of TNF- α , IL-1 β and IL-6 in CSF samples. Assays were

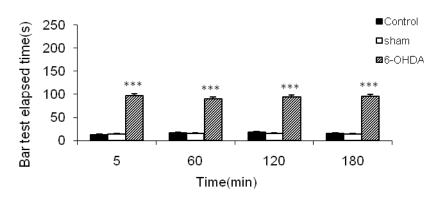


Figure 1. The results of bar test in control, sham-operated and 6-OHDA ($8 \mu g/2 \mu l/rat$)-lesioned rats. Each bar represents the mean ± SEM of bar test elapsed time (sec); n = 8 rats in each group; *** *p* <0.001 when compared with normal and sham-operated groups

performed by a commercial ELISA kit (IBL INTERN-ATIONAL GMBH) as per manufacturer's instructions and in the similar conditions for all assays. Briefly, the frozen CSF samples were diluted, added into the wells and incubated at room temperature for 120 min on a microplate shaker. Subsequent to washing, diluted Streptavidin-Horseradish peroxidas e-conjugated antimouse TNF A, IL-1B and IL-6 were reacted for 60 min at room temperature (on microplate shaker set at 200 rpm). After washing for second time, the wells were developed with tetramethyl benzidine (TMB) for 10 min and the optical densities were read at 450 nm with an ELISA reader.

Histology

All animals having guide cannula were sacrificed at the end of the experiments. Brain dissections were performed in all animals to confirm the exact implantation of guide cannula into the SNc. Brain in the injecting tube in situ was fixed in 10% formalin for 1 week. The location of the tip of the injecting tube was then verified in serial sections. Only the results from bar tests in animals with the tip of the injecting tube within the SNc area were used for statistical analysis.

Statistical analysis

Statistical analysis for each data set was calculated by SPSS software (version 16.0). Data were expressed as mean+SEM, and one-way ANOVA test was utilized to analyze in the data from behavioral and biochemical experiments. In the case of significant variation (P< 0.05), the values were compared by Tukey test.

Results

6-OHDA-induced catalepsy

6-OHDA was able to induce significant (P < 0.001) catalepsy in comparison with both normal and sham-operated rats (Figure 1).

Effect of 8-OH-DPAT on 6-OHDA-in duced catalepsy

Four groups of 6-OHDA-lesioned rats received saline or one of the three different doses of 8-OH-DPAT (0.25, 0.5 and1 mg/kg, IP), respectively for 10 days. The results showed that 8-OH-DPAT attenuated the severity of 6-OHDA-induced catalepsy (P< 0.001) (Figure 2).

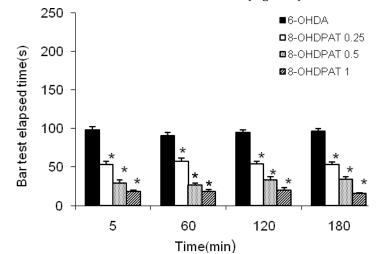


Figure 2. The results of bar test in 6-OHDA (8 μ g/2 μ l/rat)-lesioned rats treated with 8-OH-DPAT (0.25, 0.5, and 1 mg/kg, IP for 10 days). Each bar represents the mean± SEM of catalepsy time (sec); n = 8 rats in each group; * p < 0.001 when compared with 6-OHDA-lesioned rats



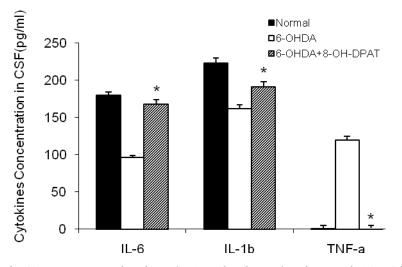


Figure 3. The CSF concentration of cytokines (IL-6, IL-1 β and TNF- α) in the control, 6-OHDA-lesioned rats and 6-OHDA -lesioned rats treated with 8-OH-DPAT (1 mg/kg, IP for 10 days). Each bar represents the mean± SEM of cytokines concentration in the CSF (pg/ml) n = 8 rats in each group; * *P*< 0.001 when compared with 6-OHDA-lesioned rats

Effect of 8-OH-DPAT on inflammatory parameters

Results showed that TNF- α was undetectable in normal rats while it was increased three weeks after 6-OHDA injection and decreased following 10 days of IP administeration of 8-OH-DPAT to normal animals. Levels of the two other cytokines were decreased in 6-OHDA-lesioned rats in comparison with normal group but increased 10 days after injection of 8-OH-DPAT (Figure 3).

Discussion

In our previous studies (13, 24) the potential anticataleptic effects of 5-HT_{1A} receptor agonists in 6-OHDA-lesioned rats has been investigated in single dose administrations. Given the chronic clinical administrations of such drugs, we investigated the potenti al anticataleptic effects of chronic administration of 8-OH-DPAT in 6-OHDA-lesioned rats in the current study. Our results showed that intra-SNc injection of 6-OHDA induced catalepsy in animals when assessed by the bar test (13). Bar test is a standard test being frequently used for evaluation of catalepsy induced by 6-OHDA and neuroleptic drugs in rodents (25). According to the results, chronic administration of 8-OH-DPAT, an agonist of 5-HT_{1A} receptors, improved catalepsy in 6-OHDA-lesioned rats in a dose dependent manner. Such finding confirms the previous studies, reporting a promising role for 5-HT_{1A} agonists in decreasing the motor disorders associated with PD (24).

 $5-HT_{1A}$ receptors are widely distributed throughout the basal ganglia. They are located on dorsal raphe neurons with efferents to the striatum and on cortical neurons that send glutamatergic projections into the basal ganglia (26). In the basal ganglia, serotonin modulates dopamine-related motor activity through affecting the $5-HT_{1A}$ receptor (27). It has been shown that $5-HT_{1A}$ agonists improve motor impairments in parkinsonian animals via stimulation of somatodendritic $5-HT_{1A}$ receptors and subsequent decrease in serotonin release from the nerve endings (28). Furthermore, studies have indicated that $5-HT_{1A}$ receptor plays a role in neuronal survival (29, 30) and has a neuroprotective effect in animal models of stroke and traumatic brain injury (29, 31). Such effect is exerted through inhibition of glutamate release that leads to a reduction in the putative excitotoxicity-mediated cell death (31).

Herein, we assessed levels of inflammatory cytokines i.e TNF- α , IL-6 and IL-1 β in the CSF of parkinsonian rats being treated with chronic injections of 8-OH-DPAT. According to our results, there was a significant increase in the amount of TNF- α in parkinsonian rats whereas its levels were resorted to normal ranges by chronic administration of 8-OH-DPAT. This is in accordance with other studies reporting that toxic effects of 6-OHDA are in part mediated through the activation of microglia and increasing levels of TNF- α in both SN and striatum (4, 9, 32). The substantia nigra (SN) has high density of microglia and it is hypothesized that DA neurons are susceptible to inflammatory damage as a major stimuli for neurodegenerative diseases (33). Activated microglia release proinflammatory cytokines such as TNF- α that play a key role in modulation of inflammatory responses (34).

The levels of IL-1 β and IL-6 were decreased in parkinsonian rats when compared with normal (nonparkinsonian) animals. In parkinsonian rats, which were treated with chronic injections of 8-OH-DPAT, the levels of IL-1 β and IL-6 were increased to that of normal rats. It has been reported that there is an increase in CSF concentration of IL-1 β and IL-6 in parkinsonian rats (32, 35). In addition to their proinflammatory effect, these are pleiotropic cytokines, which can produce neuroprotective effects in PD, Alzheimer disease (AD) and CNS injuries (32, 36). Our results showed that IL-1 β and IL-6 were decreased in 6-OHDA-lesioned rats while in 6-OHDAlesioned rats, being treated with 8-OH-DPAT, the levels of IL-1 β and IL-6 were restored to normal values. This is in agreement with previous studies which suggest a neuroprotective effect for these cytokines (22, 37).

Conclusion

Our data suggest that chronic administration of 8-OH-DPAT improves catalepsy in 6-OHDA-lesioned rats. Moreover, we suggest that 5-HT_{1A} receptor agonists can be utilized as adjuvant therapy along with commonly used anti-parkinsonian drugs. However, further clinical investigations should be carried out to prove this.

Acknowledgment

We wish to thank the director of Drug Applied Research Center, dean of Faculty of Pharmacy and Research Vice-Chancellor of Tabriz University of Medical Sciences, Tabriz, Iran for supporting this study. The results described in this paper were part of student thesis (No. 61).

References

1. Hattori N, Sato S. Animal models of Parkinson's disease: Similarities and differences between the disease and models. Neuropathology 2007; 27:479–483.

2. Bolin LM, Strycharska-Orczyk I, Murray R, Langston JW, Di Monte D. Increased vulnerability of dopaminergic neurons in MPTP-lesioned interleukin-6 deficient mice. J Neurochem 2002; 83:167–175.

3. Di Filippo M, Chiasserini D, Tozzi A, Picconi B, Calabresi P. Mitochondria and the link between Neuroinflammation and Neurodegeneration. J Alzheimers Dis 2010; 20:S369–379.

4. McGeer PL, Itagaki S, Boyes BE, McGeer EG. Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. Neurology 1988;38:1285-1291

5. Tansey MG, McCoy MK, Frank-Cannon TC. Neuroinflammatory mechanisms in Parkinson's disease: Potential environmental triggers, pathways, and targets for early therapeutic intervention. Exp Neurol 2007; 208:1–25.

6. Esposito E, Di Matteo V, Benigno A, Pierucci M, Crescimanno G, Di Giovanni G. Non-steroidal antiinflammatory drugs in Parkinson's disease. Exp Neurol 2007; 205:295–312.

7. Manthripragada AD, Schernhammer ES, Qiu J, Friis S, Wermuth L, Olsen JH, *et al.* Non-steroidal anti-inflammatory drug use and the risk of Parkinson's disease. Neuroepidemiology 2011; 36:155-161.

8. Sawada M, Imamura K, Nagatsu T. Role of cytokines in inflammatory process in Parkinson's disease. J Neural Transm Suppl 2006; 70:373–381.

9. Tansey MG, Frank-Cannon TC, McCoy MK, Lee JK, Martinez TN, McAlpine FE, *et al.* Neuroinflammation in Parkinson's disease: is there sufficient evidence for mechanism-based interventional therapy? Front Biosci 2008; 13:709-717.

10. Nagatsu T, Mogi M, Ichinose H, Togari A. Changes in cytokines and neurotrophins in Parkinson's disease.J Neural Transm Suppl 2000; 60:277-290.

11. Brodacki B, Staszewski J, Toczyłowska B, Kozłowska E, Drela N, Chalimoniuk M, *et al.* Serum interleukin (IL-2, IL-10, IL-6, IL-4), TNF_, and INF_ concentrations are elevated in patients with atypical and idiopathic parkinsonism. Neurosci Lett 2008; 441:158–162.

12. Li XZ, Bai LM, Yang YP, Luo WF, Hu WD, Chen JP, *et al.* Effects of IL-6 secreted from astrocytes on the survival of dopaminergic neurons in lipopolysaccharide-induced inflammation. Neurosci Res Suppl 2009; 65:252–258.

13. Sawada M, Imamura K, Nagatsu T. Role of cytokines in inflammatory process in Parkinson's disease. J Neural Transm 2006; 70:373–381.

14. Nayebi MA, Sheidaei H. Buspirone improves haloperidol-induced Parkinson disease in mice through 5-HT_{1A} receptors. Daru 2010; 18:41-45.

15. Fox SH, Chuang R, Brotchie JM. Serotonin and Parkinson's disease: On movement, mood, and madness. Mov Disord 2009; 24:1255-1266.

16. Huot P, Fox SH, Brotchie JM. The serotonergic system in Parkinson's disease. Prog Neurobiol 2011; 95:163–212.

17. Mignon L, Wolf WA. Postsynaptic 5-HT_{1A} receptor stimulation increases motor activity in the 6-hydroxydopamine-lesioned rat: implications for treating Parkinson's disease. Psychopharmacology (Berl) 2007; 192:49–59.

18. Riad M, Garcia S, Watkins KC, Jodoin N, Doucet E, Langlois X, *et al.* Somatodendritic localization of 5-HT_{1A} and preterminal axonal localization of 5-HT1B serotonin receptors in adult rat brain. J Comp Neural. 2000; 417:181-194.

19. Bibbiani F, Oh JD, Chase TN. Serotonin $5-HT_{1A}$ agonist improves motor complications in rodent and primate parkinsonian models. Neurology 2001; 57:1829-1834.

20. Adayev T, El-Sherif Y, Barua M, Penington NJ, Banerjee P. Agonist stimulation of the serotonin1A receptor causes suppression of anoxia-induced apoptosis via mitogen-activated protein kinase in neuronal HN2-5 cells. J Neurochem 1999; 72:1489-1496.

21. Anisman H, Merali Z, Hayley S. Neurotransmitter, peptide and cytokine processes in relation to depressive disorder: Comorbidity between depression and neurodegenerative disorders. Prog Neurobiol 2008; 85:1–74.

22. Sawada H, Hishida R, Hirata Y, Ono K, Suzuki H, Muramatsu S, *et al.* Activated microglia affect the nigro-striatal dopamine neurons differently in neonatal and aged mice treated with 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine. J Neurosci Res 2007; 85:1752–1761.

23. Bick RJ, Poindexter BJ, Kott MM, Liang YA, Dinh K, Kaur B, *et al.* Cytokines disrupt intracellular patterns of Parkinson's disease-associated proteins alphasynuclein, tau and ubiquitin in cultured glial cells. Brain Res 2008; 1217: 203-212.

24. Paxinos G, Watson C: The rat brain in stereotaxic coordinates. Academic Press, Sydney, 1982.

25. Mahmoudi J, Nayebi AM, Samini M, Reyhani-Rad S, Babapour V. Buspirone improves the anticataleptic effect of levodopa in 6-hydroxydopaminelesioned rats. Pharmacol Rep 2011; 63:908-914.

26. Dupre KB, Eskow KL, Negron G, Bishop C. The differential effects of 5-HT_{1A} receptor stimulation on dopamine receptor-mediated abnormal involuntary movements and rotations in the primed hemiparkinsonian rat. Brain Res 2007; 1158:135-143.

27. Knobelman DA, Kung HF, Lucki I. Regulation of extracellular concentrations of 5-Hydroxytryptamine (5-HT) in mouse striatum by 5-HT_{1A} and 5-HT_{1B} receptors. J Pharmacol Exp Ther 2000; 292:1111–1117.

28. Madhavan L, Freed WJ, Anantharam V, Kanthasamy AG. 5-Hydroxytryptamine 1A receptor activation protects against N-methyl-D-aspartate-induced apoptotic cell death in striatal and mesencephalic cultures. J Pharmacol Exp Ther 2003; 304:913-923.

29. Dupre KB, Eskow KL, Barnum CJ, Bishop C. Striatal 5-HT_{1A} receptor stimulation reduces D1 receptor-induced dyskinesia and improves movement in the hemiparkinsonian rat. Neuropharmacology 2008; 55:1321-1328.

30. Bezard E, Gerlach I, Moratalla R, Gross CE, Jork R. $5-HT_{1A}$ receptor agonist-mediated protection from MPTP toxicity in mouse and macaque models of

Parkinson's disease. Neurobiol Dis 2006; 23:77 – 86. 31. Raymond JR, Mukhin YV, Gelasco A, Turner J, Collinsworth G, Gettys TW, *et al.* Multiplicity of mechanisms of serotonin receptor signal transduction. Pharmacol Ther 2001; 92:179-212.

32. Charkhpour M, Nayebi AR, Doustar Y, Hassanzadeh K. 8-OH-DPAT prevents morphineinduced apoptosis in rat dorsal raphe nucleus: A possible mechanism for attenuating morphine tolerance. Anesth Analg 2010; 111:1316-1321.

33. Nagatsu T. Parkinson's disease: changes in apoptosis-related factors suggesting possible gene therapy. J Neural Transm 2009; 109:731–745.

34. Qin L, Wu X, Block ML, Liu Y, Breese GR, Hong JS, *et al.* Systemic LPS Causes Chronic Neuroinflammation and Progressive Neurodegeneration. Glia 2007;55:453-462.

35. Shadrina MI, Slominsky PA, Limborska SA. Molecular mechanisms of pathogenesis of parkinson's disease. Int Rev Cell Mol Biol 2010; 281:229-266.

36. Hirsch EC, Hunot S. Neuroinflammation in Parkinson's disease: a target for neuroprotection? Lancet Neurol 2009; 8:382–397.

37. Imamura K, Hishikawa N, Ono K, Suzuki H, Sawada M, Nagatsu T, *et al.* Cytokine production of activated microglia and decrease in neurotrophic factors of neurons in the hippocampus of Lewy body disease brains. Acta Neuropathol 2005; 109:141–150. 38. Shaftel SS, Griffin WS, O'Banion MK. The role of interleukin-1 in neuroinflammation and Alzheimer disease: an evolving perspective. J Neuroinflammation 2008; 5:7.