

Antidotal and protective effects of mangosteen (*Garcinia mangostana*) against natural and chemical toxicities: A review

Aidin Mohammadi Zonouz¹, Mahboobeh Ghasemzadeh Rahbardar², Hossein Hosseinzadeh^{2, 3*}

¹ School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

² Pharmaceutical Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran

³ Department of Pharmacodynamics and Toxicology, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

ARTICLE INFO

Article type:

Review

Article history:

Received: Jun 23, 2022

Accepted: Dec 19, 2022

Keywords:

Analgesics

Anti-inflammatory agents

Anti-oxidants

Apoptosis

Hypoglycemic agents

Neuroprotective agents

Phytotherapy

Xanthones

ABSTRACT

Chemical and natural toxic compounds can harm human health through a variety of mechanisms. Nowadays, herbal therapy is widely accepted as a safe method of treating toxicity. *Garcinia mangostana* (mangosteen) is a tree in the Clusiaceae family, and isoprenylated xanthones, its main constituents, are a class of secondary metabolites having a variety of biological properties, such as anti-inflammatory, anti-oxidant, pro-apoptotic, anti-proliferative, antinociceptive, neuroprotective, hypoglycemic, and anti-obesity. In this review, the protective activities of mangosteen and its major components against natural and chemical toxicities in both *in vivo* and *in vitro* experiments were evaluated. The protective effects of mangosteen and its components are mediated primarily through oxidative stress inhibition, a decrease in the number of inflammatory cells such as lymphocytes, neutrophils, and eosinophils, reduction of inflammatory mediators such as tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1), interleukin-6 (IL-6), interleukin-8 (IL-8), cyclooxygenase-2 (COX-2), prostaglandin (PG) E₂, inducible nitric oxide synthase, and nuclear factor- κ B (NF- κ B), modulation of apoptosis and mitogen-activated protein kinase (MAPK) signaling pathways, reducing p65 entrance into the nucleus, α -smooth muscle actin (α -SMA), transforming growth factor β 1 (TGF β 1), improving histological conditions, and inhibition in acetylcholinesterase activity.

► Please cite this article as:

Mohammadi Zonouz A, Ghasemzadeh Rahbardar M, Hosseinzadeh H. Antidotal and protective effects of mangosteen (*Garcinia mangostana*) against natural and chemical toxicities: A review. Iran J Basic Med Sci 2023; 26: 492-503. doi: <https://dx.doi.org/10.22038/IJBMS.2023.66900.14674>

Introduction

Humans and animals are now exposed to different types of toxic substances, either directly or indirectly, through a variety of routes, including food, water, soil, and air (1). Toxic agents can cause different disorders such as nephrotoxicity (2), neurotoxicity (3), hepatotoxicity (4), and cardiotoxicity (5) in human or animal bodies. The most frequent mechanism regulating chemical and natural toxicities is oxidative stress. The abundance of nucleobase products characteristic of the oxygen assault on deoxyribonucleic acid (DNA) in cultured cells and animals exposed to carcinogenic agents provides the most compelling evidence that toxicants cause genotoxic damage via an oxidative process. The potential of toxicants to generate reactive radicals, which cause DNA damage, lipid peroxidation, and protein sulfhydryl depletion, has been demonstrated (6, 7). Different toxins can cause inflammation, and these events are likely to involve a variety of pathways. An increase in the formation of reactive oxygen species (ROS) and redox-related alterations are frequently strongly linked to inflammatory processes. Likely, the first event leading to inflammation in the case of some kinds of toxicants is their capacity to stimulate ROS generation (8). Toxins are also well known for causing apoptotic cell death and playing a role in a variety of clinical diseases. The regulation of toxicant-induced apoptotic pathways appears to be largely dependent on oxidative stress (9). In apoptosis, several gene families are involved or work together, including the caspases, inhibitors of apoptosis proteins,

the B cell lymphoma-2 family of genes, the tumor necrosis factor (TNF) receptor gene superfamily, and the p53 gene (10).

Many chemical medications, such as cimetidine, cilastatin, dexrazoxane, and β -blockers, are used to treat these disorders (11, 12), and despite their protective action against toxic agents, they have numerous side effects, including fatigue, hypotension, dizziness, diarrhea, and impotence (13). As a result, seeking out new medications to combat the harmful consequences of toxins is essential.

Herbs and spices have been employed to enhance the flavor of meals and beverages around the world for centuries, as well as being used as protective agents. Today, researchers are focusing more on the possible benefits of herbal drugs as alternative therapies for illness prevention or as antitoxic agents, due to their potential efficacy, low toxicity, and minimal side effects (14-18).

Mangosteen, *Garcinia mangostana* Linn., a member of the Clusiaceae family, is a tropical evergreen tree that is native to Southeast Asia with a height of 6-25 m and a dark-brown or practically black bark. Its product, mangosteen, is a reddish/dark purple fruit that is regarded as "the queen of fruits" and has a juicy, soft, edible pulp, and delicious flavor. In folk medicine, mangosteen pericarp has been used to treat fever, convulsions, diarrhea, dysentery, stomach discomfort, trauma, pain, infected wounds, suppuration, and chronic ulcers (19, 20).

Mangosteen contains phenolic acids, xanthones,

*Corresponding author: Hossein Hosseinzadeh. Pharmaceutical Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran; Department of Pharmacodynamics and Toxicology, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran. Tel: +98-5138819042; Fax: +98-5138823251; Email: hosseinzadehh@mums.ac.ir

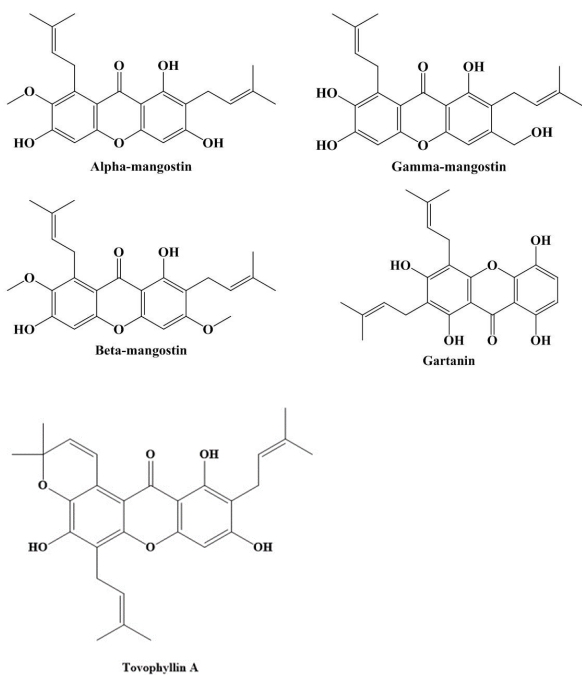


Figure 1. Structures of effective compounds of mangosteen

prenylated benzophenone derivatives, flavonoids, anthocyanins, and condensed tannins. Furthermore, it has been hypothesized that the pericarp of the mangosteen is a rich source of oligomeric proanthocyanidins with B-type linkages. Alpha-mangostin (α -MG) and gamma-mangostin (γ -MG) are the most common xanthones in mangosteen fruits, although there are also beta-mangostin (β -MG), gartanin, and other xanthones in mangosteen (Figure 1). In addition, according to previous research, most of the biological activities of *G. mangostana* are linked to the quantity of α -MG (21, 22). Mangosteen and its components have been shown to have medical and pharmacological characteristics, including antimalarial (23), anti-metabolic syndrome (19), antimicrobial (24), antifungal (25), antidiabetic (26), anticancer (27), antiproliferative (28), anti-adipogenesis (29), anti-oxidant, anti-apoptotic, anti-inflammatory, analgesic (30), and antidotal (31) activities (Figure 2).

Since mangosteen and its main constituents are potent anti-oxidants, anti-inflammatory, and anti-apoptotic agents and can regulate different cellular pathways, including the mitogen-activated protein kinase (MAPK) signaling pathway, they might be effective in managing natural and chemical toxicities. Hence, the protective effects of mangosteen and its main constituents, essential oils, and extracts against various natural and chemical toxic compounds have been reviewed in this article to help expand the mangosteen and its constituents' application as protective agents and potential lead compounds against toxicities.

Methods

In this comprehensive review article, our team argued various documents in Google Scholar, PubMed, Web of Science, and Scopus. This publication contains both *in vitro* and *in vivo* studies. This review did not consider any time constraints. The keywords for this study were *Garcinia mangostana*, mangosteen, xanthones, α -mangostin, natural toxins, chemical toxicity, nephrotoxins, neurotoxins, hepatotoxins, and cardiotoxins. The protective effects of mangosteen were investigated under two headings: biological and chemical toxic substances.

Mangosteen and its main constituents against biological agents-induced toxicity

This section discusses the antidotal and protective actions of mangosteen and its main constituents against several biological agents that cause toxicity. In addition, in this part, certain key defensive mechanisms are discussed (Figure 3).

Lipopolysaccharide

Lipopolysaccharide (LPS), a glycolipid generated by most gram-negative bacteria, is one of the most researched bacterial surface compounds. LPS is an amphiphilic molecule made up of three distinct regions, which are lipid A, the core region, and the O-antigen polysaccharide (32). The toxic effects of LPS in mammalian cells are caused by the lipid A portion binding to toll-like receptors (TLRs), which stimulate the innate immune system and trigger the production of inflammatory cytokines such as tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1), and

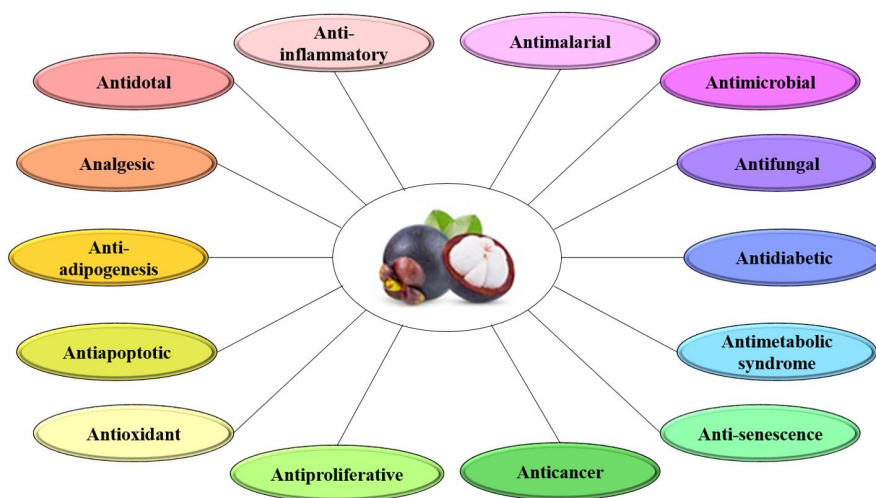


Figure 2. Pharmacological effects of mangosteen and its components

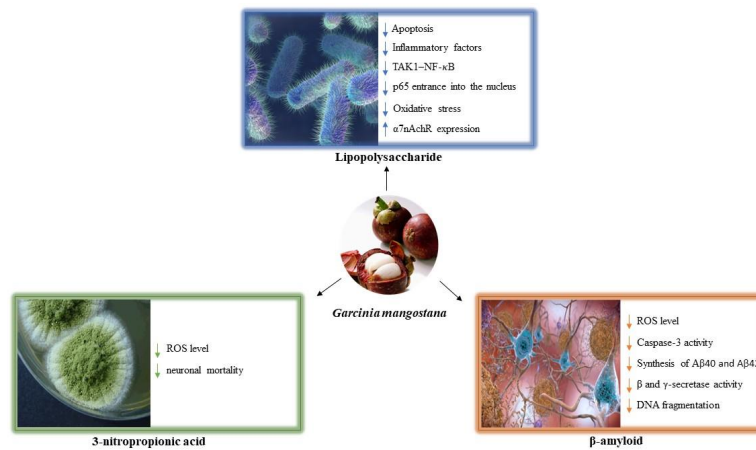


Figure 3. The mechanism of protective effects of mangosteen on natural toxicant-induced changes

interleukin-6 (IL-6), which lead to a potentially deadly systemic inflammatory response known as “septic shock” (32, 33). Below, we discuss intestinal inflammation, lung injury, the cytotoxicity of human gingival fibroblasts, and liver failure.

Intestinal inflammation

In a study, the anti-inflammatory effect of α-MG was assessed by administering α-MG (2.5, 5, and 10 μM, for 1 h) to an LPS-induced inflammation model of rat intestinal epithelial cells (IEC-6 cells) and the outcomes showed that α-MG administration demonstrated anti-inflammatory and anti-apoptotic properties by reducing apoptosis, inflammatory factors (nitric oxide (NO), prostaglandin (PG) E2, IL-6, TNF-α, and IL-1β) production, activation of transforming growth factor-activated kinase 1-nuclear factor-κB (TAK1-NF-κB) signaling pathway-related proteins, and p65 [involved in nuclear translocation and activation, nuclear factor- kappa-B (NF-κB) heterodimer formation] entrance into the nucleus in LPS-stimulated IEC-6 cells (34).

As a consequence, α-MG administration has an anti-inflammatory effect on LPS-induced intestinal inflammation by reducing apoptosis, inflammatory factors (NO, PGE2, IL-6, TNF-α, and IL-1β) production, activation of TAK1-NF-κB signaling pathway-related proteins, and p65 entrance into the nucleus (Figure 4).

Lung injury

Yang et al. examined the role of α-MG in the cholinergic anti-inflammatory pathway (CAP) and its therapeutic potential in the treatment of acute lung injury (ALI). They administered α-MG (40 mg/kg, 3 days, PO) to male Sprague Dawley rats before inducing ALI with an injection of LPS. Also, for the *in vitro* tests, they employed RAW264.7 cells to confirm the effects of α-MG ((5 μg/ml), at various times (0.5, 1, 2, 4, and 6 h)) on CAP. The findings revealed that α-MG reversed the decline in α7 nicotinic acetylcholine receptor (α7nAChR) expression in the lungs of ALI rats and enhanced α7nAChR and choline acetyltransferase (ChAT) expression in RAW 264.7 cells. Also, α-MG affected acetylcholinesterase (AChE) expression at 5 g/ml and its catalytic activity was lowered by almost 95%. Altogether, α-MG injection resulted in NF-κB suppression and acute inflammatory remission (35).

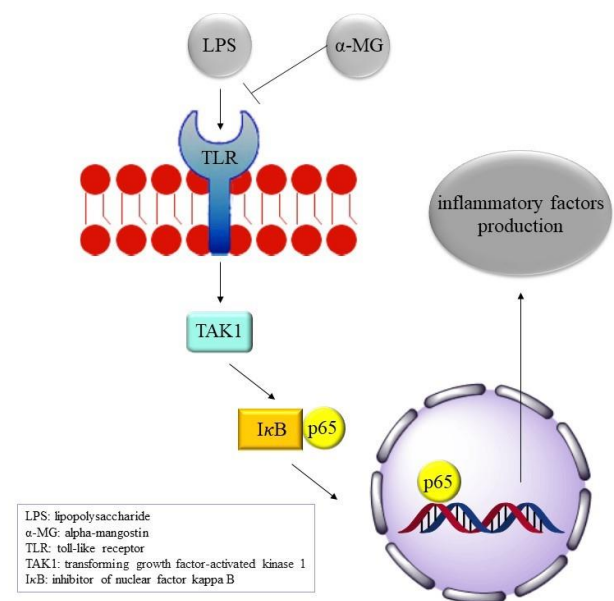


Figure 4. α-mangostin antidotal molecular mechanism against LPS-induced intestinal inflammation

In another study, researchers evaluated the therapeutic effects of α-MG (15 and 45 mg/kg/day, 3 days, PO) on LPS induce ALI in male Sprague Dawley rats. In ALI rats, it was discovered that α-MG therapy improved histological conditions (reduced interalveolar septal thickening, alveolar hemorrhage, and cells infiltration), lowered leucocyte counts, declined oxidative stress (a little recovery in superoxide dismutase (SOD) activity and reversed the elevation of malondialdehyde (MDA)), and decreased TNF-α levels. Also, α-MG therapy reduced the expressions of nicotinamide phosphoribosyltransferase (NAMPT) and sirtuin 1 (Sirt1), which was accompanied by a synchronized drop in nicotinamide adenine dinucleotide (NAD) and TNF-α. α-MG also inhibited high mobility group box 1 (HMGB1), TLR4, and p-p65 in RAW 264.7 cells. These findings implied that α-MG treatments reduced NAMPT/NAD levels, which helped to relieve TLR4/NF-κB-mediated inflammation in macrophages, which is critical for ameliorating ALI in rats (36).

As a result, α-MG treatment protects against LPS-induced lung damage through mechanisms such as increased

peripheral acetylcholine and $\alpha 7$ nAChR expression, as well as inhibition of TLR4/NF- κ B mediated inflammatory responses via modification of NAMPT/NAD.

Human gingival fibroblasts cytotoxicity

Human gingival fibroblasts were exposed to *Porphyromonas gingivalis* LPS and then treated with various concentrations of α -MG (0, 0.5, 1, 1.5, and 2 μ g/ml, 24 hr). The results from the investigation showed that α -MG attenuates the expression of IL-6 and interleukin-8 (IL-8) in *P. gingivalis* LPS-treated human gingival fibroblasts (37).

Therefore, α -MG inhibits the production of inflammatory cytokines IL-6 and IL-8 in *P. gingivalis* LPS-treated human gingival fibroblasts.

Liver failure

The hepatoprotective effect of α -MG (12.5, 25 mg/kg/day, 7 days, i.g.) on LPS/D-galactosamine-induced acute liver failure was studied and the findings revealed that α -MG protects the liver against the pathogenic effects of LPS/D-galactosamine by reducing hepatic MDA levels, serum alanine aminotransferase (ALT), aspartate transaminase (AST), TNF- α , IL-1 β , IL-6 levels, and recovering hepatic glutathione (GSH), SOD, and catalase (CAT) activities. They also discovered that α -MG suppressed LPS/D-galactosamine-induced TLR4 expression and NF- κ B activation while upregulating nuclear-related factor 2 (Nrf2) and heme oxygenase-1 expression (HO-1) (38).

Overall, α -MG protected against LPS/D-galactosamine-induced acute liver failure via activating Nrf2 and blocking the TLR4 signaling pathway. Taken together, α -MG might be a promising treatment for LPS/D-galactosamine-induced acute liver failure.

Table 1 also includes information on inflammation of the nervous system and murine macrophages.

β -Amyloid

Healthy neural and non-neural cells, such as skin and gut, release β -amyloid (A β), which circulates in both human cerebrospinal fluid and blood. Normally, low-density lipoproteins (LDL) receptor-related protein 1 transports β -amyloids through the blood-brain barrier. The clearance process by LDL receptor-related protein 1 is disturbed in Alzheimer's brains, causing the peptide to accumulate and aggregate. Vascular endothelial cells, astrocytes, or oligodendrocytes may all experience mitochondrial

malfunction and degeneration as a result of β -amyloid harmful effects. In Alzheimer's disease, mitochondrial dysfunctions are among the neurotoxic processes linked to β -amyloid (39). In an *in vivo* study on mice and *Caenorhabditis elegans*, A β maintains a low concentration and has a normal physiological function (40). Typically, LDL receptor-related protein 1 (LRP1) transports A β across the blood-brain barrier; however, factors like aging, oxidative stress, and gene mutation impair LRP1 clearance, resulting in peptide buildup and aggregation, which play a crucial role in the pathogenesis of Alzheimer's disease by causing neurotoxicity and cell death mostly through the generation of ROS (39, 41).

α -MG (0.5, 5, and 50 nM, 24 hr) attenuated the neurotoxicity caused by A β -(1-40) or A β -(1-42) oligomers (EC₅₀=3.89 nM and 4.14 nM, respectively) in cerebral cortex neurons of day 17 rat embryos, as determined by decreased cell viability and impaired neurite outgrowth. α -MG can also bind to A β , stabilizing the α -helical conformation. Furthermore, it can directly dissociate A β -(1-40) and A β -(1-42) oligomers and disrupt pre-formed fibrils while blocking fibril formation (42). It was demonstrated that preincubating SK-N-SH cells with mangosteen extract (50-400 μ g/ml) 30 min before exposure to A β -(1-42) could prevent the unpleasant effects of A β -(1-42) including cytotoxicity, increased intracellular ROS levels, caspase-3 activity, and changed the cellular proteome (43). Another investigation found that α -MG reduced the synthesis of A β 40 and A β 42. α -MG did not affect the expression of enzymes implicated in the nonamyloidogenic and amyloidogenic pathways, but it considerably reduced the activities of β -secretase and likely γ -secretase, with IC₅₀ values of 13.22 nmol/l and 16.98 nmol/l, respectively, in primary rat cerebral cortical neurons (44).

In primary cultured rat cortical cells, mangosteen pericarp water extract (<10 μ g/ml) prevented neurotoxicity and the generation of ROS caused by A β (25-35) or excitatory amino acids. Mangosteen pericarp water extract suppressed caspase-3 activation and DNA fragmentation in cells treated with A β (25-35) or N-methyl-D-aspartate, indicating an anti-apoptotic effect. Mangosteen pericarp water extract also decreased lipid peroxidation and scavenged 1,1-diphenyl-2-picrylhydrazyl radicals, indicating that it is an anti-oxidant. Mangosteen pericarp water also inhibited the activities of β -secretase and AChE. The Morris water maze test was used to assess the effect of mangosteen pericarp water extract

Table 1. Protective effects of mangosteen and its main constituents against LPS-induced nervous system inflammation and murine macrophage inflammation

Type of inflammation	Model	Protective agent	Results	Reference
Nervous system inflammation	young female C57BL/6j mice	α -mangostin (40 mg/kg/day, 14 days, PO)	↓ the levels of pro-inflammatory cytokine IL-6, COX-2, and translocator protein in the brain	(93)
Nervous system inflammation	BV-2 cell line	α -mangostin (100-500 nM, for 1 hr)	↓ LPS-induced pro-inflammatory cytokine production and iNOS expression ↓ microglia migration and phagocytosis in response to LPS, LPS-induced microglia-mediated neuronal dendritic damage, TLR4 expression, as well as TAK1 and NF- κ B activation	(94)
Nervous system inflammation	male C57BL/6 mouse	α -mangostin (50 mg/kg/day, 14 days, PO)	↓ LPS-induced microglial activation and neuroinflammation, as well as LPS-induced activation of the TLR4/TAK1/NF- κ B signaling pathway, LPS-induced dendritic damage, LPS-caused learning and memory impairments	(94)
Murine macrophage inflammation	RAW264.7 cells	<i>G. mangostana</i> peel extract (5, 10 and 20 μ g/ml) and its compounds (α -mangostin and γ -mangostin) (25, 50 and 75 μ M)	↓ COX-2, IL-6, IL-1, and NO production	(95)

IL-6: Interleukin-6; COX-2: Cyclooxygenase-2; iNOS: Inducible nitric oxide synthase; LPS: Lipopolysaccharide; TLR4: Toll-like receptor 4; TAK1: Transforming growth factor-activated kinase 1; NF- κ B: Nuclear factor- κ B; IL-1: Interleukin-1; NO: Nitric oxide

on memory impairment in scopolamine-treated mice. Mangosteen pericarp water extract administration (50, 100, or 300 mg/kg, 4 days, PO) considerably reduced the latency time to discover the platform and increased swimming time in the target quadrant (45).

As a result, α -MG could be used as a lead compound for prevention or decreasing the severity of Alzheimer's disease by mechanisms such as binding to A β and stabilizing the α -helical conformation, dissociating A β -(1-40) and A β -(1-42) oligomers, disrupting pre-formed fibrils while blocking fibril formation, preventing the increased intracellular ROS level, inhibiting the increased caspase-3 activity, and reducing the activities of β -secretase and likely γ -secretase.

3-Nitropropionic acid

Fungi (*Aspergillus flavus*; *Astragalus*, *Arthrinium*) and plants (*Indigofera endecapylla*) produce 3-nitropropionic acid, which is a natural poison. The succinate dehydrogenase (SDH; complex II) in the electron transportation chain, located inside the inner face of the mitochondrial membrane, is irreversibly inhibited by 3-nitropropionic acid, which causes neurotoxicity (46). Intoxication with 3-nitropropionic acid has no recognized antidote (47).

A study looked at the capacity of α -MG to scavenge ROS and its possible protective impact against the mitochondrial toxin 3-nitropropionic acid in primary cultures of cerebellar granule neurons. Singlet oxygen, superoxide anion, and peroxy nitrite anion were all shown to be scavenged by α -MG in a concentration-dependent manner. α -MG, on the other hand, could not scavenge hydroxyl radicals or hydrogen peroxide. α -MG was also able to reduce the neuronal mortality caused by 3-nitropropionic acid in a concentration-dependent manner. The reduction of 3-nitropropionic acid-induced ROS production was linked to this protective effect (48).

Consequently, α -MG decreased the generation of ROS caused by mitochondrial toxin 3-nitropropionic acid *in vitro*. Therefore, more research is needed to see if α -MG can penetrate the blood-brain barrier and acquire adequate bioavailability in the brain to trigger a protective response against neurodegenerative diseases such as Alzheimer's disease and Parkinson's disease.

Mangosteen and its compounds against chemical agents induced toxicity

This section discusses the antidotal and protective actions of mangosteen and its compounds against a variety of chemical agents that cause toxicity, besides their significant protective mechanisms.

Anti-cancer drugs

Besides mangosteen antidotal effects against chemotherapy agents-induced toxicity, it is interesting to mention that mangosteen phytochemicals (extracts, α -MG, β -MG, mangaxanthone B, and mangaphenone) have been shown in several *in vitro* and *in vivo* studies to suppress the growth and spread of cancer cells and to have an anti-proliferative and apoptosis-inducing impact on several human cancers, including breast, lung, liver, colon, oral, skin, leukemia, head and neck, prostate, and cervical cancers (49, 50).

Doxorubicin

Doxorubicin, one of the first two anthracyclines identified in *Streptomyces peucetius*, was isolated for the first time in the early 1960s. This drug is often used in chemotherapy to treat a variety of cancers such as carcinomas, sarcomas, hematological cancers, as well as solid tumors in children (20, 51). Despite its efficiency, it has a wide range of harmful side effects, the majority of which are a result of its intrinsic pro-oxidant activity. Doxorubicin is toxic to normal cells, including brain tissue (52).

The protective effect of a xanthone derivative of *G. mangostana* xanthones (200 mg/kg, single dose, IP) against doxorubicin-induced neuronal toxicity in male B6C3 mice was investigated and the findings revealed that xanthone could prevent doxorubicin from generating an increase in TNF- α , inducible nitric oxide synthase (iNOS) protein levels, and NO production in mononuclear cells. It also inhibited doxorubicin-mediated changes in pro- and anti-apoptotic proteins, reduced caspase-3 activity, and terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL)-positive apoptotic cells, and reduced doxorubicin-mediated increases in protein carbonyl, 3-nitrotyrosine, and protein-bound 4-hydroxy-2'-nonenal (4-HNE) in brain tissues (53).

In conclusion, xanthone derivatives derived from mangosteen may be useful in avoiding tissue harm caused by ROS-producing chemotherapy medicines.

Bleomycin

Bleomycin is the generic name for a class of antibiotics isolated from the *Streptococcus verticillus* bacteria (54). Bleomycin is an important aspect of the treatment for a variety of tumors that can be cured, but it has a major drawback: lung damage like pulmonary fibrosis (55).

An *in vivo* study showed that α -MG administration (10 mg/kg/day, 14 days, i.g.) to male C57/BL6 mice considerably reduced bleomycin-induced extracellular matrix deposition in lung tissues. Furthermore, α -MG has been shown to reduce α -smooth muscle actin (α -SMA) and collagen I protein expression as well as its mRNA levels. In addition, α -MG suppressed the TGF β 1/Smad2/3 pathway and affected matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 (TIMP-1) protein expression in lung tissues. *In vitro* data showed that α -MG (1-50 nM, for 48 hr) enhanced phosphorylated-adenosine 5'-monophosphate-activated protein kinase (p-AMPK)/AMPK but decreased the protein expression levels of α -SMA and collagen I as well as nicotinamide adenine dinucleotide phosphate oxidase-4 in activated primary lung fibroblasts (55).

Overall, α -MG therapy reduced collagen formation, altered the redox status of lung fibroblasts, and alleviated bleomycin-induced pulmonary fibrosis in mice by targeting AMPK signaling (Figure 5).

Cisplatin

Cisplatin is a chemotherapy drug that is used to treat a variety of solid tumors, including testicular, ovarian, head, neck, colorectal, bladder, and lung cancers (56). Nephrotoxicity is the most common dose-limiting adverse effect of cisplatin (57).

α -MG (5 μ M, for 24 hr) protects rats against cisplatin-induced kidney injury by diminishing the increase in ROS level, apoptotic cell death, GSH depletion, and increased p53 expression in Lilly laboratory culture porcine kidney

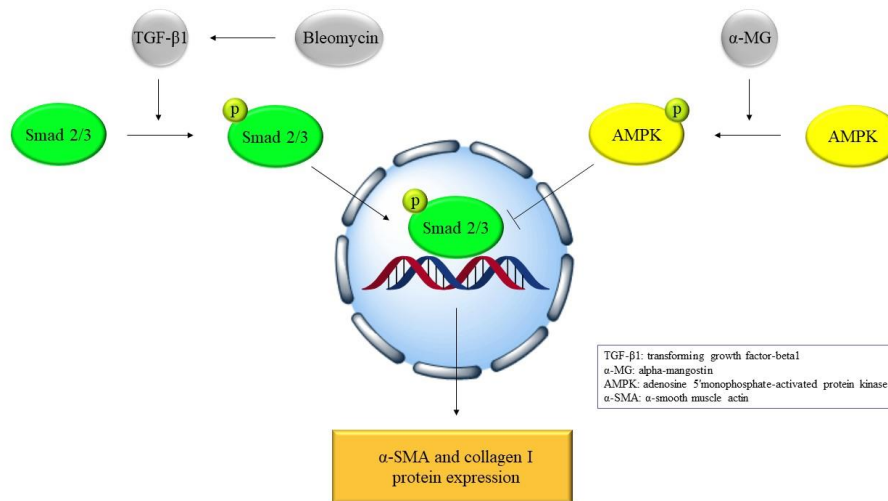


Figure 5. α -Mangostin antidotal molecular mechanism against bleomycin-induced pulmonary fibrosis

(LLC-PK1) cells (58).

In another study, α -MG (2.5 μ g/ml, for 48 hr) was found to protect HCT 116 human colorectal cancer cells from cisplatin-induced cytotoxicity due to the inhibition of ROS generation (59). Hence, this compound may have a cytoprotective impact against oxidative stress, irradiation, and chemical carcinogens, which could help prevent diseases like cancer.

In a human embryonic kidney (HEK293) cell model, the protective effect of α -MG against cisplatin-induced cytotoxicity was studied. It was discovered that α -MG (5, 10, 20, and 40 μ M, for 24 hr) reduced cisplatin-induced cell death by lowering MDA levels and increasing GSH content. Following cisplatin encounter, α -MG dramatically reduced ROS overproduction, restored phosphoinositide 3-kinases (PI3K)/ protein kinase B (Akt) activation, and downregulated the c-Jun NH2-terminal kinase (JNK) pathways. Following that, α -MG significantly prevented the cleavage of caspases and poly-ADP-ribose polymerase, implicating ROS-mediated apoptotic pathways generated by cisplatin (60).

Cisplatin induces nephrotoxicity, which is complex and appears to be connected to free radical-induced damage (oxidative/nitrosative stress), inflammatory responses, fibrotic pathways, and a reduction in catalase activity. The renoprotective effect of α -MG on cisplatin-induced nephrotoxicity in male Wistar rats was investigated in another investigation. α -MG (12.5 mg/kg/day, i.g., for 10 days) reduced renal dysfunction, structural damage, oxidative/nitrosative stress, catalase expression, and TNF- α and TGF β mRNA levels (53).

The protective effects of α -MG on cisplatin-induced damage in proximal tubule LLC-PK1 cells were investigated, and it was discovered that α -MG co-incubation (4 and 5 μ M) inhibited cisplatin-induced cell death. Furthermore, α -MG reduced cisplatin-induced reductions in cell respiratory states, the maximum capacity of the electron transfer system (E), and the respiration associated with oxidative phosphorylation. Cisplatin also reduced the protein levels of voltage-dependent anion channels and mitochondrial complex subunits, mitochondrial morphology changes, and mitochondrial mass (61).

In conclusion, α -MG attenuates cisplatin-induced

nephrotoxicity or cytotoxicity by mechanisms like decreasing ROS level, increasing GSH content, and reducing the protein levels of mitochondrial complex subunits, mitochondrial morphology changes, and mitochondrial mass. Therefore, α -MG could be used as a protective agent against cisplatin toxicity.

Streptozotocin

Streptozotocin, a monofunctional nitrosourea derivative isolated from *Streptomyces achromogenes* in 1960, has broad-spectrum antibacterial action and antineoplastic characteristics, although its diabetogenic activities were not found until 1963. Through its damaging effects on pancreatic cells, it is commonly used to cause diabetes mellitus in experimental animals and due to its selective toxicity, it is also utilized to treat β -cell pancreatic tumors (62, 63).

The hypoglycemic activity of *G. mangostana* pericarp ethanolic extract in normoglycemic and streptozotocin-induced diabetic rats was evaluated in a study. Mangosteen pericarp ethanolic extract administration (50, 100, and 200 mg/kg, single-dose study (1 day) and multiple-dose study (29 days), PO) to male normoglycemic and streptozotocin-induced diabetic Sprague-Dawley rats remarkably reduced the blood glucose level. Furthermore, in the multiple-dose research, mangosteen pericarp ethanolic extract considerably increased the body weight of the rats when compared to the diabetic control group. Triglycerides, total cholesterol, LDL, very-low-density lipoprotein (VLDL), serum glutamic oxaloacetic transaminase (SGOT), serum glutamic pyruvic transaminase (SGPT), urea, and creatinine were all significantly reduced by this extract, while high-density lipoprotein (HDL) and total protein were significantly increased. In diabetic rats, there was a slight increase in the population of β -cells (26).

The anti-glycemic and anti-hepatotoxic benefits of mangosteen vinegar rind on a high-fat diet (HFD)/ single dose STZ induced male institute of cancer research (ICR) diabetic mice are the topic of another investigation. When compared to the untreated diabetic control group, mangosteen vinegar rind administration (100 and 200 mg/kg, 1 week, PO) improved the levels of glucose, hepatic glycogen, lipid profile (lower total cholesterol, triglycerides,

LDL levels, and higher HDL levels), oxidative stress (MDA level), anti-oxidant enzyme activity (SOD and CAT), and liver function biomarkers (ALT and AST) in HFD/streptozotocin-induced type II diabetic mouse models (64).

The anti-apoptotic and reno-protective properties of mangosteen vinegar rind aqueous extract against HFD/streptozotocin-induced type II diabetes nephropathy in ICR mice were investigated, and the findings revealed that acute mangosteen vinegar rind therapy (100 and 200 mg/kg, 1 week, PO) has a reno-protective effect on type II diabetes via reducing oxidative stress and apoptosis. This protective effect could be related to improvements in glucose level and lipid metabolism, mitochondrial integrity, oxidative stress reduction, inhibition of lipid peroxidation and inflammation, insulin sensitivity enhancement, and modulation of numerous apoptotic pathways (65).

Another study assessed the renal protective effects of γ -MG in male streptozotocin-induced diabetic BALB/c mice, and the results showed that γ -MG administration (1, 2, and 4 mg/kg, 2 weeks) was able to significantly lower plasma blood urea nitrogen (BUN) and creatinine, as well as ameliorate diabetic mice's impaired renal proximal tubular cells (66).

Therefore, the constituents of *G. mangostana* could be a potential candidate for the management of hyperglycemia, hyperlipidemia, and diabetic nephropathy through several mechanisms like improvement in the level of glucose, hepatic glycogen, lipid profile (lower total cholesterol, triglycerides, LDL levels, and higher HDL level), oxidative stress (MDA level), anti-oxidant enzyme activity (SOD and CAT), liver function biomarkers (ALT and AST) and mitochondrial integrity, inhibition of lipid peroxidation and inflammation, insulin sensitivity enhancement, and modulation of numerous apoptotic pathways and lowering plasma BUN and creatinine.

Antibiotics

Isoniazid

Isoniazid (isonicotinic acid hydrazide) is one of the main therapies for tuberculosis, which is caused by infection with *Mycobacterium tuberculosis*. Isoniazid is not destructive to the bacterial cell itself, but it is a prodrug that is activated by the mycobacterial catalase-peroxidase enzyme (67). Hepatotoxicity and a potentially deadly liver injury are linked to isoniazid usage. Hepatotoxicity is frequently accompanied by nausea and right upper quadrant stomach discomfort, but it can also be asymptomatic, and the diagnosis is based on bilirubin and SGPT levels in the serum (68).

The impact of the ethanol extract of *G. mangostana* peel on isoniazid-induced liver damage in rats was examined in the research. The findings of administering mangosteen peel ethanol extract (250 and 500 mg/kg/day, 35 days) to male Wistar rats revealed that 500 mg/kg of ethanol extract of *G. mangostana* peel reduced isoniazid-induced liver damage in rats by lowering TGF- β 1, SGPT level, and liver fibrosis (69).

As a result, ethanol extracts of *G. mangostana* peel prevent isoniazid-induced liver damage in rats by lowering TGF- β 1, SGPT levels, and liver fibrosis.

Analgesics

Acetaminophen

Acetaminophen is one of the most widely used and well-

tolerated pain relievers in the world. Its exact mechanism of action is unknown, however, it appears to selectively inhibit cyclooxygenase (COX) in the brain, and its capacity to relieve fever and discomfort is a result of this effect. In the central nervous system, it may also suppress PGs production. Acetaminophen has an antipyretic effect by acting directly on the hypothalamus (70). Because of its widespread availability, acetaminophen is commonly linked to overdoses, both deliberate and unintentional, resulting in severe liver damage and even acute liver failure (71).

α -MG treatment (100, 200 mg/kg/day, 7 days, i.g.) reduces the adverse effects of overdose acetaminophen-induced hepatic injury in male ICR mice, partly by restoring anti-oxidative activity and modulating inflammation, apoptosis, and autophagy via increasing serum aminotransferase levels and GSH content, reducing MDA, inhibiting increases in TNF- α and IL-1 β , and the protein expression of autophagy-related microtubule-associated protein light chain 3 and BCL2/adenovirus E1B protein-interacting protein 3. Furthermore, the protective effect of α -MG on acetaminophen-induced acute liver injury might be attributed to changes in the liver Akt/mTOR signaling pathway (72).

Moreover, α -MG protective effect against acetaminophen-induced acute liver injury was evaluated in another study in male ICR mice. α -MG (12.5 and 25 mg/kg/day, 6 days, i.g.) reduced serum levels of ALT, AST, TNF- α , IL-1 β , IL-6, and hepatic MDA, while restoring hepatic GSH, SOD, and CAT activity. Moreover, α -MG pretreatment effectively decreased acetaminophen-induced phosphorylation of ERK, JNK, and p38 MAPK, which was associated with alterations in TNF- α , IL-1 β , and IL-6 levels; phosphorylation of I κ B α and translocation of NF- κ Bp65 were also suppressed by α -MG (38).

In another study, totophyllin A (50 and 100 mg/kg/day, 5 days) protected male BALB/c mice against acetaminophen-induced hepatic damage through its significant anti-oxidant (reversing the elevation of MDA and 4-hydroxynonenal levels, recovering hepatic GSH, SOD, and CAT, and decreasing the level of NOx) and anti-inflammatory effects, which increased Nrf2 activation and disrupted the NF- κ B pathway (73).

Consequently, α -MG and totophyllin A reduce acetaminophen-induced liver damage by mechanisms like lowering serum levels of ALT, AST, TNF- α , IL-1 β , IL-6, and hepatic MDA, boosting Nrf2 activation, and disrupting the NF- κ B pathway. As a result, α -MG and totophyllin A may be used as an antidote to acetaminophen toxicity.

β -Adrenergic agonist

Isoproterenol

Isoproterenol is a nonselective sympathomimetic β -adrenergic agonist manufactured synthetically. It is most commonly used to treat bradycardia, thioridazine-induced torsade de pointes, and heart block. Isoproterenol produces highly cytotoxic free radicals through auto-oxidation, which causes oxidative stress, resulting in gradual mitochondrial damage and changes in cardiac biochemical parameters, leading to heart injury (74, 75).

Isoproterenol induction resulted in a considerable rise in the activity of serum and cardiac lysosomal hydrolases (β -D-glucuronidase, β -D-galactosidase, β -D-N-acetylglucosaminidase, acid phosphatase, and cathepsin-D)

in adult male Wistar rats. In the hearts of isoproterenol-administered rats, the aberrant activity of membrane-bound phosphatases ($\text{Na}^+\text{-K}^+$ ATPase, Ca^{2+} ATPase, and Mg^{2+} ATPase) was noted, as well as a considerable rise in cardiac sodium and calcium levels with a decrease in potassium levels. TNF- α and COX-2 expression in the heart of isoproterenol-intoxicated rats were considerably higher. When compared to the isoproterenol-intoxicated group of rats, pre-co-treatment with α -MG (200 mg/kg, 8 days, PO) considerably reduced these anomalies and restored levels to near normalcy (76).

The induction of adult male Wistar rats with isoproterenol caused a considerable increase in lipid peroxidation, serum marker enzymes (lactate dehydrogenase (LDH), creatine phosphokinase (CPK), glutamate oxaloacetate transaminase (GOT), and glutamate pyruvate transaminase (GPT)), and a significant decrease in endogenous anti-oxidant activity (SOD, CAT, glutathione peroxidase (GPx), glutathione-S-transferase (GST), and GSH). When compared to the individual treatment groups, pre-treatment with α -MG (200 mg/kg/day, 6 days, PO) before isoproterenol administration and 2 days after ISO administration considerably reduced these alterations (77).

The potential role of mangosteen in isoproterenol-induced myocardial infarction in adult male albino rats was investigated. The administration of mangosteen (18 mg/200 mg, p.o.) resulted in a partial improvement in heart muscle fibers as well as a reduction in inflammatory cellular infiltration (78).

As a result of these findings, mangosteen and α -MG can be used as a cardiotoxic preventive against β -adrenergic catecholamine-induced myocardial toxicity and oxidative stress.

Anticholinergic

Scopolamine

Scopolamine is a muscarinic receptor blocker that impairs cholinergic neurotransmission, causing memory loss in Alzheimer's disease patients (79).

γ -MG (3~10 μM , for 24 hr) protected rat cerebrocortical cells from H_2O_2 - or xanthine/xanthine oxidase-induced oxidative neuronal death and reduced the formation of ROS generated by these oxidative insults. It also prevented H_2O_2 -induced DNA fragmentation and caspase-3 and 9 activations, confirming its antiapoptotic properties. Furthermore, γ -MG was discovered to efficiently prevent lipid peroxidation, the generation of 1,1-diphenyl-2-picrylhydrazyl radicals, and the activity of β -secretase. The effect of γ -MG on scopolamine-induced memory impairment in ICR mice was assessed using the passive avoidance test, and it was discovered that γ -MG (10 and 30 mg/kg, PO) significantly reduced scopolamine-induced memory impairment (80).

Mangosteen extract (200 g/ml, for 3 hr) might partially counteract the effects of H_2O_2 on cell survival, ROS level, and caspase-3 activity in SK-N-SH cell cultures. Mangosteen extract (200, 400, or 800 g/ml, for 3 hr) lowered SK-N-SH cells' AChE activity to around 60% of the control. The Morris water maze and passive avoidance tests were utilized to examine the memory of male ICR mice in an *in vivo* investigation. Mangosteen extract (100 mg/kg/day, PO) for the passive avoidance test and the Morris water maze test improved the animal's memory and antagonized the effect of

scopolamine on memory. The mangosteen extract therapy counteracted the rise in ROS level and caspase-3 activity in the brains of scopolamine-treated mice (81).

Therefore, γ -MG and mangosteen extract might be used as a preventive treatment against scopolamine-induced memory impairment through mechanisms such as reduction in ROS production and prevention of caspase-3 and 9 activations.

Thioacetamide

Thioacetamide contains thiono-sulfur and has been employed as a fungicide, an organic solvent, a rubber accelerator, and a motor oil stabilizer (82). Thioacetamide has been shown to cause liver fibrosis and cirrhosis in experimental animals (15).

In male Wistar rats, the effects of α -MG on thioacetamide-induced liver cirrhosis were examined. α -MG (100 mg/kg, 3 times per week, 4 weeks, IP) reduced fibrotic nodules and lowered AST and ALT levels in the blood. It also decreased the risk of liver fibrosis by lowering p53 expression (83).

Thioacetamide caused histologically detectable liver damage and fibrosis in rats in another investigation. It also elevated TGF- β 1, α -SMA, and TIMP-1 immunohistochemically detectable levels. The effects of thioacetamide treatment alone were avoided or ameliorated by co-administration of α -MG (5 mg/kg, 3 times per week, 4 weeks, IP) with thioacetamide (84).

Consequently, α -MG acts as a protective agent against thioacetamide-induced liver fibrosis and cirrhosis by reducing fibrotic nodules, lowering AST and ALT levels, lowering p53 expression, and ameliorating the elevated TGF- β 1, α -SMA, and TIMP-1.

Lead

Lead is found all over nature. It may be found in a variety of forms, including bullets, inorganic compounds like lead oxide, lead chromate, and lead sulfide, and organic compounds like tetraethyl lead (85). Abdominal discomfort, constipation, anemia, hearing loss, exhaustion, neuropathy and neurotoxicity, encephalopathy, renal disorders, abortions, osteopenia, and even mortality are among the symptoms of lead poisoning (86).

Xanthone (100 and 200 mg/kg, 38 days, PO) alleviated lead-induced neurotoxicity in ICR mice, in part by suppressing oxidative damage and reversing AChE activity. Forced swimming and Morris water maze tests have shown that it has substantial protective benefits against lead-induced learning deficits and memory loss (87).

The potential preventive benefits of xanthones against lead acetate-induced chronic renal disease in ICR mice were investigated. Xanthones (100 and 200 mg/kg/day, 38 days, PO) scavenged the radicals 2,2-diphenyl-1-picrylhydrazyl, superoxide, hydroxyl, and NO. Lead acetate-induced oxidative stress, renal dysfunction, inflammatory markers (plasma TNF- α concentration and protein expression of TNF- α , COX-2, and iNOS in kidney tissue), and kidney apoptosis were reduced by co-treatment with xanthones. The tissue architecture was significantly enhanced as a result of the therapy. An *in-silico* prediction of activity investigation revealed that xanthones' protective effects may be owing to their ability to activate Nrf2, control intracellular $[\text{Ca}^{2+}]$, and downregulate the NF- κ B, MAPK pathway (88).

As a result, by reducing oxidative damage, reversing

Table 2. Protective effects of mangosteen and its main constituents against chemical toxicants

Compound	Study model	Protective agent	Results	Reference
Rotenone	<i>In vivo</i> , Sprague Dawley rats	α -mangostin (10 mg/kg, 21 days, IP)	↑ antioxidant enzyme levels ↓ phosphorylated α -synuclein levels, TH+ dopaminergic neuronal loss in the substantia nigra pars compacta, memory impairments	(31)
Paraquat and NaNO ₂	<i>In vitro</i> , Chinese hamster lung cells	Xanthone (0, 0.1, 0.5, 1, 5, 10 μ M, 30 min prior to paraquat and NaNO ₂ exposure)	↓ sister chromatid exchange and decreased cell cycle rate	(96)
Iodoacetate	<i>In vitro</i> , cerebellar granule neurons	α -mangostin (8, 12, and 14 μ M, for 16 hrs)	↑ antioxidant induction of heme oxygenase-1 ↓ ROS generation	(97)
3,8. tert-Butyl hydroperoxide	<i>In vitro</i> , human normal hepatocytes (HL-7702)	γ -mangostin (0.63, 1.25, 2.50, and 5.00 μ M)	↑ SOD amount ↓ lipid peroxidation, GSH levels, loss of mitochondrial membrane potential	(98)
Cigarette smoke	<i>In vivo</i> , rats	mangosteen peel extract (150, 300, and 600 mg/kg, 21 days, PO)	↑ SOD activity ↓ MDA level	(99)
Iodixanol	<i>In vitro</i> , LLC-PK1 cells	α -mangostin (2.5 and 5 μ M, 3 hr)	↑ cell viability ↓ phosphorylation of p38, ERK, and caspase-3 cleavage	(100)

ERK: extracellular-signal-regulated kinase; GSH: glutathione; MDA: malondialdehyde; ROS: reactive oxygen species; SOD: superoxide dismutase

AChE activity, and lowering inflammatory markers, xanthones protect against lead and lead acetate-induced toxicity.

The protective effects of mangosteen and its main constituents against some other chemical toxicants can be found in Table 2.

Mangosteen's toxicity

In previous research, the lethal dose (LD₅₀) for intraperitoneal administration of the crude methanolic extract to female BALB/c mice was determined to be 1000 mg/kg, while the appropriate dose for short-term studies should be 200 ≤ mg/kg (27). Rahmayanti *et al.* determined that the LD₅₀ of the ethyl acetate fraction from mangosteen pericarp extract was >15.480 mg/kg after oral administration to female Sprague-Dawley rats (89). Another study found that oral administration of mangosteen skin extract to BALB/c mice in doses of up to 5000 mg/kg was not toxic and could be used as a natural herbal medicine (90). It has also been reported that intraperitoneal administration of α -MG and mangosteen extract to mice resulted in LC₅₀ of 150 and 231 mg/kg, respectively (91). The acute and subchronic toxicity of a tannin-rich extract from the mangosteen fruit pericarp was studied in Swiss albino mice via intragastric administration, and the results showed that the extract at the doses tested (2 and 5 g/kg for acute toxicity and 400, 600, and 1,200 mg extract/kg for subchronic toxicity) had no significant negative effects in the experimental animals (92).

Conclusion

In this review, we summarized the findings from several *in vitro* and *in vivo* studies on *G. mangostana* and its principal components, particularly α -MG, γ -MG, and tophoyllin A, to highlight mangosteen's antidotal and protective activities against biological and chemical toxicities. Mangosteen protects against a variety of biological toxins, including LPS, ovalbumin, β -amyloid, and 3-nitropropionic acid, through different mechanisms like anti-inflammatory and anti-apoptotic properties, histological condition improvement, and reduction of oxidative stress. It also has a protective role against chemical toxic agents including anti-cancer drugs, β -adrenergic agonists, anticholinergic, isoniazid, acetaminophen, thioacetamide, and lead. Mangosteen and its constituents exert their effects primarily through different mechanisms such as anti-oxidant, radical scavenging, anti-apoptotic properties, anti-inflammatory

effects, and the regulation of the renal, hepatic, and cardiac enzymes. All current evidence shows that *G. mangostana* and its components have extremely promising benefits and that more study, including clinical trials, is required.

Acknowledgment

We thank the Vice-Chancellor of Research, Mashhad University of Medical Sciences, Iran.

Authors' Contributions

HH Study conception, design and supervision of the research; MGR Critical revision of the paper, supervision of the research; AMZ Preparation of the original draft. All authors have agreed to the contents and approved the final version for publication

Data availability statement

The data that support the findings of this study are available from the corresponding author upon responsible request.

Conflicts of Interest

The authors declare no conflicts of interest.

References

- Najafi N, Ghasemzadeh Rahbardar M, Hosseinzadeh H, Hayes AW, Karimi G. Chemical agents protective against rotenone-induced neurotoxicity. *Toxicol Environ Chem* 2022; 104: 149-175.
- Wu H, Huang J. Drug-induced nephrotoxicity: Pathogenic mechanisms, biomarkers and prevention strategies. *Curr Drug Metab* 2018; 19: 559-567.
- Ghasemzadeh Rahbardar M, Hemadeh B, Razavi BM, Eisvand F, Hosseinzadeh H. Effect of carnosic acid on acrylamide induced neurotoxicity: *in vivo* and *in vitro* experiments. *Drug Chem Toxicol* 2020; 45: 1528-1535.
- Rahbardar MG, Farmad HC, Hosseinzadeh H, Mehri S. Protective effects of selenium on acrylamide-induced neurotoxicity and hepatotoxicity in rats. *Iran J Basic Med Sci* 2021; 24: 1041-1049.
- Sampath PD, Vijayaragavan K. Ameliorative prospective of alpha-mangostin, a xanthone derivative from *Garcinia mangostana* against beta-adrenergic catecholamine-induced myocardial toxicity and anomalous cardiac TNF-alpha and COX-2 expressions in rats. *Exp Toxicol Pathol* 2008; 60: 357-364.
- Dwivedi S, Kushalan S, Paithankar JG, D'Souza LC, Hegde S, Sharma A. Environmental toxicants, oxidative stress and health adversities: interventions of phytochemicals. *J Pharm Pharmacol* 2022; 74: 516-536.
- Valko M, Morris H, Cronin M. Metals, toxicity and oxidative stress. *Curr Med Chem* 2005; 12:1161-1208.

8. Bondy SC. Metal toxicity and neuroinflammation. *Curr Opin Toxicol* 2021; 26: 8-13.
9. Franco R, Sánchez-Olea R, Reyes-Reyes EM, Panayiotidis MI. Environmental toxicity, oxidative stress and apoptosis: manage a trois. *Mutat Res Genet Toxicol Environ Mutagen* 2009; 674: 3-22.
10. Kiraz Y, Adan A, Kartal Yandim M, Baran Y. Major apoptotic mechanisms and genes involved in apoptosis. *Tumor Biol* 2016; 37: 8471-8486.
11. Avila MS, Siqueira SRR, Ferreira SMA, Bocchi EA. Prevention and treatment of chemotherapy-induced cardiotoxicity. *Methodist Debakey Cardiovasc J* 2019; 15: 267-273.
12. Fang C-y, Lou D-y, Zhou L-q, Wang J-c, Yang B, He Q-j, et al. Natural products: potential treatments for cisplatin-induced nephrotoxicity. *Acta Pharmacol Sin* 2021; 42: 1951-1969.
13. Barron AJ, Zaman N, Cole GD, Wensel R, Okonko DO, Francis DP. Systematic review of genuine versus spurious side-effects of beta-blockers in heart failure using placebo control: Recommendations for patient information. *Int J Cardiol* 2013; 168: 3572-3579.
14. Naraki K, Rameshrad M, Hosseinzadeh H. Protective effects and therapeutic applications of ellagic acid against natural and synthetic toxicants: A review article. *Iran J Basic Med Sci* 2022; 25: 1402-1415.
15. Alavi MS, Fanoudi S, Ghasemzadeh Rahbardar M, Mehri S, Hosseinzadeh H. An updated review of protective effects of rosemary and its active constituents against natural and chemical toxicities. *Phytother Res* 2021; 35: 1313-1328.
16. Hosseini M, Pkan P, Rakhshandeh H, Aghaie A, Sadeghnia HR, Rahbardar MG. The effect of hydro-alcoholic extract of citrus flower on pentylentetrazole and maximal electroshock-induced seizures in mice. *World Appl Sci J* 2011; 15: 1104-1109.
17. Yarmohammadi F, Rahbardar MG, Hosseinzadeh H. Effect of eggplant (*Solanum melongena*) on the metabolic syndrome: A review. *Iran J Basic Med Sci* 2021; 24: 420-427.
18. Boskabady MH, Vatanprast A, Parsee H, Ghasemzadeh M. Effect of aqueous-ethanolic extract from *Rosa damascena* on guinea pig isolated heart. *Iran J Basic Med Sci* 2011; 14: 116-121.
19. Ardakanian A, Rahbardar MG, Omidkhoda F, Razavi BM, Hosseinzadeh H. Effect of alpha-mangostin on olanzapine-induced metabolic disorders in rats. *Iran J Basic Med Sci* 2022; 25: 198-207.
20. Eisvand F, Imenshahidi M, Ghasemzadeh Rahbardar M, Tabatabaei Yazdi SA, Rameshrad M, Razavi BM, et al. Cardioprotective effects of alpha-mangostin on doxorubicin-induced cardiotoxicity in rats. *Phytother Res* 2022; 36: 506-524.
21. Ghasemzadeh A, Jaafar HZ, Baghdadi A, Tayebi-Meigooni A. Alpha-mangostin-rich extracts from mangosteen pericarp: optimization of green extraction protocol and evaluation of biological activity. *Molecules* 2018; 23: 1852.
22. Im A, Kim Y-M, Chin Y-W, Chae S. Protective effects of compounds from *Garcinia mangostana* L. (mangosteen) against UVB damage in HaCaT cells and hairless mice. *Int J Mol Med* 2017; 40: 1941-1949.
23. Ming-Hui W, Zhang K-J, Qin-Lan G, Xiao-Ling B, Jin-Xin W. Pharmacology of mangostins and their derivatives: A comprehensive review. *Chin J Nat Med* 2017; 15: 81-93.
24. Geetha R, Roy A, Lakshmi T. Evaluation of antibacterial activity of fruit rind extract of *Garcinia mangostana* Linn on enteric pathogens-an *in vitro* study. *Asian J Pharm Clin Res* 2011; 4: 115-118.
25. Gopalakrishnan G, Banumathi B, Suresh G. Evaluation of the antifungal activity of natural xanthenes from *Garcinia mangostana* and their synthetic derivatives. *J Nat Prod* 1997; 60: 519-524.
26. Taher M, Zakaria TMFST, Susanti D, Zakaria ZA. Hypoglycaemic activity of ethanolic extract of *Garcinia mangostana* Linn. in normoglycaemic and streptozotocin-induced diabetic rats. *BMC Complement Altern Med* 2016; 16: 1-12.
27. Kosem N, Ichikawa K, Utsumi H, Moongkarndi P. *In vivo* toxicity and antitumor activity of mangosteen extract. *J Nat Med* 2013; 67: 255-263.
28. Moongkarndi P, Kosem N, Kaslungka S, Luanratana O, Pongpan N, Neungton N. Antiproliferation, antioxidation and induction of apoptosis by *Garcinia mangostana* (mangosteen) on SKBR3 human breast cancer cell line. *J Ethnopharmacol* 2004; 90: 161-166.
29. Liu Q-Y, Wang Y-T, Lin L-G. New insights into the anti-obesity activity of xanthenes from *Garcinia mangostana*. *Food Funct* 2015; 6: 383-393.
30. Ghasemzadeh Rahbardar M, Razavi BM, Hosseinzadeh H. Investigating the ameliorative effect of alpha-mangostin on development and existing pain in a rat model of neuropathic pain. *Phytother Res* 2020; 34: 3211-3225.
31. Parkhe A, Parekh P, Nalla LV, Sharma N, Sharma M, Gadepalli A, et al. Protective effect of alpha mangostin on rotenone induced toxicity in rat model of Parkinson's disease. *Neuroscience Lett* 2020; 716: 134652.
32. Jo S, Wu EL, Stuhlsatz D, Klauda JB, MacKerell AD, Widmalm G, et al. Lipopolysaccharide membrane building and simulation. *Methods Mol Biol* 2015; 1273: 391-406.
33. Miller KA, Suresh Kumar E, Wood SJ, Cromer JR, Datta A, David SA. Lipopolysaccharide sequestrants: structural correlates of activity and toxicity in novel acylhomospermines. *J Med Chem* 2005; 48: 2589-2599.
34. Zou W, Yin P, Shi Y, Jin N, Gao Q, Li J, et al. A novel biological role of α -mangostin via TAK1-NF- κ B pathway against inflammatory. *Inflammation* 2019; 42: 103-112.
35. Yang Z, Yin Q, Olatunji OJ, Li Y, Pan S, Wang DD, et al. Activation of cholinergic anti-inflammatory pathway involved in therapeutic actions of α -mangostin on lipopolysaccharide-induced acute lung injury in rats. *Int J Immunopathol Pharmacol* 2020; 34: 2058738420954941.
36. Tao M, Jiang J, Wang L, Li Y, Mao Q, Dong J, et al. α -Mangostin alleviated lipopolysaccharide induced acute lung injury in rats by suppressing NAMPT/NAD controlled inflammatory reactions. *Evid Based Complement Altern Med* 2018; 2018: 5470187.
37. Yiemwattana I, Kaomongkolgit R. Alpha-mangostin suppresses IL-6 and IL-8 expression in *P. gingivalis* LPS-stimulated human gingival fibroblasts. *Odontology* 2015; 103: 348-355.
38. Fu T, Wang S, Liu J, Cai E, Li H, Li P, et al. Protective effects of α -mangostin against acetaminophen-induced acute liver injury in mice. *Eur J Pharmacol* 2018; 827: 173-180.
39. Tillement L, Lecanu L, Papadopoulos V. Alzheimer's disease: effects of β -amyloid on mitochondria. *Mitochondrion* 2011; 11: 13-21.
40. Morley JE, Farr SA, Nguyen AD, Xu F. What is the physiological function of amyloid-beta protein? *J Nutr Health Aging* 2019; 23: 225-226.
41. Huang YR, Liu RT. The toxicity and polymorphism of β -amyloid oligomers. *Int J Mol Sci* 2020; 21: 4477.
42. Wang Y, Xia Z, Xu JR, Wang YX, Hou LN, Qiu Y, et al. α -mangostin, a polyphenolic xanthone derivative from mangosteen, attenuates β -amyloid oligomers-induced neurotoxicity by inhibiting amyloid aggregation. *Neuropharmacology* 2012; 62: 871-881.
43. Moongkarndi P, Srisawat C, Saetun P, Jantaravid J, Peerapittayamongkol C, Soi-ampornkul R, et al. Protective effect of mangosteen extract against beta-amyloid-induced cytotoxicity, oxidative stress and altered proteome in SK-N-SH cells. *J Proteome Res* 2010; 9: 2076-2086.
44. Zhao LX, Wang Y, Liu T, Wang YX, Chen HZ, Xu JR, et al. α -Mangostin decreases β -amyloid peptides production via modulation of amyloidogenic pathway. *CNS Neurosci Ther* 2017; 23: 526-534.
45. Oh Y, Do HTT, Kim S, Kim Y-M, Chin Y-W, Cho J. Memory-enhancing effects of mangosteen pericarp water extract through antioxidative neuroprotection and anti-apoptotic action. *Antioxidants* 2021; 10: 34.
46. Torabi A, Joneidi M, Mohammadzadeh I, Abdollahifar M-a, Khatmi A, Ezi S, et al. The effect of 3-nitropropionic acid on behavioral dysfunction, neuron loss and gliosis in the brain of adult male rats: The case of prefrontal cortex, hippocampus and

- the cerebellum. *Toxicol* 2020; 183: 44-50.
47. Bendiksen Skogvold H, Yazdani M, Sandås EM, Østebø Vassli A, Kristensen E, Haarr D, et al. A pioneer study on human 3-nitropropionic acid intoxication: Contributions from metabolomics. *J Appl Toxicol* 2021; 45: 818-829.
 48. Pedraza-Chaverri J, Reyes-Fermin LM, Nolasco-Amaya EG, Orozco-Ibarra M, Medina-Campos ON, González-Cuahutencos O, et al. ROS scavenging capacity and neuroprotective effect of α -mangostin against 3-nitropropionic acid in cerebellar granule neurons. *Exp Toxicol Pathol* 2009; 61: 491-501.
 49. Muchtaridi M, Wijaya CA. Anticancer potential of α -mangostin. *Asian J Pharm Clin Res* 2017; 10: 440.
 50. Rizaldy D, Hartati R, Nadhifa T, Fidrianny I. Chemical compounds and pharmacological activities of mangosteen (*Garcinia mangostana* L.)—Updated Review. *Biointerface Res Appl Chem* 2021; 12: 2503-2516.
 51. Rahbardar MG, Eisvand F, Rameshrad M, Razavi BM, Hosseinzadeh H. *In vivo* and *in vitro* protective effects of rosmarinic acid against doxorubicin-induced cardiotoxicity. *Nutr Cancer* 2021; 74: 747-760.
 52. Tangpong J, Miriyala S, Noel T, Sinthupibulyakit C, Jungsuwadee P, Clair DS. Doxorubicin-induced central nervous system toxicity and protection by xanthone derivative of *Garcinia mangostana*. *Neuroscience* 2011; 175: 292-299.
 53. Tangpong J, Miriyala S, Noel T, Sinthupibulyakit C, Jungsuwadee P, St Clair DK. Doxorubicin-induced central nervous system toxicity and protection by xanthone derivative of *Garcinia mangostana*. *Neuroscience* 2011; 175: 292-299.
 54. Blum RH, Carter SK, Agre K. A clinical review of bleomycin—a new antineoplastic agent. *Cancer* 1973; 31: 903-914.
 55. Li R-s, Xu G-h, Cao J, Liu B, Xie H-f, Ishii Y, et al. Alpha-mangostin ameliorates bleomycin-induced pulmonary fibrosis in mice partly through activating adenosine 5'-monophosphate-activated protein kinase. *Front Pharmacol* 2019; 10: 1305.
 56. Galluzzi L, Senovilla L, Vitale I, Michels J, Martins I, Kepp O, et al. Molecular mechanisms of cisplatin resistance. *Oncogene* 2012; 31: 1869-1883.
 57. Yao X, Panichpisal K, Kurtzman N, Nugent K. Cisplatin nephrotoxicity: A review. *Am J Med Sci* 2007; 334: 115-124.
 58. Sánchez-Pérez Y, Morales-Bárcenas R, García-Cuellar CM, López-Marure R, Calderon-Oliver M, Pedraza-Chaverri J, et al. The alpha-mangostin prevention on cisplatin-induced apoptotic death in LLC-PK1 cells is associated to an inhibition of ROS production and p53 induction. *Chem Biol Interact* 2010; 188: 144-150.
 59. Aisha AF, Abu-Salah KM, Ismail Z, Majid AMSA. α -Mangostin enhances betulinic acid cytotoxicity and inhibits cisplatin cytotoxicity on HCT 116 colorectal carcinoma cells. *Molecules* 2012; 17: 2939-2954.
 60. Li Q, Yan X-t, Zhao L-c, Ren S, He Y-f, Liu W-c, et al. α -Mangostin, a dietary xanthone, exerts protective effects on cisplatin-induced renal injury via PI3K/Akt and JNK signaling pathways in HEK293 cells. *ACS omega* 2020; 5: 19960-19967.
 61. Reyes-Fermin LM, Avila-Rojas SH, Aparicio-Trejo OE, Tapia E, Rivero I, Pedraza-Chaverri J. The protective effect of alpha-mangostin against cisplatin-induced cell death in LLC-PK1 cells is associated to mitochondrial function preservation. *Anti-oxidants* 2019; 8: 133.
 62. Wang-Fischer Y, Garyantes T. Improving the reliability and utility of streptozotocin-induced rat diabetic model. *J Diabetes Res* 2018; 2018: 8054073.
 63. Furman BL. Streptozotocin-induced diabetic models in mice and rats. *Curr Protoc Pharmacol* 2015; 70: 1-5.47. 20.
 64. Karim N, Jeenduang N, Tangpong J. Anti-glycemic and anti-hepatotoxic effects of mangosteen vinegar rind from *Garcinia mangostana* against HFD/STZ-induced type II diabetes in mice. *Polish J Food Nutr Sci* 2018; 68: 163-169.
 65. Karim N, Rahman A, Chanudom L, Thongsom M, Tangpong J. Mangosteen vinegar rind from *Garcinia mangostana* prevents high-fat diet and streptozotocin-induced type II diabetes nephropathy and apoptosis. *J Food Sci* 2019; 84: 1208-1215.
 66. Husen SA, Ansori M, Nur A, Hayaza S, Susilo K, Joko R, et al. Renal protective effects of gamma-mangostin in streptozotocin-induced diabetic mice. *Indian J Forensic Med Toxicol* 2020; 14: 1251-1256.
 67. Timmins GS, Deretic V. Mechanisms of action of isoniazid. *Mol Microbiol* 2006; 62:1220-1227.
 68. Metushi I, Cai P, Zhu X, Nakagawa T, Uetrecht J. A fresh look at the mechanism of isoniazid-induced hepatotoxicity. *Clin Pharmacol Ther* 2011; 89: 911-914.
 69. Pramana TY, Wasita B, Widyaningsih V, Cilmiaty R, Suroto S, Mudigdo A, et al. The ethanol extract of *Garcinia mangostana* L peel reduces the isoniazid-induced liver damage in rats. *Bali Med J* 2021; 10: 156-159.
 70. Agrawal S, Khazaeni B. Acetaminophen toxicity. *Statpearls [internet]* 2021.
 71. Ramachandran A, Jaeschke H, editors. Acetaminophen hepatotoxicity. *Seminars in liver disease*; 2019: Thieme Medical Publishers.
 72. Yan X-t, Sun Y-s, Ren S, Zhao L-c, Liu W-c, Chen C, et al. Dietary α -mangostin provides protective effects against acetaminophen-induced hepatotoxicity in mice via Akt/mTOR-mediated inhibition of autophagy and apoptosis. *Int J Mol Sci* 2018; 19: 1335.
 73. Ibrahim SR, El-Agamy DS, Abdallah HM, Ahmed N, Elkablawy MA, Mohamed GA. Protective activity of totophyllin A, a xanthone isolated from *Garcinia mangostana* pericarps, against acetaminophen-induced liver damage: role of Nrf2 activation. *Food Funct* 2018; 9: 3291-3300.
 74. Allawadhi P, Khurana A, Sayed N, Kumari P, Godugu C. Isoproterenol-induced cardiac ischemia and fibrosis: Plant-based approaches for intervention. *Phytother Res* 2018; 32: 1908-1932.
 75. Boarescu P-M, Chirilă I, Bulboacă AE, Bocşan IC, Pop RM, Gheban D, et al. Effects of curcumin nanoparticles in isoproterenol-induced myocardial infarction. *Oxid Med Cell Longev* 2019; 2019: 7847142.
 76. Devi SP, Kannan V. Ameliorative prospective of alpha-mangostin, a xanthone derivative from *Garcinia mangostana* against beta adrenergic catecholamine induced myocardial toxicity and anomalous cardiac TNF-alpha and COX-2 expressions in rat. *Exp Toxicol Pathol* 2008; 60: 357-364.
 77. Devi Sampath P, Vijayaraghavan K. Cardioprotective effect of α -mangostin, a xanthone derivative from mangosteen on tissue defense system against isoproterenol-induced myocardial infarction in rats. *J Biochem Mol Toxicol* 2007; 21: 336-339.
 78. Ismail Z, Morcos MA, EL-Shafei MDE-D, Helmy FA-ZM. Histological and immunohistochemical study on the possible effect of mangosteen and mesenchymal stem cells on isoproterenol induced myocardial infarction in adult male Albino rats. *Egypt J Histol* 2019; 42: 651-666.
 79. El-Marasy SA, Abd-Elsalam RM, Ahmed-Farid OA. Ameliorative effect of silymarin on scopolamine-induced dementia in rats. *Open Access Maced J Med Sci* 2018; 6: 1215-1224.
 80. Lee Y, Kim S, Oh Y, Kim Y-M, Chin Y-W, Cho J. Inhibition of oxidative neurotoxicity and scopolamine-induced memory impairment by γ -mangostin: *In vitro* and *in vivo* evidence. *Oxid Med Cell Longev* 2019; 2019: 3640753.
 81. Sattayasai J, Chaonapan P, Arkaravichie T, Soi-Ampornkul R, Junnu S, Charoensilp P, et al. Protective effects of mangosteen extract on H₂O₂-induced cytotoxicity in SK-N-SH cells and scopolamine-induced memory impairment in mice. *PLoS one* 2013; 8: e85053.
 82. Staňková P, Kučera O, Lotková H, Roušar T, Endlicher R, Červinková Z. The toxic effect of thioacetamide on rat liver *in vitro*. *Toxicol In Vitro* 2010; 24: 2097-2103.
 83. Nilbunga S. Investigation of therapeutic effects of α -mangostin on thioacetamide-induced cirrhosis in rats. *J Med Assoc Thai* 2015; 98: S91-S97.
 84. Rodniem S, Tiyao V, Nilbu-Nga C, Poonkhum R, Pongmayteegul S, Pradidarcheep W. Protective effect of alpha-

- mangostin on thioacetamide-induced liver fibrosis in rats as revealed by morpho-functional analysis. *Histol Histopathol* 2018; 34: 419-430.
85. Shukla V, Shukla P, Tiwari A. Lead poisoning. *India J Med specialities* 2018; 9: 146-149.
86. Shabani M, Hadeiy SK, Parhizgar P, Zamani N, Mehrad H, Hassanian-Moghaddam H, et al. Lead poisoning; a neglected potential diagnosis in abdominal pain. *BMC Gastroenterol* 2020; 20: 134.
87. Phyu MP, Tangpong J. Neuroprotective effects of xanthone derivative of *Garcinia mangostana* against lead-induced acetylcholinesterase dysfunction and cognitive impairment. *Food Chem Toxicol* 2014; 70: 151-156.
88. Rana MN, Tangpong J, Rahman MA. Xanthenes protects lead-induced chronic kidney disease (CKD) via activating Nrf-2 and modulating NF- κ B, MAPK pathway. *Biochem Biophys Rep* 2020; 21: 100718.
89. Rahmayanti F, Sastradipura DFS, Mas'Ud Z, Bachtiar B, Wimardhani YS, Permana G. Acute oral toxicity testing of ethyl acetate fraction from *Garcinia mangostana* Linn extract in sprague-dawley rats. *Res J Med Plant* 2016; 10: 261-264.
90. Sunarjo L, Suharti O, Susanto H. The preliminary study on safety of using mangosteen peel extract as natural herbs. *J med sci clin res* 2017; 50: 24851-24856.
91. Choi YH, Han SY, Kim Y-J, Kim Y-M, Chin Y-W. Absorption, tissue distribution, tissue metabolism and safety of α -mangostin in mangosteen extract using mouse models. *Food Chem Toxicol* 2014; 66: 140-146.
92. Hutadilok-Towatana N, Reanmongkol W, Wattanapiromsakul C, Bunkrongcheap R. Acute and subchronic toxicity evaluation of the hydroethanolic extract of mangosteen pericarp. *J Med Plant Res* 2010; 4: 969-974.
93. Catorce MN, Acero G, Pedraza-Chaverri J, Fragoso G, Govezensky T, Gevorkian G. Alpha-mangostin attenuates brain inflammation induced by peripheral lipopolysaccharide administration in C57BL/6J mice. *J Neuroimmunol* 2016; 297: 20-27.
94. Guan H, Li J, Tan X, Luo S, Liu Y, Meng Y, et al. Natural xanthone α -mangostin inhibits LPS-induced microglial inflammatory responses and memory impairment by blocking the TAK1/NF- κ B signaling pathway. *Mol Nutr Food Res* 2020; 64: 2000096.
95. Widowati W, Darsono L, Suherman J, Fauziah N, Maesaroh M, Erawijantari PP. Anti-inflammatory effect of mangosteen (*Garcinia mangostana* L.) peel extract and its compounds in LPS-induced RAW264. 7 cells. *Nat Prod Sci* 2016; 22: 147-153.
96. Tanaka R. Inhibitory effects of xanthone on paraquat-and NaNO₂-induced genotoxicity in cultured cells. *Toxicol Sci* 2007; 32: 571-574.
97. Reyes-Fermin LM, González-Reyes S, Tarco-Álvarez NG, Hernández-Nava M, Orozco-Ibarra M, Pedraza-Chaverri J. Neuroprotective effect of α -mangostin and curcumin against iodoacetate-induced cell death. *Nutr Neurosci* 2012; 15: 34-41.
98. Wang A, Liu Q, Ye Y, Wang Y, Lin L. Identification of hepatoprotective xanthenes from the pericarps of *Garcinia mangostana*, guided with tert-butyl hydroperoxide induced oxidative injury in HL-7702 cells. *Food Funct* 2015; 6: 3013-3021.
99. Wuragil DK. Inhibition inflammation process on cigarette smoke induced rats by extract of mangosteen Peel (*Garcinia mangostana* L) based on oxidant-anti-oxidant profile. *Int J Chemtech Res* 2015; 8: 528-533.
100. Lee D, Choi YO, Kim KH, Chin Y-W, Namgung H, Yamabe N, et al. Protective effect of α -mangostin against iodixanol-induced apoptotic damage in LLC-PK1 cells. *Bioorg Med Chem Lett* 2016; 26: 3806-3809.