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Mesenchymal stem cell therapy in amyotrophic lateral sclerosis (ALS) patients: A comprehensive review of disease information and future perspectives

Shahrzad Najafi ¹, Parizad Najafi ¹, Najmeh Kaffash Farkhad ², Ghazal Hosseini Torshizi ¹, Reza Assaran Darban ¹, Amir Reza Boroumand ³, Sajad Sahab-Negah ^{3, 4}, Mohammad Ali Khodadoust ², Jalil Tavakol-Afshari ^{2*}

¹ Department of Biology, Mashhad Branch, Islamic Azad University, Mashhad, Iran

² Immunology Research Center, Department of Immunology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

³ Neuroscience Research Center, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

⁴ Shefa Neuroscience Research Center, Khatam Alanbia Hospital, Tehran, Iran

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Keywords: Amyotrophic lateral sclerosis Clinical trial Mesenchymal stem cells Motor neurons Neurological disease Amyotrophic lateral sclerosis (ALS) is a rare deadly progressive neurological disease that primarily affects the upper and lower motor neurons with an annual incidence rate of 0.6 to 3.8 per 100,000 people. Weakening and gradual atrophy of the voluntary muscles are the first signs of the disease onset affecting all aspects of patients' lives, including eating, speaking, moving, and even breathing. Only 5-10% of patients have a familial type of the disease and show an autosomal dominant pattern, but the cause of the disease is unknown in the remaining 90% of patients (Sporadic ALS). However, in both types of disease, the patient's survival is 2 to 5 years from the disease onset. Some clinical and molecular biomarkers, magnetic resonance imaging (MRI), blood or urine test, muscle biopsy, and genetic testing are complementary methods for disease diagnosis. Unfortunately, with the exception of Riluzole, the only medically approved drug for the management of this disease, there is still no definitive cure for it. In this regard, the use of mesenchymal stem cells (MSCs) for the treatment or management of the disease has been common in preclinical and clinical studies for many years. MSCs are multipotent cells having immunoregulatory, anti-inflammatory, and differentiation ability that makes them a good candidate for this purpose. This review article aims to discuss multiple aspects of ALS disease and focus on MSCs' role in disease management based on performed clinical trials.

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Introduction

Amyotrophic lateral sclerosis (ALS) is a rare, deadly, idiopathic neurological disease of the human motor system mainly affecting the neurons responsible for controlling voluntary muscle movements (1). The disorder was first described by Jean-Martin Charcot in 1869 as a special neuromotor disorder (2) and is commonly known as Lou Gehrig's disease following the retirement of this famous ballplayer in the 1940s due to this disease (3). Because of the progressive nature of ALS, the symptoms become more severe over time (4). In fact, the disease onset is usually very imperceptible but gradually the symptoms progress into palpable weakness or atrophy (5). Early symptoms include twitching muscle involvement (such as arm, leg, shoulder, or tongue), muscle stiffness (spasticity), muscle spasms, and swallowing problems (6, 7). These symptoms are initially focal, but as the disease progresses, they tend to spread throughout the body (5) which usually leads to swallowing difficulty (dysphagia), speech difficulty (dysarthria), and breathing disorder (dyspnea) (8). In this context, respiratory muscle dysfunction is the main cause of death in ALS patients within 3 to 5 years of the symptom's

onset (5). Unfortunately, despite extensive research on ALS, today there is no clinical or prophylactic treatment for it. However, there is only one approved drug called Riluzole to modify the disease in this area (9, 10) with effectiveness of 3 to 6 months in increasing the patient's mean lifetime. But, this effectiveness varies from person to person (11).

Respecting the above information, finding new treatment strategies can be promising to increase the ALS patient's life span. In this regard, Mesenchymal stem cells (MSCs) have been used for many years in the treatment of ALS patients in preclinical and clinical studies (12, 13). The migratory behavior, tissue regenerative effect (12, 14), and the ability to differentiate into various cell types like neuron cells (15) has made MSCs suitable candidates in this field. This review article tries to study the role of MSCs in the treatment of ALS in addition to familiarity with different aspects of the disease.

Factors involved in ALS development

To date, the cause of ALS is unknown and it remains an unanswered riddle for researchers. However, there is scientific evidence that both genetics and the environment are key players in this scenario (16) as follows:

*Corresponding author: Jalil Tavakol-Afshari. Immunology Research Center, Department of Immunology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran. Email: Tavakolaj@mums.ac.ir

Genetics

Research shows that some particular gene (more than 30 different genes) mutations are associated with motor neuron destruction and ALS development (17). Some of these mutations are directly responsible for the disease induction because they are inherited from parents with infected children and are known as mutated familial ALS genes including SOD1, TARDBP, FUS, OPTN, VCP, UBQLN2, C9ORF72, ANG, SETX, and SQSTM1 (18, 19).

Environmental factors

Many epidemiological studies have examined the environmental factors involved in ALS development (20, 21). Obtained results show that inappropriate lifestyle and various environmental factors may affect the onset and spread of the disease, including exposure to toxins, heavy metals, pesticides, agricultural chemicals, electrical magnetic fields, and viruses (22-24).

Also, physical activity, physical trauma, diet, smoking, occupational hazards, geographic region, cluster (ALS/ Parkinson dementia complex), age, and male gender are considered as other influential environmental factors (25-28). For example, it has been proven that ALS prevalence is higher in men than women (approximately 2 times).

Diagnosis

The ALS diagnosis is mainly based on clinical manifestation showing both upper and lower motor neuron failure and there is no specific single test to diagnose it (29). Also, since the disease can mimic the condition of other neurological diseases, it is difficult and sometimes impossible to diagnose in the early stages (30). Some common tests for

distinguishing from other neurological diseases include Nerve Conduction Study (NCS), Magnetic Resonance Imaging (MRI), blood and urine tests, Electromyogram (EMG), muscle biopsy, and Genetic Testing (30, 31). History of notable patient pain or muscle atrophy, physical examination like spirometry test, neuroimaging laboratory, and electrodiagnostic testing could be very helpful in disease diagnosis (32). In addition, diagnostic criteria have been validated and derived from the El Escorial and modified Airlie House criteria (33). Today, another suggestive complementary method for disease diagnosis is the use of clinical and molecular biomarkers (22).

ALS biomarkers

The lack of a definitive diagnostic method for ALS, and consequently the rapid progression of the disease due to delayed diagnosis, highlights the need to discover new diagnostic solutions. In this regard, there are various clinical and molecular biomarkers with crucial diagnostic roles (34).

Some of the most important of them are electromyography (to detect motor neuron damage), transcranial magnetic stimulation (functional integrity of neurons), electrical impedance myography (functional integrity and muscle structure), and neurophysiological approaches (35, 36). Also, molecular biomarkers like inflammatory cytokines are key helpers in this scenario and can be detected in body fluids including CSF, urine, blood, and saliva (35, 37). These biomarkers can be categorized as follows: biomarkers related to excitotoxicity, oxidative stress, inflammation, metabolic dysfunction, neurodegeneration, and other blood biomarkers (37, 38). Some of the most important molecular biomarkers based on previous studies are summarized in Table 1.

Table 1. Selective molecular biomarkers of amyotrophic lateral sclerosis (ALS)

Biomarkers	Associated process	Identification method	Finding	Ref	
CCR2 Inflammation		chemiluminescent assay & ELISA	Low monocyte expression Low PBMC expression Less CCR2+PBMCs in limb versus bulbar onset	(37, 39)	
IL-4 & IL-6	Inflammation	ELISA	High serum level in hypoxic patients	(40, 41)	
MCP-1	Inflammation	RT-PCR	High plasma level	(37, 42)	
TNF-α	Inflammation	ELISA	High plasma level	(37, 43)	
Caspase-9	Neurodegeneration	ELISA	High serum level Correlated with severity and duration	(37, 44)	
NFL	Neurodegeneration	ELISA	High serum level	(37, 42)	
PNF-H	Neurodegeneration	ELISA	Plasma and serum level correlated with ALSFRS-R decline	(37, 42)	
Cystatin C	Neurodegeneration	Enzyme-linked immunosorbent assay & ELISA	High plasma level	(37, 45, 46	
Nitric Oxide	Excitotoxicity and oxidative stress	Griess nitric colorimetric assay/ELISA	High serum level correlated with duration	(37, 47)	
SOD1	Excitotoxicity and oxidative stress	Enzymatic activity assay/ELISA	Low erythrocyte activity correlated with disease status	(37, 45, 48	
G6PD	Excitotoxicity and oxidative stress	Enzymatic activity assay/ELISA	Correlated with severity	(37, 49)	
Prostaglandin E2	Excitotoxicity and oxidative stress	ELISA	High serum level	(36, 44)	
LDL/HDL ratio	Metabolic dysfunction	Not specified	High plasma level correlated with survival	(37, 50)	
CNTF	Metabolic dysfunction	ELISA	High serum level	(37, 51)	
N-acetyl aspartate	Metabolic dysfunction	MRI & ELISA	High serum level correlated with progression	(37, 51)	
Apolipoprotein E	Metabolic dysfunction	PCR & ELISA	Plasma level correlated with progression and survival	(37, 52)	
MMP-2	Other blood biomarkers	Sandwich ELISA	Correlated with severity	(37, 53)	
MMP-9	Other blood biomarkers	ELISA	High serum level	(37, 53, 54	
TDP-43	Other blood biomarkers	ELISA/ Western Blot & NMR	Cytoplasmic lymphomonocyte location in ALS subtype	(37, 55)	

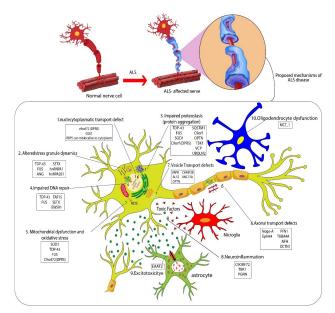
CCR2: C-C chemokine receptor type 2; IL: interleukin; MCP-1: monocyte chemoattractant protein; TNF- α: tumor necrosis factor α; NFL: neurofilament light chain; pNF-H: phosphorylated neurofilament heavy chain; SOD1: Cu/Zn superoxide dismutase/ G6P; Glucose-6-phosphate dehydrogenase; HDL: high-density lipoprotein; LDL: low-density lipoprotein; CNTF: ciliary neurotrophic factor; MMP: matrix metalloproteinase; TDP-43: TAR DNA binding pro

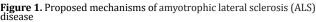
Proposed mechanisms of disease development

Like many other autoimmune diseases, despite global efforts of researchers and clinicians, the causative mechanisms of ALS (particularly in sporadic patients) are still unknown. In fact, several factors and interactions of environment, genetics, age, and other elements are involved in the disease development and progression (56). On the other side, the variety of genetic and phenotypic features between cases has prevented its discovery and general conclusions about the mechanisms of pathogenesis (57). However, some of the most commonly suggested pathogenic mechanisms are: disturbances in RNA metabolism, impaired protein homeostasis, defects in nucleocytoplasmic transport, impaired DNA repair, excitotoxicity, mitochondrial dysfunction, oxidative stress, disturbances in axonal transport, neuroinflammation, oligodendrocyte dysfunction, vesicular transport defects, and alteration in nucleocytoplasmic transport (56, 58) which are schematically given in Figure 1.

Current treatments strategies

The complexity of ALS pathology, both from a molecular mechanisms to clinical symptoms, makes it difficult to identify the exact causative factor, and so develop a single drug or targeted treatment (59). On the other hand, unfortunately, despite the increasing prevalence of the disease following the industrialization of most countries in recent decades (which increases the chance of exposure to environmental risk factors) and the efforts of researchers, there is still no definitive cure for ALS (59). However, in most European societies, Riluzole (Sanofi-Aventis, USA) as an anti-glutamatergic drug is used (50 mg twice daily) as the only approved drug in disease management, but it has adverse effects such as liver problems and diarrhea and in the best case can increase the average life expectancy of patients by 3 to 6 months (9, 60). Also, anti-oxidant drugs





Nucleocytoplasmatic transport defect, altered stress granule dynamics, impaired proteostasis, impaired DNA repair, mitochondrial dysfunction and oxidative stress, axonal transport defects, vesicle transport defects, Neuroinflammation, excitotoxicity, and oligodendrocyte dysfunction are the main proposed mechanisms of ALS onset in patients. (The figure is created using Adobe Illustrator 2020) have been considered by many researchers today due to the crucial role of oxidative stress in ALS induction and/ or progression (56). One of the most common anti-oxidant drugs in this field is Edaravone (Mitsubishi Tanabe Pharma Corporation (MTPC), USA), and its safety and effectiveness have recently been studied in several clinical trials (61-63). In this regard, analysis of the results of 2- years treatment of ALS patients with Edaravone on 621 ALS patients (331 patients in intervention- and 290 patients in control groups) showed that the drug is well tolerated by patients but no significant effect on disease progression and respiratory function was reported (64). However, in another study of 22 ALS patients in Korea, the use of Edaravone showed a modest effect on ALSFRS- score and FVC in patients. Also, only minor side effects were reported in this study (63). Although there are still contradictions in the obtained results in this field, today Edaravone as an approved drug to reduce the progression of ALS is accepted in the USA, Japan, Canada, South Korea, and Switzerland (5).

Other proposed treatment strategies

In addition to the drug treatments mentioned above, some other treatment methods have also been suggested to control and reduce the disease symptoms, including respiratory support (65), psychological and social support (66), occupational therapy (67), speech therapy (68), and physical therapy (69). Also, using trophic factors, respecting their key role in the motor neurons' survival and maintenance, has been proposed as potential therapeutic alternatives in ALS disease. Although, unfortunately, the subcutaneous infusion for some of these trophic factors like CNTF and IGF-1 has not resulted in significant therapeutic benefits in clinical trial studies (70), intrathecally injection (IGF-1 and BDNF) has shown moderate improvement results in some cases, without any severe side effects (49), which indicates the need of conducting more animal and human studies in this field.

Genetically modifications like using silencing RNA of the mutant SOD1 or TARDBP, ALS2, and ALS4 genes, in familial ALS patients could be considered as a good optional therapeutic method (71). Design of iron chelating multifunctional molecules like M30 and HLA20, alone or in combination with other compounds, is another valuable approach to promote the motor nerves' survival via supporting effects on neuro-differentiation and sprouting of axons, leading to reinnervation of muscle fibers (72). Another common method that has been used from the past to the present to treat ALS patients or slow down the disease progression in preclinical and clinical studies is the use of mesenchymal stem cells (which are explained in more detail below) (12, 73).

Mesenchymal Stem cells (MSCs) and their role in ALS improvement

MSCs are adult stromal multipotent cells first isolated from Bone Marrow (BM) as fibroblastic colony-forming units in 1976 by Friedenstein *et al.* and later detected in many other tissues (74). In the human body, they remain uncommitted until receiving a signal to develop into a specialized cell with new specialized cellular functions (75). In many tissues, they serve as an internal reservoir and are also essential for the growth, development, survival, and repair and construction of various body parts (76). MSCs are found in all multicellular organisms and must exhibit three essential properties based on the International Society of Cellular Therapy (ISCT) guideline. First, proliferation by mitotic cell division to produce progeny is the same as the originating cell (76). Second, MSCs should be able to self-renew over long periods (77). Third, they possess the pluripotent ability to differentiate into different multilineages (e.g., osteocytes, adipocytes, and chondrocytes) under certain physiological conditions (14, 78). They are easily accessible and can be isolated from two cell types: adult sources like BM (79), adipose tissue (80), peripheral blood (81), etc., and fetal sources such as amniotic membrane (82), placenta (83), and umbilical cord (84). To date, these cells and their derivatives have been used in the treatment of various diseases and many clinical trials, including diabetes (85), Covid-19 (86-89), and also neurodegenerative diseases like Alzheimer's (90), ALS (12), ataxia (91), and Parkinson's disease (92). These cells have several salient features that make them suitable candidates for the treatment of diseases, including: (1) they do not face ethical considerations associated with the use of Embryonic Stem Cells (ESCs) (93), (2) they can be isolated and expanded both in vitro and in vivo using a simple method (14, 94), (3) they have multiple immunomodulatory and anti-apoptotic properties through various mechanisms (95, 96), (4) Their limited replication time reduces the possibility of malignant transformation after infusion compared with ESCs and iPSCs (97), (5) they are not immunogenic and do not need immune-suppressive drug consumption before injection due to lack of expression of MHCs given the possibility of autologous transplantation (98), (6) and also they have migratory behavior and can differentiate into multiple cell lines like differentiation of BM-MSCs (15) and chorion-MSCs (99) to functional motor neuron-like cells. The golden role of mesenchymal stem cells in neurological diseases like ALS is due to their role of differentiating into neuronal cells and replacing dead and damaged cells with new functional cells. Also, they help to improve the surrounding environment of neurons by secreting trophic factors and removing toxic molecules, and play a protective role for neurons. (12). Repairing damaged nerve sequences such as dendrites and axons and stimulating alternative brain pathways to improve movement and coordination are other effective mechanisms of these cells in the treatment of ALS patients (98). All these brilliant features make MSCs a good source for cell therapy and regenerative medicine. Figure 2 schematically shows some of the effects of mesenchymal stem cells on neuronal restoration in ALS patients.

Pre-clinical studies using MSCs in ALS models

Preclinical research investigating the causes and potential treatments of ALS primarily relies on rat and mouse models, which overexpress mutated human SOD1 genes and exhibit similar patterns of pathology and disease progression to those observed in humans (100). Through the use of these models, researchers have discovered that the transplantation of MSCs via various routes such as intrathecal (IT), intravenous (IV), intramuscular (IM), and intracerebral (IC) can be a safe and effective approach in delaying the decline of motor functions and promoting neurogenesis (101). The secretion of various factors such as cytokines and growth factors like TGF-1 and VEGF is also believed to contribute to the therapeutic protection of neurons

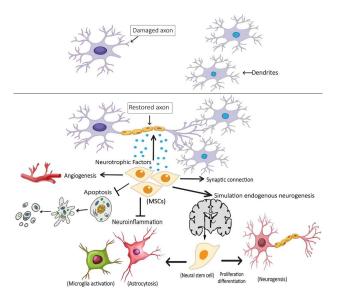


Figure 2. Proposed mechanisms of neurorestoration by mesenchymal stem cells

and the reduction of inflammation after transplantation (102). Furthermore, studies involving the systematic or intra-spinal administration of MSCs from BM or adipose tissue on standard SOD1 mutant SOD1-G93A mouse/rat have shown significant advantages in terms of delaying degeneration of motor neurons, improving motor function, and extending lifespan (73, 103). A study investigated the impact of combined intra-spinal and systemic injection of MSCs in symptomatic SOD-G93A transgenic rats. The outcomes indicated that MSC grafting had a significant effect on motor activity, grip strength, and lifespan, and led to a greater number of motor neurons that were bigger in size with less apoptosis (104). Another investigation revealed that the intravenous injection of MSCs in SOD1 mice had a notable impact on prolonging survival and reducing symptoms, along with the improvement of multiple histological and biochemical parameters (105). These successful experiments have led to the belief that treating ALS with MSCs could improve neuroprotective or neuro-regenerative properties that modulate biological functions (106). To better understand the significance of these experiments, Table 2 provides a summary of the results of preclinical studies conducted in models of ALS with MSCs.

As the results of the above studies show, the use of MSCs in ALS animal models has shown effective results. Delay in motor dysfunction and increase in lifespan are promising results that can be explained by the various mechanisms of action of MSCs, including anti-inflammatory, antiapoptotic, and immunoregulatory features (110).

Clinical studies using MSCs in ALS patients

The first clinical trial that used MSCs to evaluate their safety and potency in the treatment of ALS patients was performed in 2003 by Mazzini *et al.* (111). In this study,

Mesenchymal stem vells (MSCs) secrete various cytokines, and growth factors including neurotrophic factors like transforming growth factor (TGF)-1 and vascular endothelial growth factor (VEGF) contribute to therapeutic protection of neurons, promote endogenous neuronal growth, neurogenesis, and angiogenesis, promote synaptic connection and remyelination of damaged axons, reduce apoptosis, and regulate inflammation mainly through paracrine actions. (Figure is created by Adobe Illustrator 2020)

Rodent model	MSC Source	Dose	Administration	Main Results	Ref	
Irradiated pre-symptomatic male	hBM-MSC	3×10 ⁶ cells	IV (Tail vein)	Increase in survival	(73)	
SOD1G93A mice	nBM-MSC	3×10° cells	IV (Tall Velh)	Delayed in disease onset/progression Delayed the loss of motor function	(75)	
				Increase lifespan of 18-28 days		
0.0 7 - (02)	1.014.040.0			Preservation of neuromuscular junctions and	()	
SOD1 ^{G93A} rat	hBM-MSC	3.6×10 ⁵ cells	IM	corresponding motor neurons	(100)	
				Delay in motor dysfunction		
				Increased survival		
Symptomatic SOD1 ^{G93A} rat	rBM-MSC	2×10 ⁶ cells	IT	Delayed in disease onset/progression	(105)	
				Delayed the loss of motor function		
				Increase lifespan of 8 days		
SOD1 ^{G93A} mice	hBM-MSC	1×10 ⁶ cells	IT	Slowed decline in rotarod test	(106)	
				Increase motor neuro survival		
				Increase lifespan of 17 days		
SOD1 ^{G93A} mice	mBM-MSC	1×10 ⁶ cells	IV	Decreased activated astrocyte and microglial cells	(107)	
				Improvement in profile of oxidative stress/ antioxidant enzyme expression		
	hBM-MSC	5×10 ⁵ cells		Increase lifespan of 14 days		
SOD1 ^{G93A} mice			IT	Delayed in disease onset	(108)	
	110111 11100	Sitte Cells		Reduced astrogliosis	(100)	

hBM-MSCs: Human bone marrow mesenchymal stem cell; rBM-MSC: Rat bone marrow mesenchymal stem cell; mBM-MSC: Mouse bone marrow mesenchymal stem cell; IV: Intravenous; IM: Intramuscular; MSCs: mesenchymal stem cells; ALS: amyotrophic lateral sclerosis

autologous bone marrow MSCs dissolved in the patient's autologous cerebrospinal fluid were injected intrathecally into 7 ALS patients. In terms of safety, these cells were well tolerated in all patients and no serious side effects were observed. Also, MRI images did not show any abnormal structural changes in the spinal cord following cell injection. However, due to the lack of a control group in this study, no results were reported on the effectiveness of the cells, but the good tolerability of cells provided new hope for further research in this area (111). In a 36-month follow-up period, in another study by Mazzini et al. in 2006 another promising result was reported. In this study, out of 9 ALS patients participating, 5 patients showed a significant decrease in forced vital capacity (FVC) and ALS- FRS score following cell injection. Also, no serious side effects were observed in any of the participants (112). Taken together, Mazzini et al.'s results showed that direct injection of autologous expanded MSCs into the spinal cord of ALS patients is protected without any toxicity and well tolerated by patients (112). In 2012, Mazzini et al. reported the results of a 9-year longterm follow-up of 19 ALS patients receiving autologous BM-MSCs. In this study, as expected, no serious side effects

related to cell injection like tumor formation were reported in any of the patients. There was also a significant decrease in disease progression and an increase in life expectancy in 6 patients (113). In another exploratory clinical trial new aspects of the immunoregulatory properties of MSCs appeared. Flow cytometry analysis of peripheral blood monocytes of 5 ALS patients showed significant changes in T lymphocyte subtypes following intrathecal and intravenous injection of MSCs. Obtained results showed a 72% increase in T-regulatory subsets (CD4+ CD25+ T cells) and 30-60% decrease in CD86⁺, CD83⁺, HLADR⁺ myeloid dendritic cells, and CD4+ activated cells 24 hr after MSCs transplantation (114). Following the advancement of sciences, the number of clinical trials associated with ALS has increased in recent years. For example, in a recent study conducted by our research team on 15 ALS patients, promising results were observed. In this study, 3 months after simultaneous IV and IT injections of BM-MSCs, a significant increase in ALS-FRS and FVC was observed. Also, no serious side effects were observed in any of the patients (12). Table 3 summarizes the results of some performed clinical trials in this field.

Table 3. Clinical applications of mesenchymal stem cells in amyotrophic lateral sclerosis patients

Limitation		Study							
	Efficacy	Safety	Observation	Objective	Dose (Injection)	Administration route (Region)	MSC Source	Patients (Age range)	Trial/ Phase
Small sample size Lake of control group Low average age of patients compared to other clinical trials Short observation period before surgery (1 month) Short follow-up period (6 months)	Demonstrates transien efficacy in slowing disease progression Provides evidence of potential clinical benefit	t No serious AEs during the 6-months follow-up period	Stable ALSFRS-R scores and FVC values during the first 3 months follow-up ↓ in ALSFRS-R scores and FVC values 6 months after cell transplantation	and potency of BM-	(2 Simultaneous	Combined IV and IT	Autologous BM-MSC	15 (23-60 years)	Tavakol- Afshari et al. (2021) Phase I/II (12)
Lack of placebo-control group Not evaluating of efficacy	Not evaluated	No serious adverse events were AEs observed during 6 months of follow-up Confirms safety of IT injections of WJ-MSC in patients with ALS			Mean: 0.42×10 ⁶ cells/kg (3 injections in 14 patients, 2 injections in 20 patients, and 1 injection in 9 patients with 2-month intervals in each administration)	IT (Injection region, depended on the clinical symptoms)	Allogenic WJ-MSC	43 (Mean age, 57.3 years)	Barczewska et al. (2019) (101)
Case-control study was weaker than other randomized, double bilnd, controlled trials Short cut-off for follow-up time of ALSFR-R No control for genetic factors that may influence survival even in SALS cases	Demonstrated a positive clinical response and a 70% reduction in risk of death	No new or unexpected reactions were observed after lumbar puncture and administration of MSCs during the 6-month treatment period None of the patients reported symptoms other than ALS symptoms during the 2-month follow-up period Acceptable short-term safety	↓ in disease progression in 21 Patients ↑ in disease progression in 13 patients No significant	of WJ-MSCs on disability development and survival in ALS	30×10° cells/kg (3 injections at 2- month intervals)	IT (L3-L4)	Allogenic WJ-MSC	Treated: 67 Reference: 67 (20-78 years)	Barczewska <i>et a</i> (2020) (13)

Continued Table 3

Lack of postmortem material prohibits any definitive conclusion regarding the fate of the MSCs after injections. Perform CSF analysis only in 2 patients Lack of adequate power to detect a meaningful efficacy Small sample size	IT injection of MSCs may be associated with the positive effect on immune response in ALS patients	h No serious AEs were observed during the 12-	Stable ALSFRS-R scores and FVC during the 6-months post-transplantation 1 in levels of 11-10, TGF-B TGF-β2, TGF-β, an IL-6 compared with the baseline 1 in the levels of MCP-1	Evaluation the safe of 2 repeated IT injections of autologous BM-MS	1×10 ⁶ cells/kg (2 injections at 26-day	- IT (L2-L4)	Autologous BM-MSC	8 (25-75 years)	Oh et al. (2015) Phase I (114)
Not blinding and no sham treatment in the control group Short follow-up time after injection Low mean age of participants compared to other studies Possibly underpowerad to detect a difference in FVC	↓ The ALSFRS-R scores in intervention group at 4 and 6- month follow-up Significant difference i No significant difference i No significant change in FVS and SF-36 during the 4-month h period ↑ in mean levels of TGF-β1-3, IL-6, and 1 -10 between the first and second visits wean levels of TNF α and MCP-1 between the first and second visits No significant difference in survival rate	n S Significantly higher functional stability at 4 and 6 months after MSC L transplantation in intervention group		C Assess the safety and efficacy of 2 repeat IT injections of autologous BM-MS	ed 1×10 ⁶ cells/kg (2 injections at 26-day	. IT (L2-1.4)	Autologous BM-MSC	Treated: 33 Control: 31 (27-75 years)	Oh <i>et al.</i> (2018) Phase II (115)
Small sample size Disease variabilitybetween patients Lack of a control group	Decrease in disease progression in a certair subpopulation of ALS patients		↓ in ALSFR-8 scores during the 6 months post- transplantation period. (from 12.33±1.48 to 0.70±0.949) Points/period in responders and from 0.90±0.39 to 1.60±0.84 points/period In non-responders)	Investigation of safety and efficacy o MSCs in ALS patients	f 15×10 ⁶ cells/kg	IT (Injection region, depended on the clinical symptoms)	Autologous BM-MSC	Responders: 15 Non-responders: 10 Total: 25 (18-65 years)	Siwek <i>et al.</i> (2018) Phase I (116)
Small sample size Short number of injections No logistical support programs		No serious AEs related to BM collection or lumbar puncture reported in any of the patients during each treatment in the 6- month follow-up period No immediate or chronic surgical or local complications observed after MSC injection	↓ in monthly ALSFR-S scores in 2 patients ↑ in monthly ALSFR-S scores in 2 patients No change in disease	Investigate repeated		IT Injection region, depended on the clinical symptoms)	Autologous BM-MSC	8 (41-72 years)	Siwek <i>et al.</i> (2020) Phase II (117)
Further investigation and repeated administration of BM-MSC are required		month follow-up period No new intradural CSF pathology was detected in patients by MRI	A significant reduction/stabilization in ALSFRS-4 decline at 3- months after administration found FVC values remained stable or above 70% for 9- months period in 80% of cases and remained in about 60% of patients at 12-months after application WS values remained stable in 75% of the patients at 3- months after application, and deccreased at 12-months in the	potency of BM-MSC application on the disease progression rate	∑ 15±4.5×10 ⁶ cells/kg	IT (L3-L4)	Autologous BM-MSC	Treated: 26 Control: 23 (18-65 years)	Syková <i>et al.</i> (2017) Phase I/IIa (118)
Small sample size	No notable improvements in or cessation of progression were observed, but 17 patients reported specific mild transient clinical improvements	changes in lumbosacral spine MRI, parameters, and transient lumbosacral radicular pain were observed Appears to be safe at the	aseline to week 1 and week 4 ↑ in CSF monocytes from baseline to week 1 ↓ in CSF monocytes by week 4 ↓ in glucose between baseline	of IT autologous AD-MSC treatment for ALS	$\begin{array}{l} Group 1: 1 \times 10^7\\ cells/kg (1 injection)\\ Group 2: 5 \times 10^7\\ cells/kg (1 injection)\\ Group 3: 5 \times 10^7\\ Group 4: 1 \times 10^6\\ cells/kg (1 injection)\\ Group 4: 1 \times 10^6\\ cells/kg (2 monthly injection)\\ Group 5: 1 \times 10^6\\ cells/kg (2 monthly injections)\\ \end{array}$	IT (L3-L4)	Autologous AD-MSC	27 (36-75 years)	Staff <i>et al.</i> (2016) Phase I/II (119)
Small sample size	Evidence of clinical stabilization or improvement in some patients	No significant AEs were noted during a 25-months follow-up period No significant unexpected pathology findings in MRI of brain and whole spine during the follow-up Safe and effective in some ALS patients.	ALSFRS-R scores were observed after the first 2- months ↑ in CD4* CD25* regulatory T calle	Evaluate the feasibility, safety, and immunologic effects of IT and IV administration of autologous MSCs in ALS patients	63.2×10 ⁶ cells/kg (1 injection intrathecally) 63.2×10 ⁶ cells/kg (1 injection intravenously)	Combined IT [n=10] and IV [n=9] (L3-L4)	Autologous BM-MSC	19 (Mean: 53.0 years)	Karussis et al. (2010) Phase I/II Pilot (113)
Small sample size Short follow-up period. Lack of controls	Short-term follow-up of ALSFRS-R scores suggests a trend towards stabilization of disease	No significant AEs were reported and demonstrated that autologous BM-MSC therapy is safe and feasible in patients of ALS	No significant deterioration in ALSFRS-R composite score from baseline at one-year follow-up	Assess the feasibility efficacy and safety of autologous BM-MSC in ALS patients	f (12-3 or 13-4)	Autologous BM-MSC	10 (23-75 years)	10 (23-75 years)	Prabhakar <i>et al.</i> (2012) (120)

Continued Table 3

Numbers of injected cells were differed from the optimal number Small sample size	No significant changes in disease progression was detected	No serious AEs was	MRI showed no structural changes (including tumor formation) in either the brain or the spinal cord No significant modification of the decline of all clinical and instrumental measures were observed between pre- and post-treatment periods	Assess the safety and efficacy of MSCs transplantation	Mean: 75×10° cells Range: 11.4-120×10° cells	Intraspinal (T4-T5; T5-T6)	Autologous BM-MSC	10 (20-61 years)	Mazzini et al. (2010) (121)
Disease variability between patients Small sample size	The clinical results seem to be promising	No evidence of structural changes of spinal cord or	A significant slowing down of linear decline of the FVC and the ALSFRS was observed after transplantation of MSCs in 5 patients	feasibility and safety of autologous BM-	Mean: 32×10 ⁶ cells Range: 7.0-152×10 ⁶ cells	Intraspinal	Autologous BM-MSC	9 (23-75 years)	Mazzini et al. (2006) (111)
Small sample size	Not evaluated	All patients showed good acceptance of the procedure and no significant changes in psychological status or quality of life were noted	presented major AEs. No bladder and bowel	Verify the safety and tolerability of autologous BM-MSC after directly into the spinal cord of humans	8×10 ⁵ cells/cm ²	Intraspinal	Autologous BM-MSC	9 (23-74 years)	Mazzini et al. (2003) (110)

IV: Intravenous; IT: Intrathecal; BM-MSC: Bone morrow mesenchymal stem cell; WJ-MSC: Wharton's jelly MSCs; ALSFRS-R: Amyotrophic lateral sclerosis functional rating scale-revised; Adverse event: AE; FVC: Forced vital capacity; SALS: Sporadic ALS; AALS: Appel ALS rating scale; SF-36: 36-Item short-form health survey; TGF: Tumor growth factor; II: Interleukin; TNF: Tumor necrosis factor; MCP: Monocyte chemoattractant protein; MRI: Magnetic resonance images

Conclusion

To sum up, ALS is a fatal neuro-degenerative disease affecting all aspects of the sufferer's lifestyle including speaking, swallowing, breathing, moving, and their survival. To date, with the exception of Riluzole for disease management, there is no definitive cure for ALS. In this regard, MSCs are considered a good therapeutic approach due to their brilliant features like anti-inflammatory, immunoregulatory, and differentiation ability. There are many pre-clinical and clinical studies using MSCs in ALS management with promising results. This article aimed to collect general information and available data in this field.

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Authors' Contributions

S SN had the idea for the article. SH N, P N, N KF, GH H, and MA KH performed the literature search and provided the first draft of the manuscript. SH N, P N and GH H made the first draft of art works. J TA, AR B, R AD, N KF and S SN scientifically updated the literature search and critically revised the whole work including art works. All authors read and commented on the final draft of the manuscript.

Conflicts of Interest

The authors declare no conflicts of interest.

References

1. Kiernan MC, Vucic S, Cheah BC, Turner MR, Eisen A, Hardiman O, *et al.* Amyotrophic lateral sclerosis. Lancet 2011;377:942-955.

2. Goetz CG. Amyotrophic lateral sclerosis: Early contributions of Jean-Martin Charcot. Muscle Nerve 2000;23:336-343.

3. Walling A. Amyotrophic lateral sclerosis: Lou Gehrig's disease. Am Fam Physician 1999;59:1489-1496.

4. Irwin D, Lippa CF, Swearer J. Cognition and amyotrophic lateral sclerosis (ALS). Am J Alzheimers Dis Other Demen 2007;22:300-312.

5. Masrori P, Van Damme P. Amyotrophic lateral sclerosis: A clinical review. Eur J Neurol 2020;27:1918-1929.

6. Štětkářová I, Ehler E. Diagnostics of amyotrophic lateral sclerosis: Up to date. Diagnostics 2021;11:231-243.

7. Galvin M, Gaffney R, Corr B, Mays I, Hardiman O. From first symptoms to diagnosis of amyotrophic lateral sclerosis: Perspectives of an Irish informal caregiver cohort-a thematic analysis. BMJ Open 2017;7:e014985-e014992.

8. Richards D, Morren JA, Pioro EP. Time to diagnosis and factors affecting diagnostic delay in amyotrophic lateral sclerosis. In: Amyotrophic Lateral Sclerosis. Brisbane (AU): Exon Publications; 2021 Jul 25. Chapter 2.

9. Hinchcliffe M, Smith A. Riluzole: Real-world evidence supports significant extension of median survival times in patients with amyotrophic lateral sclerosis. Degener Neurol Neuromuscul Dis 2017;7:61-70.

10. Bensimon G, Lacomblez L, Meininger V, Group ARS. A controlled trial of riluzole in amyotrophic lateral sclerosis. New Engl J Med 1994;330:585-591.

11. Group II RS, Lacomblez L, Bensimon G, Meininger V, Leigh P, Guillet P. Dose-ranging study of riluzole in amyotrophic lateral sclerosis. Lancet 1996;347:1425-1431.

12. Tavakol-Afshari J, Boroumand AR, Farkhad NK, Moghadam AA, Sahab-Negah S, Gorji A. Safety and efficacy of bone marrow derived-mesenchymal stem cells transplantation in patients with amyotrophic lateral sclerosis. Regen Ther 2021;18:268-74.

13. Barczewska M, Maksymowicz S, Zdolińska-Malinowska I, Siwek T, Grudniak M. Umbilical cord mesenchymal stem cells in amyotrophic lateral sclerosis: an original study. Stem Cell Rev Rep 2020;16:922-932.

14. Farkhad NK, Mahmoudi A, Mahdipour E. How similar are human mesenchymal stem cells derived from different origins? A review of comparative studies. Curr Stem Cell Res Ther 2021;16:980-993.

15. Abdullah RH, Yaseen NY, Salih SM, Al-Juboory AA, Hassan A, Al-Shammari AM. Induction of mice adult bone marrow mesenchymal stem cells into functional motor neuron-like cells. J Chem Neuroanatom 2016;77:129-142.

16. Figueroa-Romero C, Mikhail KA, Gennings C, Curtin P, Bello GA, Botero TM, *et al.* Early life metal dysregulation in amyotrophic lateral sclerosis. Ann Clin Transl Neurol 2020;7:872-882.

17. Vance C, Rogelj B, Hortobágyi T, De Vos KJ, Nishimura AL, Sreedharan J, *et al.* Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. Science 2009;323:1208-1211.

18. Leblond CS, Kaneb HM, Dion PA, Rouleau GA. Dissection of genetic factors associated with amyotrophic lateral sclerosis. Exp Neurol 2014;262:91-101.

19. Renton AE, Chiò A, Traynor BJ. State of play in amyotrophic lateral sclerosis genetics. Nat Neurosci 2014;17:17-23.

20. Yu Y, Hayashi S, Cai X, Fang C, Shi W, Tsutsui H, *et al.* Pu-erh tea extract induces the degradation of FET family proteins involved

in the pathogenesis of amyotrophic lateral sclerosis. Biomed Res Int 2014;2014:254680-254692.

21. Raymond J, Oskarsson B, Mehta P, Larson T, Horton DK. Clinical Characteristics of a Large Cohort of US Patients Enrolled in the National Amyotrophic Lateral Sclerosis (ALS) Registry, 2010–2015 (P4. 6-001). AAN Enterprises; 2019.

22. Zarei S, Carr K, Reiley L, Diaz K, Guerra O, Altamirano PF, *et al.* A comprehensive review of amyotrophic lateral sclerosis. Surg Neurol Int 2015;6:171-193.

23. Weisskopf M, Morozova N, O'Reilly E, McCullough M, Calle E, Thun M, *et al.* Prospective study of chemical exposures and amyotrophic lateral sclerosis. J Neurol Neurosurg Psychiatr 2009;80:558-561.

24. Zhou H, Chen G, Chen C, Yu Y, Xu Z. Association between extremely low-frequency electromagnetic fields occupations and amyotrophic lateral sclerosis: A meta-analysis. PLoS One 2012;7:e48354-e48360.

25. Morozova N, Weisskopf MG, McCullough ML, Munger KL, Calle EE, Thun MJ, *et al.* Diet and amyotrophic lateral sclerosis. Epidemiology 2008;19:324-337.

26. Beghi E, Logroscino G, Chiò A, Hardiman O, Millul A, Mitchell D, *et al.* Amyotrophic lateral sclerosis, physical exercise, trauma and sports: Results of a population-based pilot case-control study. Amyotroph Lateral Scler 2010;11:289-292.

27. Bozzoni V, Pansarasa O, Diamanti L, Nosari G, Cereda C, Ceroni M. Amyotrophic lateral sclerosis and environmental factors. Funct Neurol 2016;31:7-19.

28. Filippini T, Tesauro M, Fiore M, Malagoli C, Consonni M, Violi F. Environmental and occupational risk factors of amyotrophic lateral sclerosis: A population-based case-control study. Int J Environ Res Public Health 2020;17:6490-6502.

29. Brooks BR, Miller RG, Swash M, Munsat TL. El Escorial revisited: Revised criteria for the diagnosis of amyotrophic lateral sclerosis. Amyotroph Lateral Scler Other Motor Neuron Disord 2000;1:293-299.

30. Campanari M-L, Bourefis A-R, Kabashi E. Diagnostic challenge and neuromuscular junction contribution to ALS pathogenesis. Front Neurology 2019;10:68-75.

 Ferguson TA, Elman LB. Clinical presentation and diagnosis of amyotrophic lateral sclerosis. NeuroRehabilitation 2007;22:409-416.
 Mills KR. Detecting fasciculations in amyotrophic lateral sclerosis: duration of observation required. J Neurol Neurosurg Psychiatr 2011;82:549-551.

33. Luna J, Diagana M, Aissa LA, Tazir M, Pacha LA, Kacem I, *et al.* Clinical features and prognosis of amyotrophic lateral sclerosis in Africa: The TROPALS study. J Neurol Neurosurg Psychiatr 2019;90:20-29.

34. Vejux A, Namsi A, Nury T, Moreau T, Lizard G. Biomarkers of amyotrophic lateral sclerosis: Current status and interest of oxysterols and phytosterols. Front Mol Neurosci 2018;11:12-24.

35. Turner MR, Bowser R, Bruijn L, Dupuis L, Ludolph A, McGrath M, *et al.* Mechanisms, models and biomarkers in amyotrophic lateral sclerosis. Amyotroph Lateral Scler Frontotemporal Degener 2013;14:19-32.

36. Verber NS, Shepheard SR, Sassani M, McDonough HE, Moore SA, Alix JJ, *et al.* Biomarkers in motor neuron disease: A state of the art review. Front Neurol 2019;10:291-318.

37. Robelin L, Gonzalez De Aguilar JL. Blood biomarkers for amyotrophic lateral sclerosis: myth or reality? Biomed Res Int 2014;2014:525097-525107.

38. Chew S, Atassi N. Positron emission tomography molecular imaging biomarkers for amyotrophic lateral sclerosis. Front Neurol 2019;10:135-147.

39. Komiya H, Takeuchi H, Ogawa Y, Hatooka Y, Takahashi K, Katsumoto A, *et al*. CCR2 is localized in microglia and neurons, as well as infiltrating monocytes, in the lumbar spinal cord of ALS mice. Mol Brain 2020;13:1-4.

40. Furukawa T, Matsui N, Fujita K, Nodera H, Shimizu F, Miyamoto K, *et al.* CSF cytokine profile distinguishes multifocal motor neuropathy from progressive muscular atrophy. Neurol

Neuroimmunol Neuroinflamm 2015;2:1-11.

41. Mack CL. Serum cytokines as biomarkers of disease and clues to pathogenesis. Wiley Online Library; 2007. p. 6-8.

42. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, *et al.* Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020;395:497-506.

43. Guidotti G, Scarlata C, Brambilla L, Rossi D. Tumor necrosis factor alpha in amyotrophic lateral sclerosis: Friend or foe? Cells 2021;10:518-536.

44. Iłżecka J. Serum caspase-9 levels are increased in patients with amyotrophic lateral sclerosis. Neurol Sci 2012;33:825-829.

45. Vu LT, Bowser R. Fluid-based biomarkers for amyotrophic lateral sclerosis. Neurotherapeutics 2017;14:119-134.

46. Zhu Y, Yang M, Li F, Li M, Xu Z, Yang F, *et al.* Aberrant Levels of cystatin C in amyotrophic lateral sclerosis: A systematic review and meta analysis. Int J Biol Sci 2018;14:1041-1053.

47. Shimohama MU, Shun. The role of nitric oxide in amyotrophic lateral sclerosis. Amyotroph Lateral Scler Other Motor Neuron Disord 2001;2:71-81.

48. Wright GS, Antonyuk SV, Hasnain SS. The biophysics of superoxide dismutase-1 and amyotrophic lateral sclerosis. Q Rev Biophys 2019;52:1-39.

49. Gouel F, Rolland A-S, Devedjian J-C, Burnouf T, Devos D. Past and future of neurotrophic growth factors therapies in ALS: From single neurotrophic growth factor to stem cells and human platelet lysates. Front Neurol 2019;10:835-845.

50. Laaksovirta H, Soinila S, Hukkanen V, Röyttä M, Soilu Hänninen M. Serum level of CNTF is elevated in patients with amyotrophic lateral sclerosis and correlates with site of disease onset. Eur J Neurol 2008;15:355-359.

51. Simone IL, Ruggieri M, Tortelli R, Ceci E, D'Errico E, Leo A, *et al.* Serum N-acetylaspartate level in amyotrophic lateral sclerosis. Arch Neurol 2011;68:1308-1312.

52. González De Aguilar J-L. Lipid biomarkers for amyotrophic lateral sclerosis. Front Neurol 2019;10:284-289.

53. Sánchez-Guijo F, García-Arranz M, López-Parra M, Monedero P, Mata-Martínez C, Santos A, *et al.* Adipose-derived mesenchymal stromal cells for the treatment of patients with severe SARS-CoV-2 pneumonia requiring mechanical ventilation. A proof of concept study. EClinicalMedicine 2020;25:100454-100462.

54. Łukaszewicz-Zając M, Mroczko B, Słowik A. Matrix metalloproteinases (MMPs) and their tissue inhibitors (TIMPs) in amyotrophic lateral sclerosis (ALS). J Neural Transm 2014;121:1387-1397.

55. Beyer L, Günther R, Koch JC, Klebe S, Hagenacker T, Lingor P, *et al.* TDP 43 as structure based biomarker in amyotrophic lateral sclerosis. Ann Clin Transl Neurol 2021;8:271-277.

56. Mejzini R, Flynn LL, Pitout IL, Fletcher S, Wilton SD, Akkari PA. ALS genetics, mechanisms, and therapeutics: Where are we now? Fronti Neurosci 2019;13:1310-1336.

57. Le Gall L, Anakor E, Connolly O, Vijayakumar UG, Duddy WJ, Duguez S. Molecular and cellular mechanisms affected in ALS. J Pers Med 2020;10:101.

58. Hardiman O, Al-Chalabi A, Chio A, Corr EM, Logroscino G, Robberecht W, *et al.* Amyotrophic lateral sclerosis. Nat Rev Dis primers 2017;3:1-19.

59. Mitsumoto H, Brooks BR, Silani V. Clinical trials in amyotrophic lateral sclerosis: Why so many negative trials and how can trials be improved? Lancet Neurol 2014;13:1127-1138.

60. Lacomblez L, Bensimon G, Leigh PN, Guillet P, Meininger V. Dose-ranging study of riluzole in amyotrophic lateral sclerosis. Amyotrophic Lateral Sclerosis/Riluzole Study Group II. Lancet 1996;347:1425-1431.

61. Okada M, Yamashita S, Ueyama H, Ishizaki M, Maeda Y, Ando Y. Long-term effects of edaravone on survival of patients with amyotrophic lateral sclerosis. eNeurologicalSci 2018;11:11-4.

62. Quarracino C, Bendersky M, Rey R, Rodríguez G. Logistics and safety of edaravone treatment for amyotrophic lateral sclerosis: Experience in Argentina. Acta Neurol Belg 2021;121:1519-1523.

63. Park J-M, Kim S-Y, Park D, Park J-S. Effect of edaravone

therapy in Korean amyotrophic lateral sclerosis (ALS) patients. Neurol Sci 2020;41:119-23.

64. Lunetta C, Moglia C, Lizio A, Caponnetto C, Dubbioso R, Giannini F, *et al.* The Italian multicenter experience with edaravone in amyotrophic lateral sclerosis. J Neurol 2020;267:3258-3267.

65. Hansen-Flaschen J. Respiratory care for patients with amyotrophic lateral sclerosis in the us: In need of support. JAMA Neurol 2021;78:1047-1048.

66. Matuz T, Birbaumer N, Hautzinger M, Kübler A. Psychosocial adjustment to ALS: A longitudinal study. Front Psychol 2015;6:1197-1208.

67. Arbesman M, Lieberman D, Berlanstein DR. Method for the systematic reviews on occupational therapy and neurodegenerative diseases. Am J Occup Ther 2014;68:15-19.

68. Lévêque N. Speech therapy guidelines in patients with amyotrophic lateral sclerosis. Rev Neurol 2006;162:4S269-4S272.

69. Dal Bello-Haas V. Physical therapy for individuals with amyotrophic lateral sclerosis: Current insights. Degener neurol neuromuscul Dis 2018;8:45-54.

70. Henriques A, Pitzer C, Schneider A. Neurotrophic growth factors for the treatment of amyotrophic lateral sclerosis: Where do we stand? Front Neurosci 2010:32-45.

71. Benkler C, Offen D, Melamed E, Kupershmidt L, Amit T, Mandel S, *et al.* Recent advances in amyotrophic lateral sclerosis research: perspectives for personalized clinical application. EPMA J 2010;1:343-361.

72. Golko-Perez S, Mandel S, Amit T, Kupershmidt L, Youdim MB, Weinreb O. Additive neuroprotective effects of the multifunctional iron chelator M30 with enriched diet in a mouse model of amyotrophic lateral sclerosis. Neurotoxic Res 2016;29:208-217.

73. Zhao C, Zhang C, Zhou S, Xie Y, Wang Y, Huang H, *et al.* Human mesenchymal stromal cells ameliorate the phenotype of SOD1-G93A ALS mice. Cytotherapy 2007;9:414-246.

74. Friedenstein AJ, Gorskaja J, Kulagina N. Fibroblast precursors in normal and irradiated mouse hematopoietic organs. Exp Hematol 1976;4:267-274.

75. Bongso A, Lee EH. Stem cells: their definition, classification and sources. Stem Cells: From bench to bedside. 2005;1.

76. Biehl JK, Russell B. Introduction to stem cell therapy. J Cardiovasc Nurs 2009;24:98-107.

77. Sivakumar M, Dineshshankar J, Sunil P, Nirmal RM, Sathiyajeeva J, Saravanan B, *et al.* Stem cells: An insight into the therapeutic aspects from medical and dental perspectives. J Pharm Bioallied Sci 2015;7:S361-371.

78. Zakrzewski W, Dobrzyński M, Szymonowicz M, Rybak Z. Stem cells: Past, present, and future. Stem Cell Res Ther 2019;10:1-22.

79. Aboushady IM, Salem ZA, Sabry D, Mohamed A. Comparative study of the osteogenic potential of mesenchymal stem cells derived from different sources. J clin exp Dent 2018;10:e7-e13.

80. Waldner M, Zhang W, James IB, Allbright K, Havis E, Bliley JM, *et al.* Characteristics and immunomodulating functions of adipose-derived and bone marrow-derived mesenchymal stem cells across defined human leukocyte antigen barriers. Front immunol 2018;9:1642-1654.

81. Ukai R, Honmou O, Harada K, Houkin K, Hamada H, Kocsis JD. Mesenchymal stem cells derived from peripheral blood protects against ischemia. J Neurotrauma 2007;24:508-520.

82. Navas A, Magaña-Guerrero FS, Domínguez-López A, Chávez-García C, Partido G, Graue-Hernández EO, *et al*. Antiinflammatory and anti-fibrotic effects of human amniotic membrane mesenchymal stem cells and their potential in corneal repair. Stem Cells Transl Med 2018;7:906-917.

83. Jiang R, Han Z, Zhuo G, Qu X, Li X, Wang X, *et al.* Transplantation of placenta-derived mesenchymal stem cells in type 2 diabetes: A pilot study. Front Med 2011;5:94-100.

84. Farkhad NK, Reihani H, Moghadam AA, Moghadam AB, Tavakol-Afshari J. Are mesenchymal stem cells able to manage cytokine storm in COVID-19 patients? A review of recent studies. Regen Ther 2021;18:152-160.

85. Liu X, Zheng P, Wang X, Dai G, Cheng H, Zhang Z, et al. A

preliminary evaluation of efficacy and safety of Wharton's jelly mesenchymal stem cell transplantation in patients with type 2 diabetes mellitus. Stem Cell Res Ther 2014;5:1-9.

86. Shetty AK. Mesenchymal stem cell infusion shows promise for combating coronavirus (COVID-19)-induced pneumonia. Aging Dis 2020;11:462-464.

87. Kaffash Farkhad N, Sedaghat A, Reihani H, Adhami Moghadam A, Bagheri Moghadam A, Khadem Ghaebi N, *et al.* Mesenchymal stromal cell therapy for COVID-19-induced ARDS patients: A successful phase 1, control-placebo group, clinical trial. Stem Cell Res Ther 2022;13:1-13.

88. Farkhad NK, Mahmoudi A, Mahdipour E. Regenerative therapy by using mesenchymal stem cells-derived exosomes in COVID-19 treatment. The potential role and underlying mechanisms. Regen Ther 2022;20:61-71.

89. Farkhad NK, Sedaghat A, Reihani H, Moghadam AA, Moghadam AB, Ghaebi NK, *et al.* Specific clinical and immunological changes following mesenchymal stem cell transplantation in COVID-19–induced acute respiratory distress syndrome patients: A phase-I clinical trial. Iran J Allergy Asthma Immunol 2022;21:1-17.

90. Chakari-Khiavi F, Dolati S, Chakari-Khiavi A, Abbaszadeh H, Aghebati-Maleki L, Pourlak T, *et al.* Prospects for the application of mesenchymal stem cells in Alzheimer's disease treatment. Life Sci 2019;231:116564-116564.

91. Nakamura K, Mieda T, Suto N, Matsuura S, Hirai H. Mesenchymal stem cells as a potential therapeutic tool for spinocerebellar ataxia. Cerebellum 2015;14:165-170.

92. Venkataramana NK, Kumar SK, Balaraju S, Radhakrishnan RC, Bansal A, Dixit A, *et al.* Open-labeled study of unilateral autologous bone-marrow-derived mesenchymal stem cell transplantation in Parkinson's disease. Transl Res 2010;155:62-70.

93. Wei X, Yang X, Han Z-p, Qu F-f, Shao L, Shi Y-f. Mesenchymal stem cells: a new trend for cell therapy. Acta Pharmacol Sin 2013;34:747-754.

94. Nardi NB, Meirelles L. Mesenchymal stem cells: Isolation, in vitro expansion and characterization. Handb Exp Pharmacol 2006;174:249-82.

95. Sykova E, Cizkova D, Kubinova S. Mesenchymal stem cells in treatment of spinal cord injury and amyotrophic lateral sclerosis. Front Cell Dev Biol 2021;9:695900-695916.

96. Soleymaninejadian E, Pramanik K, Samadian E. Immunomodulatory properties of mesenchymal stem cells: Cytokines and factors. Am J Reprod Immunol 2012;67:1-8.

97. Kim HJ, Park J-S. Usage of human mesenchymal stem cells in cell-based therapy: Advantages and disadvantages. Dev reprod 2017;21:1-10.

98. Bonafede R, Mariotti R. ALS pathogenesis and therapeutic approaches: the role of mesenchymal stem cells and extracellular vesicles. Front Cell Neurosci 2017;11:80-95.

99. Faghihi F, Mirzaei E, Ai J, Lotfi A, Sayahpour FA, Barough SE, *et al.* Differentiation potential of human chorion-derived mesenchymal stem cells into motor neuron-like cells in two-and three-dimensional culture systems. Mol Neurobiol 2016;53:1862-1872.

100. Lewis CM, Suzuki M. Therapeutic applications of mesenchymal stem cells for amyotrophic lateral sclerosis. Stem Cell Res Ther 2014;5:1-10.

101. Syková E, Rychmach P, Drahorádová I, Konrádová Š, Růžičková K, Voříšek I, *et al.* Transplantation of mesenchymal stromal cells in patients with amyotrophic lateral sclerosis: Results of phase I/IIa clinical trial. Cell Transplant 2017;26:647-58.

102. Wu Y, Chen L, Scott PG, Tredget EE. Mesenchymal stem cells enhance wound healing through differentiation and angiogenesis. Stem Cells 2007;25:2648-2659.

103. Suzuki M, McHugh J, Tork C, Shelley B, Hayes A, Bellantuono I, *et al.* Direct muscle delivery of GDNF with human mesenchymal stem cells improves motor neuron survival and function in a rat model of familial ALS. Mol Ther 2008;16:2002-2010.

104. Forostyak S, Jendelova P, Kapcalova M, Arboleda D, Sykova E. Mesenchymal stromal cells prolong the lifespan in a rat model

of amyotrophic lateral sclerosis. Cytotherapy 2011;13:1036-1046. 105. Uccelli A, Milanese M, Principato MC, Morando S, Bonifacino

T, Vergani L, *et al.* Intravenous mesenchymal stem cells improve survival and motor function in experimental amyotrophic lateral sclerosis. Mol Med 2012;18:794-804.

106. Barczewska M, Grudniak M, Maksymowicz S, Siwek T, Ołdak T, Jezierska-Woźniak K, *et al.* Safety of intrathecal injection of Wharton's jelly-derived mesenchymal stem cells in amyotrophic lateral sclerosis therapy. Neural Regen Res 2019;14:313-318.

107. Boucherie C, Schäfer S, Lavand'homme P, Maloteaux JM, Hermans E. Chimerization of astroglial population in the lumbar spinal cord after mesenchymal stem cell transplantation prolongs survival in a rat model of amyotrophic lateral sclerosis. J Neurosci Res 2009;87:2034-2046.

108. Kim H, Kim HY, Choi MR, Hwang S, Nam K-H, Kim H-C, *et al.* Dose-dependent efficacy of ALS-human mesenchymal stem cells transplantation into cisterna magna in SOD1-G93A ALS mice. Neurosci Lett 2010;468:190-194.

109. Zhou C, Zhang C, Zhao R, Chi S, Ge P, Zhang C. Human marrow stromal cells reduce microglial activation to protect motor neurons in a transgenic mouse model of amyotrophic lateral sclerosis. J Neuroinflamm 2013;10:1-11.

110. Gugliandolo A, Bramanti P, Mazzon E. Mesenchymal stem cells: a potential therapeutic approach for amyotrophic lateral sclerosis? Stem Cells Int 2019;2019:3675627.

111. Mazzini L, Fagioli F, Boccaletti R, Mareschi K, Oliveri G, Olivieri C, *et al.* Stem cell therapy in amyotrophic lateral sclerosis: a methodological approach in humans. Amyotroph Lateral Scler Other Motor Neuron Disord 2003;4:158-161.

112. Mazzini L, Mareschi K, Ferrero I, Vassallo E, Oliveri G, Boccaletti R, *et al.* Autologous mesenchymal stem cells: clinical applications in amyotrophic lateral sclerosis. Neurol Res 2006;28:523-526.

113. Mazzini L, Mareschi K, Ferrero I, Miglioretti M, Stecco A, Servo S, *et al.* Mesenchymal stromal cell transplantation in amyotrophic

lateral sclerosis: A long-term safety study. Cytotherapy 2012;14:56-60.

114. Karussis D, Karageorgiou C, Vaknin-Dembinsky A, Gowda-Kurkalli B, Gomori JM, Kassis I, *et al.* Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis. Arch Neurol 2010;67:1187-1194.

115. Oh K-W, Moon C, Kim HY, Oh S-i, Park J, Lee JH, *et al.* Phase I trial of repeated intrathecal autologous bone marrow-derived mesenchymal stromal cells in amyotrophic lateral sclerosis. Stem Cells Transl Med 2015;4:590-597.

116. Oh KW, Noh MY, Kwon MS, Kim HY, Oh Si, Park J, *et al.* Repeated intrathecal mesenchymal stem cells for amyotrophic lateral sclerosis. Annals Neurol 2018;84:361-373.

117. Siwek T, Barczewska M, Sowa M, Jezierska-Wozniak K, Wojtkiewicz J, Maksymowicz W. Mesenchymal stem cell (MSC) transplantation in patients with amyotrophic lateral sclerosis (ALS): Is there a responder population? J Neurol Neurosci 2018;9:1-6.

118. Siwek T, Jezierska-Woźniak K, Maksymowicz S, Barczewska M, Sowa M, Badowska W, *et al.* Repeat administration of bone marrow-derived mesenchymal stem cells for treatment of amyotrophic lateral sclerosis. Med Sci Monit 2020;26:e927484.

119. Staff NP, Madigan NN, Morris J, Jentoft M, Sorenson EJ, Butler G, *et al.* Safety of intrathecal autologous adipose-derived mesenchymal stromal cells in patients with ALS. Neurology 2016;87:2230-2234.

120. Prabhakar S, Marwaha N, Lal V, Sharma RR, Rajan R, Khandelwal N. Autologous bone marrow-derived stem cells in amyotrophic lateral sclerosis: A pilot study. Neurol India 2012;60:465-469.

121. Mazzini L, Ferrero I, Luparello V, Rustichelli D, Gunetti M, Mareschi K, *et al.* Mesenchymal stem cell transplantation in amyotrophic lateral sclerosis: A phase I clinical trial. Exp Neurol 2010;223:229-237.