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Gliosis induction on locus coeruleus in a living liver donor experimental model: A brief review

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Living Donor Liver Transplantation (LDLT) is a promising approach to treating end-stage liver diseases,

however, some post-operatory complications such as pneumonia, bacteremia, urinary tract infections, and hepatic dysfunction have been reported. In murine models using partial hepatectomy (PHx), a

model that emulates LDLT, it has been determined that the synthesis of hepatic cell proliferation factors

that are associated with noradrenaline synthesis are produced in locus coeruleus (LC). In addition, studies have shown that PHx decreases GABA and 5-HT2A receptors, promotes loss of dendritic

spines, and favors microgliosis in rat hippocampus. The GABA and serotonin-altered circuits suggest

that catecholaminergic neurons such as dopamine and noradrenaline neurons, which are highly

susceptible to cellular stress, can also be damaged. To understand post-transplant affections and to

perform well-controlled studies it is necessary to know the potential causes that explain as a liver surgical procedure can produce brain damage. In this paper, we review several cellular processes

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that could induce gliosis in LC after rat PHx.

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ABSTRACT

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Introduction

Liver transplantation is a treatment for end-stage liver diseases (1, 2), however, most of the organs (or their portions) coe from postmortem donors. The number of patients exceeds the number of livers available for donation. To resolve this problem, the use of liver transplantation from Living Donor Liver Transplantation (LDLT) has gained great relevance (3-5). LDLT consists of the removal and reinsertion of a fraction of the liver (16-30%) from a living, healthy, and compatible donor to a sick patient (6, 7). However, according to the Clavien-Dindo scale, postoperative complications such as keloids, hernias, bilomas, hemorrhages, pneumonia, gram-negative-related bacteremia, urinary tract infections, hepatic dysfunction, impaired regeneration, and severe ischemia/reperfusion injury (IRI) in the graft, result in small-for-size syndrome (SFSS), thrombocytopenia, and others reported in 10-30% of all cases around the world (8-10).

At the preclinical level, the partial hepatectomy (PHx), an animal model designed by Higgins and Anderson in 1931 (11), emulates LDLT. PHx allows cutting at a maximum of 95 percent of liver parenchyma (12, 13). Studies using 30%- or 70%-PHx murine models lead to identifying proliferation, cell cycle regulation, and cell death pathways (14-16). Highlighting that to reach an efficient liver regeneration transcription factor (e.g. Nrf2, Mir33, Keap1), activation of proteins, for example, cleavage caspase-3, epidermal growth factor (EGF), transforming growth factor (TGF), tumor necrosis factor-alpha (TNF-a), humoral response factors such as interleukin 6 (IL-6), and other biomolecules such as insulin, bile acids, and noradrenaline (NA), also named norepinephrine (NE), are tightly involved (13, 17-19).

NA is produced on the locus coeruleus (LC) which leads to the production of growth factors that regulate the proliferation of hepatocytes (20, 21). Brain-liver communication is carried out through the hepatic plexus (Figure 1), Due to this PHx favors neuronal dysfunction due to anterograde nerve retraction (22, 23), which could produce hepatic encephalopathy or neurological abnormalities. PHx induces decreased expression of gamma-aminobutyric acid (GABA) and serotonin (5-HT2A) receptors; furthermore, PHx favors dendritic spine loss and microgliosis/astrocytosis in rat hippocampus (24-26).

Susceptibility of noradrenergic neurons

Stress-based susceptibility of NA neurons could favor degeneration. The clues that can explain this phenomenon are discussed in the next lines.

First, LC is a bilateral nucleus, is the most important noradrenergic source, and is involved in the regulation of learning, pain modulation, and among others, memory consolidation and retrieval. The morphotype-based subsets

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Figure 1. Schematic view of brain-liver axis

The vagus nerve originates from the posterior lateral sulcus of the medulla oblongata, the right and left portions of this nerve make a connection with the hepatic plexus, which is attached to the hepatic artery to integrate into the hepatic parenchyma. Artwork was performed in Biorender.

arranged in the dorsal-ventral region of the LC extend afferents to CA1 from the hippocampus and cerebellum. It has been determined that damage induced by stereotaxic manipulation in the CA1 produces hyperactivity and motor incoordination. Elevated levels of iNOS, IL-1 β , and prostaglandin E2 are identified when cerebellum lose connectivity with LC, which are associated with atypical motor behavior in rats (21, 24, 27-29).

Second, NA is not the only neurotransmitter produced on the LC. One subset of LC neurons co-releases galanin (Gal) in the dorsal and central regions, and another subset co-releases Neuropeptide Y (NPY) in the dorsal portion. In addition, the presence of receptors for the neurotransmitter acetylcholine (nicotinic receptors, nAChRs: α 3, α 6, β 3, and β 4), GABA, orexin/hypocretin, and opioid peptides has also been identified. The fact that the same morphotypesbased subset co-releases two different neurotransmitters can compromise, in pathological conditions, their mitochondrial homeostasis, which could be a disadvantage in preventing oxidative/nitrosative imbalance (27, 30).

Third, LC noradrenergic neurons can be stressed easily due to their low anti-oxidant ability. Epigenetic or morphological changes can be performed. Genomic disturbances include altered expression of genes required for the synthesis and release of NA, as well as genetic transcription errors focused on protein kinases that activate transcription factors or atypical expression of growth factors such as brain-derived neurotrophic factor (BDNF). Regarding morphology, the distress can generate aberrant neuronal arborization and loss of dendritic spines (31-33).

Fourth, NA participates as an inducer of the synthesis of growth factors during liver regeneration. This route requires a complementary source of NA. The alternative source of NA is the chromaffin cells of the adrenal medulla, where NA is synthesized from tyrosine, a process that is regulated by the hypothalamus and cerebral cortex. The loss of connectivity between the brain and the liver can potentiate the activation of systemic inflammation (34-36).

However, joining susceptibility neuronal plus PHx physical effects leads analyze other factors involved in the presence of gliosis in LC.

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Neuronal damage linked to PHx

Morphometric changes in neurons after PHx have been described. Le *et al.* (24) reported a decrease in dendritic spines in neurons of the CA1 region and dentate gyrus from the hippocampus at 1 and 3 days post-PHx. Zhang's group (37) identified pyknotic nuclei, nuclear membrane shrinkage, widened perinuclear space, and decreased synaptic surface area in hippocampal neurons, according to Shilpa's laboratory which reported neuronal apoptosis after ablation of two-70%-PHx (25). To explain this fact it is necessary to revise the hot points of the physical impact of the hepatectomy procedure (Figure 2).

Amino acid metabolism alteration

Studies have shown that the availability of tyrosine modulates the synthesis of NA by phenylalanine hydroxylase enzyme complex. The lack of tyrosine and consequently NA due to PHx could induce distress (38, 39).

Post-operative infection

Lipopolysaccharide (LPS)-related endotoxemia can produce damage to the central nervous system. For example, the infection with *Lactobacillus rhamnosus* alters the GABA receptor expression on the cingulate cortex, hippocampus, amygdala, and LC (39-41).

Redox balance

At the liver level it has been demonstrated that the antioxidant capacity remains stable during liver resection but decreases during the post-surgical period in humans, which can induce post-operative complications such as infections, keloids, or delayed liver regeneration (42-44).

Clinical relevance

Neuronal dysfunction can be given in the first instance by a loss of phenotype, atrophy, and cell death mainly by apoptosis (24-26, 37). The relationship between LC and the development of neurodegenerative diseases has been studied, finding that structural and functional alterations are related to pathologies such as Alzheimer's, Parkinson's,



Figure 2. Potential risk factors to induce gliosis on LC Artwork was performed in Biorender. LC: Locus coeruleus

and Dementia (27). In Alzheimer's Disease, high expression of the Tau protein has been observed in LC (45-46), as well as a neural decrease of 15 to 55%, and abnormal NA content in the LC affecting memory, perception, and visuospatial ability (47); while in Parkinson's disease and dementia there is a neuronal decrease of LC (27) followed by microgliosis and disruption in the biosynthesis of NA (48).

Conclusion

The data support the hypothesis that a transient neuroinflammatory process on LC is activated during liver regeneration, but information regarding the neuroprotective or cytotoxic effect of glial activation on this brain nucleus is of great interest to the researchers.

Authors' Contributions

AA BB, R N and D HB wrote the manuscript. PB PG, AC SG and LM ZF revised the review. All the authors read and approved the final manuscript

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Conflicts of Interest

The authors declare no conflicts of interest.

References

1. Jadlowiec CC, Taner T. Liver transplantation: Current status and challenges. World J Gastroenterol 2016; 22:4438-4445.

 Meirelles Júnior RF, Salvalaggio P, Rezende MBd, Evangelista AS, Guardia BD, Matielo CEL, *et al.* Liver transplantation: history, outcomes and perspectives. Einstein (Sao Paulo) 2015; 13:149-152.
 Emond JC. Live donor liver transplantation: An international perspective. Transplantation 2016; 100:1182-1183.

4. Au KP, Chan ACY. Is living donor liver transplantation justified in high model for end-stage liver disease candidates (35+)? Curr Opin Organ Transplant 2019; 24:637-643.

5. Moghe A, Ganesh S, Humar A, Molinari M, Jonassaint N. Expanding donor selection and recipient indications for living donor liver transplantation. Clin Liver Dis 2021; 25:121-135.

6. Center KOT. Living liver donor. In: Hospital NM, editor. Northwestern Memorial Hospital. Chicago, USA: Northwestern Memorial Hospital; 2018.

7. Li J, Hou Y, Liu J, Liu B, Li L. A better way to do small-for-size liver transplantation in rats. Front Med 2011; 5:106-110.

8. Shaji Mathew J, Manikandan K, Santosh Kumar KY, Binoj ST, Balakrishnan D, Gopalakrishnan U, *et al.* Biliary complications among live donors following live donor liver transplantation. Surgeon 2018; 16:214-219.

9. Liang C, Takahashi K, Furuya K, Oda T, Ohkohchi N. Platelets stimulate liver regeneration in a rat model of partial liver transplantation. Liver Transpl 2021; 27:719-734.

10. Liang C, Takahashi K, Furuya K, Ohkohchi N, Oda T. Dualistic role of platelets in living donor liver transplantation: Are they harmful? World J Gastroenterol 2022; 28:897-908.

11. Michalopoulos GK, Bhushan B. Liver regeneration: Biological and pathological mechanisms and implications. Nat Rev Gastroenterol Hepatol 2021; 18:40-55.

12. Aller MA, Arias N, Prieto I, Agudo S, Gilsanz C, Lorente L, *et al.* A half century (1961-2011) of applying microsurgery to experimental liver research. World J Hepatol 2012; 4:199-208.

 Michalopoulos GK. Hepatostat: Liver regeneration and normal liver tissue maintenance. Hepatology 2017; 65:1384-1392.
 Inderbitzin D, Studer P, Sidler D, Beldi G, Djonov V, Keogh A, *et al.* Regenerative capacity of individual liver lobes in the

microsurgical mouse model. Microsurgery 2006; 26:465-469.
15. Vdoviakova K, Vdoviakova K, Petrovova E, Kresakova L, Maloveska M, Teleky J, *et al.* Importance rat liver morphology and vasculature in surgical research. Med Sci Monit 2016; 22:4716-4728.
16. Hori T, Kirino I, Uemoto S. Right posterior segment graft in living donor liver transplantation. Hepatol Res 2015; 45:1076-1082.

17. Zhou Y, Xu JC, Jia YF, Xu CS. Role of death receptors in the regulation of hepatocyte proliferation and apoptosis during rat liver regeneration. Genet Mol Res 2015; 14:14066-14075.

18. Cirera-Salinas D, Pauta M, Allen RM, Salerno AG, Ramirez CM, Chamorro-Jorganes A, *et al*. Mir-33 regulates cell proliferation and cell cycle progression. Cell Cycle 2012; 11:922-933.

19. Pachowka M, Zegarska J, Ciecierski R, Korczak-Kowalska G. The role of IL-6 during the late phase of liver regeneration. Ann Transplant 2008; 13:15-19.

20. Jensen KJ, Alpini G, Glaser S. Hepatic nervous system and neurobiology of the liver. Compr Physiol 2013; 3:655-665.

21. Llorca-Torralba M, Borges G, Neto F, Mico JA, Berrocoso E. Noradrenergic locus coeruleus pathways in pain modulation. Neuroscience 2016; 338:93-113.

22. Breit S, Kupferberg A, Rogler G, Hasler G. Vagus nerve as modulator of the brain-gut axis in psychiatric and inflammatory disorders. Front Psychiatry 2018; 9:44-58.

23. Hadjihambi A, Arias N, Sheikh M, Jalan R. Hepatic encephalopathy: A critical current review. Hepatol Int 2018; 12:135-147.

24. Le Y, Liu S, Peng M, Tan C, Liao Q, Duan K, *et al.* Aging differentially affects the loss of neuronal dendritic spine, neuroinflammation and memory impairment at rats after surgery. PLoS One 2014; 9:e106837.

25. Shilpa J, Anitha M, Paulose CS. Increased neuronal survival in the brainstem during liver injury: Role of gamma-aminobutyric acid and serotonin chitosan nanoparticles. J Neurosci Res 2013; 91:1203-1214.

26. Yirmiya R, Goshen I. Immune modulation of learning, memory, neural plasticity and neurogenesis. Brain Behav Immun 2011; 25:181-213.

27. Betts MJ, Kirilina E, Otaduy MCG, Ivanov D, Acosta-Cabronero J, Callaghan MF, *et al.* Locus coeruleus imaging as a biomarker for noradrenergic dysfunction in neurodegenerative diseases. Brain 2019; 142:2558-2571.

28. Schwarz LA, Miyamichi K, Gao XJ, Beier KT, Weissbourd B, DeLoach KE, *et al.* Viral-genetic tracing of the input–output organization of a central noradrenaline circuit. Nature 2015; 524:88-92.

29. Abg Abd Wahab DY, Gau CH, Zakaria R, Muthu Karuppan MK, BS AR, Abdullah Z, *et al.* Review on cross talk between neurotransmitters and neuroinflammation in striatum and cerebellum in the mediation of motor behaviour. Biomed Res Int 2019; 2019:1767203.

30. Schwarz LA, Luo L. Organization of the locus coeruleusnorepinephrine system. Curr Biol 2015; 25:R1051-R1056.

31. Borodovitsyna O, Joshi N, Chandler D. Persistent stress-induced neuroplastic changes in the locus coeruleus/ norepinephrine system. Neural Plast 2018; 2018:1892570.

 Hernandez Baltazar D, Nadella R, Cibrian Llanderal T, Puga Olguin A, Barrientos Bonilla AA, Zavala Flores LM, Villanueva Olivo A, *et al.* The causative and curative roles of brain-derived neurotrophic factor in Parkinson's disease. In: Parkinson's Disease and Beyond - A Neurocognitive Approach. InTechOpen 2018; 3:1-21.
 Swinny JD, Valentino RJ. Corticotropin-releasing factor promotes growth of brain norepinephrine neuronal processes through Rho GTPase regulators of the actin cytoskeleton in rat. Eur J Neurosci 2006; 24:2481-2490.

34. Li J, Yan HT, Che JX, Bai SR, Qiu QM, Ren L, *et al.* Effects of neurolytic celiac plexus block on liver regeneration in rats with partial hepatectomy. PLoS One 2013; 8:e73101.

35. Dum RP, Levinthal DJ, Strick PL. Motor, cognitive, and

affective areas of the cerebral cortex influence the adrenal medulla. Proc Natl Acad Sci U S A 2016; 113:9922-9927.

36. Bondarenko NS, Murtazina AR, Nikishina YO, Sapronova AY, Ugrumov MV. Molecular mechanisms of synthesis of noradrenaline as an inducer of development in the adrenal glands of rats in ontogenesis. Dokl Biochem Biophys 2017; 472:23-26.

37. Zhang Y, Lv YL, Si YN, Zhou J, Qian Y, Bao HG. Alpha-lipoic acid attenuates spatial learning and memory impairment induced by hepatectomy. Exp Ther Med 2019; 17:2329-2333.

38. Garabal MV, Arévalo RM, Díaz-Palarea MD, Castro R, Rodríguez M. Tyrosine availability and brain noradrenaline synthesis in the fetus: control by maternal tyrosine ingestion. Brain Res 1988; 457:330-337.

39. Vargas-Caraveo A, Sayd A, Maus SR, Caso JR, Madrigal JLM, Garcia-Bueno B, *et al.* Lipopolysaccharide enters the rat brain by a lipoprotein-mediated transport mechanism in physiological conditions. Sci Rep 2017; 7:13113.

40. Hernandez Baltazar D, Nadella R, Barrientos Bonilla A, Flores Martinez Y, Olguin A, Heman Bozadas P, *et al.* Does lipopolysaccharide-based neuroinflammation induce microglia polarization? Folia Neuropathol 2020; 58:113-122.

41. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, *et al.* Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proc Natl Acad Sci U S A 2011; 108:16050-16055.

42. Schwarz C, Fitschek F, Bar-Or D, Klaus DA, Tudor B, Fleischmann E, *et al.* Inflammatory response and oxidative stress during liver resection. PLoS One 2017; 12:e0185685.

43. Senoner T, Schindler S, Stattner S, Ofner D, Troppmair J, Primavesi F. Associations of oxidative stress and postoperative outcome in liver surgery with an outlook to future potential therapeutic options. Oxid Med Cell Longev 2019; 2019:3950818.

44. Li S, Tan HY, Wang N, Zhang ZJ, Lao L, Wong CW, *et al.* The role of oxidative stress and anti-oxidants in liver diseases. Int J Mol Sci 2015; 16:26087-26124.

45. Liu KY, Marijatta F, Hammerer D, Acosta-Cabronero J, Duzel E, Howard RJ. Magnetic resonance imaging of the human locus coeruleus: A systematic review. Neurosci Biobehav Rev 2017; 83:325-355.

46. Seidel K, Mahlke J, Siswanto S, Kruger R, Heinsen H, Auburger G, *et al*. The brainstem pathologies of Parkinson's disease and dementia with Lewy bodies. Brain Pathol 2015; 25:121-135.

47. Kelly SC, He B, Perez SE, Ginsberg SD, Mufson EJ, Counts SE. Locus coeruleus cellular and molecular pathology during the progression of Alzheimer's disease. Acta Neuropathol Commun 2017; 5:8-21.

48. Tabrez S, Jabir NR, Shakil S, Greig NH, Alam Q, Abuzenadah AM, *et al.* A synopsis on the role of tyrosine hydroxylase in Parkinson's disease. CNS Neurol Disord Drug Targets 2012; 11:395-409.