

Candidiasis in breast cancer: Tumor progression or not?

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ABSTRACT

Candida albicans is an "opportunistic fungal agent" in cancer patients that can become colonized in both mucosal and deep tissues and cause severe infections. Most evidence has shown that *C. albicans* can enhance the progress of different cancers by several mechanisms such as generating virulence factors, participation in endogenous production of pro-inflammatory mediators, and stimulating a wide range of immune cells in the host. The main idea of this review is to describe a range of *Candida*-used mechanisms that are important in candidiasis-associated malignant processes and cancer development, particularly breast cancer. This review intends to provide a detailed discussion on different regulatory mechanisms of *C. albicans* that undoubtedly help to open new therapeutic horizons of cancer therapy in patients with fungal infection. The current therapeutic approach is not fully effective in immunocompromised and cancer patients, and further studies are required to find new products with effective antifungal properties and minimal side effects to increase the susceptibility of opportunistic fungal infections to conventional antifungal agents. So, in this situation, a special therapy should be considered to control the infection and simultaneously have the most therapeutic index on tumor patients.

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Introduction

Breast cancer is the most common cancer and one of the leading causes of death among females worldwide (1). Therapeutic interventions like chemotherapy, targeted therapy, and immunotherapy may lead to alterations of the immune system rendering these patients susceptible to infectious complications. Studies have demonstrated that most women with breast cancer exhibit a high prevalence of opportunistic fungal infection, especially *Candida albicans* (2).

C. albicans is a commensal organism living with humans responsible for opportunistic infections in immunodeficient patients (3). Systemic *C. albicans* infections may negatively affect the outcome of malignancy and produce lengthy hospital stays, intensify the economic burden of disease, and increase morbidity and mortality (4). In patients with a

moderately compromised immune system, *C. albicans* often induce mild or superficial infections, however, this fungus may also establish life-threatening diseases (5).

The role of microorganisms in cancer incidence and morbidity along with their interactions with the immune system and host responses have been explored. However, the impact of fungi on carcinogenesis is not clearly understood mainly because of their much lower prevalence and cumbersome investigation techniques. Interrelation of *C. albicans* to cancer development like colorectal carcinogenesis has been shown (6-8). The primary purpose of this review is to explore the effects of candidiasis on the immune system and tumor cells and highlight some recent findings suggesting that *C. albicans* may have a more extensive role in breast cancer status or progression.

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The host immune response against *Candida albicans*

Immunological crosstalk between the host immune system and *C. albicans* is complex and dynamic since the pathogen employs various strategies to escape antimicrobial immunity (9, 10). Upon initial fungal infection, both innate and adaptive immune reactions restrict fungal proliferation through various approaches.

Innate immune response against *C. albicans*

Candida expresses many pathogen/microbe-associated molecular patterns (PAMPs/MAMPs), including N-linked mannan, β -glucan, α -mannans, acylated lipoprotein, and β -(1-2) oligomannan, which are involved in anti-fungal immunity (11). The interplay between PAMPs and pattern recognition receptors (PRRs) determines the path of inflammation or infection. PRRs activation is accompanied by activation of phagocytosis, transcriptional factors, nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), activator protein 1 (AP-1), interferon regulatory factors (IRFs), CCAAT/enhancer-binding protein beta (C/EBP β), proinflammatory cytokines production, and inflammasome activation which trigger inflammatory pathways (12, 13).

The primary cells that are involved in innate immunity against *Candida* are epithelial and phagocytic cells, including polymorphonuclear neutrophils (PMNs), mononuclear phagocytes, monocytes/macrophages, dendritic cells (DCs), and natural killer cells (NKs) (11). Among them, epithelial cells of the skin, urogenital, gastrointestinal, and respiratory systems are the first line of host defense. The fungus exploits two ways to attack the human host tissues and spread to all other regions of the human body: passive penetration (endocytosis) and active penetration (14). Passive penetration is mediated through the interaction of several adhesions of *C. albicans* including hyphal wall protein1 (HWP1), agglutinin-like sequence 1-9 (ALS1-9), and integrin-like protein 1 (INT1) (15). In contrast, active penetration is directly dependent on fungus features including touch (thigmotropism), hyphal-induced physical pressure, and the secretion of extracellular hydrolases like Saps, class B phospholipase (Plb), and lipase (Lip) families (16, 17).

The interplay between *C. albicans* and epithelial surfaces encourages signaling pathways like NF- κ B and biphasic Mitogen-activated protein kinase (MAPK). It has been shown that activation of NF- κ B, first MAPK phase, ERK1/2, and JNK signaling further promote the expression of antimicrobial peptides like defensins, cathelicidins, and statins (18, 19). The activation of the second MAPK phase is dependent on the hyphal form of *C. albicans* and is associated with inducing c-Fos activity that triggers the secretion of pro-inflammatory molecules such as IL-1 α / β , IL-6, IL-8, Tumor necrosis factor alpha (TNF- α), Granulocyte-macrophage colony-stimulating factor (GM-CSF), and Granulocyte colony-stimulating factor (G-CSF) in vulvovaginal candidiasis (20, 21).

Furthermore, the secretion of some pro-inflammatory mediators such as IL-22 leads to immune cell proliferation, differentiation, and activation (22). The overexpression of IL-22 together with other mediators such as TNF- α and IL-17 stimulates the production of the complement system components C1r, C1s, and anti-fungal peptides by epithelial cells (23, 24). Depending on the type of infection, epithelial cells may promote and generate chemokines to call up neutrophils towards infectious niches to directly kill *Candida*

cells (25). These cells recruit several anti-fungal mechanisms to demolish *Candida* cells including phagocytosis, cytokine secretion, granule enzymes production, antimicrobial peptides, and oxidative burst; latent of which results in the generation of reactive oxygen species (ROS), nitrogen intermediates, and myeloperoxidase (MPO) (26, 27).

The infiltration of neutrophils inhibits *C. albicans* growth and promotes the yeast-to-hyphal transition (9). Urban *et al.* showed that neutrophils are equipped with neutrophil extracellular traps (NETs), which can destroy both yeast and hyphal forms of the fungus, enabling extracellular killing of the microorganism. (28). The bactericidal/permeability-increasing (BPI) protein, lactoferrin, and defensins are among the anti-fungal proteins found in NET granules and are involved in pathogen killing (25).

Monocytes and macrophages are other phagocytic cells directly involved in anti-*Candida* immune responses (29, 30). Macrophages utilize a combination of several oxidative and non-oxidative anti-fungal mechanisms including phagocytosis, antimicrobial peptides, degradative enzymes inducing ROS and nitric oxide synthase (iNOS), and formation of macrophage extracellular traps (METs) to attack the invasive pathogens (31, 32). As professional antigen presenting cells (APCs). DCs also play an essential role in regulating immune responses and bridging the innate to adaptive anti-fungal immune trajectories (33).

DCs possess several PRRs enabling them to localize infection and activate naïve T cells (34). Furthermore, cytokines secreted by DCs lead T cells to differentiate into Th1, Th2, Th17, and Tregs. In response to the yeast form of the pathogen, DCs are activated and secrete IL-12 which promotes Th1 cells, while ingestion of hypha form triggers IL-4 production and Th2 differentiation (35). Interestingly, Dectin-1 interaction with the yeast form induces IL-17 and IL-6 overexpression promoting Th17 cell responses that protect cutaneous infection. Filamentous form, in contrast, provokes just Th1 differentiation, which is essential for the control of systemic infection (36).

Additionally, the cell wall protein fraction (CPF) of *C. albicans* initiates MHC-II, CD86, and CD40 expression on dendritic cells, which indicates DCs maturation (37). Van de Veerdonk studied the innate immune mechanisms involved in triggering Th17 responses and showed that *C. albicans* mannan, macrophage mannose receptor (MR), and TLR2/dectin-1 pathway activate and induce IL-17 production as a pathogen-specific defense (38). Moreover, it is shown that vaccination with recombinant cell surface glycoprotein Als3p, a significant component of the hyphal form, induces Th1, Th17, and Th1/17 lymphocytes, resulting in decreased tissue infectious burden (39).

NK cells are innate cytotoxic lymphocytes that can directly and potentially recognize and phagocytize *C. albicans* cells by releasing molecular contents such as perforin and granzymes, causing receptor-mediated apoptosis (40, 41). They usually have effective roles in anti-viral and anti-tumor immunity as well (42). These cells modulate various innate and adaptive immune cells and responses through secretion of various pro-inflammatory mediators (43). A study showed that NK cells in defense against *C. albicans* infection exhibit different roles depending on the state of host defense and immunological context. This study demonstrates that NK cells cause hyper/chronic inflammation in candidiasis and immunocompetent hosts by stimulating excessive pro-inflammatory mediators, which may be redundant and even

detrimental to host defense and results in the exacerbation of infection (44).

Adaptive immune response against *C. albicans*

Adaptive immune responses are mainly carried out by two cell types: T lymphocytes and B lymphocytes. T lymphocytes are divided into CD8+ cytotoxic cells and CD4+ helper (Th) cells, both of which participate in anti-fungal immunity and their activation is monitored by DC subsets that migrate to the local lymph nodes. Activation of different subtypes of DCs via distinct signaling pathways can shape diverse T-cell responses against *Candida* infections (45, 46). The importance of CD4+ helper T cells is well recognized in host defense against *Candida* infections (47). Th1, Th2, Th17, and Tregs, all play pivotal roles in *Candida*-specific immune reactions.

Th2 and Th17 mediated reactions are initiated by collaboration of myeloid (inflammatory) dendritic cells via TLR-MyD88 pathways, whereas Th1 and T regulatory (Treg) cell responses originate in plasmacytoid (tolerogenic) dendritic cell interaction via TRIF signaling pathways (48). Several inflammatory cytokines including IFN- γ , IL-12, IL-1 β , IL-2, IL-6, TNF- α , IL-17, IL-21, IL-22, and IL-23 are bound to Th1 and Th17 cell mediated immunity (46). In a 2007 research, it was demonstrated that IFN- γ knockout mice show significantly lower surveillance while surveillance rate was increased in IL-4, a Th2 cytokine, knockout mice (49). This finding indicates a protective role of Th1 and detrimental effect of Th2 response, which confirms the importance of Th1/Th2 balance against *Candida* infections (48, 50).

Notably, another study represented that as with epithelial cell responses, various Th phenotypes-specific responses against *Candida* infections are tissue-specific (51). Recognition of *Candida* by PRRs on DCs such as dectin-1 and dectin-2 promotes Th17 proliferation and development (52). On the other hand, Th17 and its related cytokines, IL-17 and IL-23, play a crucial role in protective immunity and reduce fungal burdens in vaginal candidiasis (53). Additionally, Th1- and Th17-associated cytokines, IFN- γ and IL-17, can prohibit cancer development, and as a result, suppressing these cytokines may increase the risk of cancer in the candidiasis tissue environment (54).

Considering the importance of neutrophils for anti-fungal immunity, IL-17R deficiency could indirectly affect

these cells by impairing NK cell functions. Simultaneously, GM-CSF not only induces neutrophil migration and *Candida* killing but also stimulates oxidative metabolism (55-57). In addition, defects in GM-CSF signaling increase the risk of invasive aspergillosis infection, and aligned with this finding, GM-CSF treatment could exacerbate the fungicidal activity of neutrophils and monocytes and improve fungal clearance in the lung (58).

Treg cells are a subpopulation of CD4+ helper T cells, which are involved in controlling inflammation, autoimmunity, and hemostasis (59, 60). Tregs can potentially exhibit opposite features during infections and breast cancer. For instance, they may facilitate tumor growth and metastasis by suppressing most immune cells including CD4+ and CD8+ T cells, B cells, NK cells, and APCs; or enhance microbial clearance and cancer improvement through stimulation of immune mechanisms (61). Despite the immune-suppressing function of Tregs, their role in combatting *Candida* is controversial. In a report of oral *Candida* infection in mice, Treg cells enhanced Th17 differentiation by inducing IL-17 secretion and IL-2 consumption, which helped fungal clearance (59). However, in another study on mouse models, Treg cell depletion or IL-10 administration did not result in Th17 cell response to *Candida* in the murine oral epithelium infection (62). Nonetheless, the intravenous injection of infection-induced Foxp3+ Treg cells in C57BL/6 mice was associated with fungal kidney infection (63). In line with previous findings, this study indicated the role of Tregs in Th17 responses but showed suppressive effects of these regulatory cells in the down-regulation of Th1 and Th2 pathways (59, 62). Also, our previous finding showed that *C. albicans* could induce Treg homing in the tumor microenvironment, which led to increased tumor growth (64).

Collectively, innate and adaptive immune responses consist of a wide range of molecules and receptors that communicate with each other in a coordinated manner to provide host protection against pathogens. However, our knowledge of immune responses against fungal pathogens is inadequate and incomplete, and possibly further research would help us better understand how immune signals interact and how invading fungi hide themselves from immune cells and escape from the immune system to develop cancer-related processes (Figure 1 and 2).

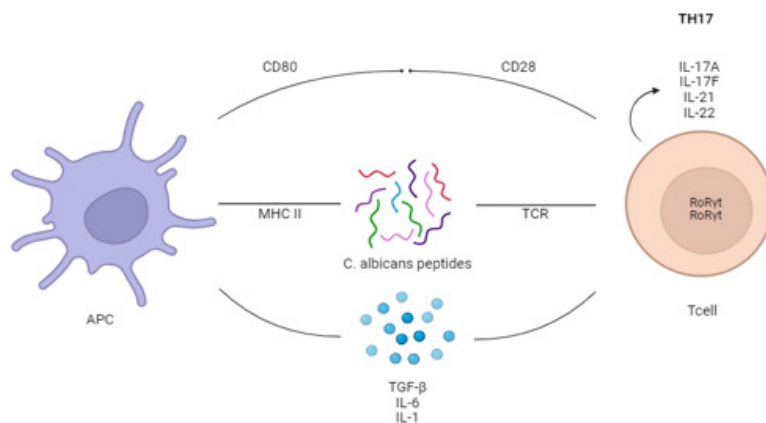


Figure 1. The effect of *Candida albicans* products on the modulation of T cells and induction of suppressor cytokines

CD28: Cluster of Differentiation 28; MHC-II: Major histocompatibility complex-II; TCR: T-cell receptor; TGF- β : Transforming growth factor beta; APC: Antigen-presenting cell; ROR- γ t: Retinoic acid receptor- γ t

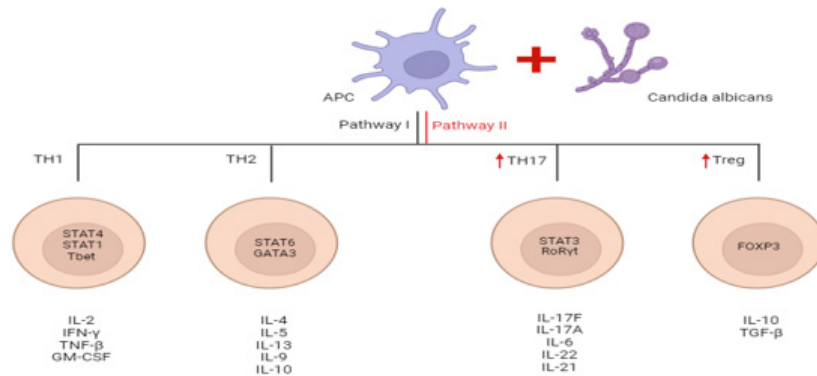


Figure 2. Interaction of *Candida albicans* and antigen presenting cells can modulate T cells response and cytokines pattern

TH: T helper; Treg: Regulatory T cell; STAT: Signal transducer and activator of transcription proteins; Tbet: T-box transcription factor; IFN- γ : Interferon gamma- γ ; TNF- α : Tumor necrosis factor- α ; GM-CSF: Granulocyte-macrophage colony-stimulating factor; GATA-3: GATA binding protein 3; FOXP3: Forkhead box protein 3

Role of *C. albicans* in tumorigenesis

A large body of evidence exists that fungi participate in the processes that encourage carcinogenesis and cancer progression (65, 66). There are several pathways through which, *Candida* spp. increase the risk of cancer and metastasis (67), namely the generation of carcinogenic products (nitrosamine, acetaldehyde), inducing inflammation (inflammatory mediators, cytokines, and chemokines), molecular mimicry, and epigenetic modifications.

Generation of carcinogenic by-products

Nitrosamine

N-nitroso compounds or nitrosamines are chemical carcinogens of nitrogen oxides (e.g., nitrites and nitrates). Nitrosamines react with DNA and form adducts with phosphate residues that may stimulate the activation of specific proto-oncogenes (68, 69). There is much evidence linking nitrosamines to the incidence of various types of cancer, including colorectal, stomach, esophagus, nasopharynx, bladder, and breast (70-73). Studies have shown that 4-(methylnitrosamino)-1-(3-pyridyl) 1-butanone (NNK), the most potent carcinogen among tobacco-related nitrosamines, can affect various neoplastic processes and increase the risk of developing breast cancer by stimulation of estrogen (73).

Moreover, a study reported that some yeasts like *C. albicans* might have a role in oral cancers through endogenous nitrosamine breakdown (74). Invasive esophageal *Candida* infection is also associated with neoplastic alterations in the oral epithelium and esophageal squamous cell carcinoma (OSCCs) (75-77). Furthermore, among different grades of leukoplakia, *C. albicans* growth was observed only in severe dysplastic patients, while there was no growth in mild or moderate cases. The evidence demonstrated that *C. albicans* production of nitrosamines might indirectly promote cancer progression (78). In addition, the expression of wild-type BRCA1 suppresses the growth of breast and ovarian epithelial tumor cell lines (79). The study by Humphrey *et al.* showed that BRCA1 inhibited yeast growth by several mechanisms, and mutations in this gene may promote breast cancer progression and yeast dissemination (80).

Furthermore, previous reports showed that the level of active-matrix metalloproteinases (MMPs), especially MMP-2, could be considered a breast cancer metastasis indicator. MMPs are secreted as pro-enzymes, activated by proteolytic cleavage, and regulated by a family of inhibitors (i.e., tissue inhibitors of matrix metalloproteinases; TIMPs). Previous

studies showed that TIMP-1 expression is enhanced in breast cancer and possibly other types of cancer. In this regard, Taheri *et al.* showed that *C. albicans* stimulates MMP-9 secretion in mice-bearing tumors and seems to utilize MMP-9 to degrade the tissue and disseminate. Also, they demonstrated that TIMP-1 was increased in the presence of infection and tumor. Overall, the results showed that candidiasis positively affected tumor progression and metastasis. (4, 81). It can be concluded that *Candida*, by alteration of tumor suppressor genes and proto-oncogenes, provides a context for tumor initiation and progression.

Alcohol-derived carcinogenic agents

Alongside nitrosamines, *Candida* takes advantage of another pathway to promote carcinogenesis, especially for oral carcinoma. *C. albicans* utilizes the enzyme alcohol dehydrogenase (ADH1) to oxidize ethanol of alcoholic beverages and some substances such as carbohydrates, producing elevated levels of acetaldehyde (ACH) (>100 μ M), which is highly toxic, mutagenic, and carcinogenic; thus indisputably elevates the risk of carcinoma (82). Binding to proteins and DNA, acetaldehyde creates abnormal chromosomal aberrations, alters molecules' typical structure and function, and induces inflammation by producing inflammatory mediators such as NF- κ B in the trachea (83). Moreover, acetaldehyde triggers mitochondrial damage and boosts ROS (84). These established alterations cause genome instability, suppression of the apoptotic process, proto-oncogene activation, and cell cycle disturbances, which may favor tumor progression (85).

C. albicans could also induce oral epithelial dysplasia (86). In an *in vitro* study, L-2-hydroxyisocaproic acid (HICA), a novel antifungal agent, completely inhibited ACH production, reducing the mutagenic potential of *C. albicans* biofilms (87). In addition, a study represented that intracellular ethanol metabolism to acetaldehyde causes DNA damage, causing Fanconi anemia-breast cancer (FA-BRCA) susceptibility in alcohol-associated breast and liver cancers (88). Overall, it seems that *Candida*, via production of alcohol-derived carcinogenic agents, can stimulate breast cancer.

Heme oxygenase (HO) enzymes

Hemoglobin (Hb) is considered an iron-containing oxygen-carrier metalloprotein (89). Heme oxygenase enzymes are found in bacteria, fungi, and mammals. *C. albicans* heme oxygenase gene (CaHMX1) exploits extracellular heme or Hb as a significant source of iron to

degrade the ferroheme to free iron, biliverdin, and carbon monoxide (CO) and allow the fungus to feed on the iron to support microbial growth and pathogenesis (90, 91). Moreover, when *Candida* faces iron deprivation, hemin induces the Hmx1 gene, sustaining fungal survival and virulence (91).

In addition to hemolytic capacity, Rbt5 and Pga7, two extracellular membrane proteins, help the yeast to transfer iron from heme and hemoglobin (92, 93). Besides these two proteins, in the hyphal form, another member of the Rbt5 protein family, Csa2, is also involved in iron absorption (94). In 1966, it was shown that endogenous CO production is correlated with blood heme destruction (95). This study showed that CO, despite its poisonous effects, could be considered an immune modulator at therapeutic doses. Up-regulating Hmx1, *Candida* can alter the immune system. Like human heme oxygenase, *Candida* Hmx1 could produce CO, which leads the immune system toward Th2 expression by diminishing cellular Th1 immunity and weakening antigen presentation. It must be considered that Th2 cells facilitate tumor progression. Evidence shows that Th2 predominance enables tumor growth in pancreatic cancer, and a higher Th2/Th1 lymphocyte ratio could affect the survival rate after surgery (95).

Our previous findings demonstrated that *C. albicans* infection was followed by decreased IFN- γ /IL-4 ratio and increased IL-10 and TGF- β , which results in augmented tumor growth (95). Although it is expected that Hmx1, by shifting immune responses to Th2, might facilitate tumor growth, evidence showed the beneficial role of HO-1 in blocking breast tumor invasion (96, 97). Other investigations also represented HO-1 inhibiting effect on the TPA-induced MMP-9 expression and invasiveness with activation of PKC/ROS/extracellular signal-regulated kinases (ERK) cascade in the human breast carcinoma cells (97). Taken together, *C. albicans* can contribute to cancer progression by producing HO-1 and its derivatives, but this factor may not be its only promoter for cancer development.

Inflammatory response

Interactions between specific PAMPs and PRRs, such as TLR2, TLR4, dectin-2, dectin-1, etc., on the surface of epithelial and myeloid cells, activate the inflammatory cascades by triggering the expression and secretion of a broad range of molecules including cytokines, cell growth factors, cell adhesion molecules, and immune receptors (98, 99). In immunocompromised cancer patients with a reduced number of leukocytes and other inflammatory mediators, circulating tumor cells can adhere and attach to the endothelium instead of leukocytes, which could potentially be the first step in creating secondary tumors and metastasis (67).

Inflammatory response of endothelial cells mediates high tumor cell adhesion and metastasis after being stimulated by *C. albicans*. *Candida* recognition by PRRs leads to stimulation and activation of multiple intracellular signaling pathways including Nuclear factor of activated T-cells (NFAT), NF- κ B, Mitogen-activated protein kinases (MAPK), and extracellular-signal-regulated kinase (ERK), which result in the secretion of several cytokines like IL-1, IL-2, TNF- α , IL-8, IL-6, IL-10, and IL-12 (11). It is reported that *Candida* infection would interrupt the integrity of gut mucosa in the intestinal epithelial cells (IEC) model (100). Moreover, many findings indicate the prominent role of the

MAPK and NF- κ B pathways in inducing proinflammatory milieu in *Candida* infection. The extracellular signal-regulated kinase (Erk) MAPK pathway prevents apoptosis of CD8+ T cells by modulating the expression of Bcl-2-interacting mediator of cell death (BIM), B-cell lymphoma 2 (BCL-2), and B-cell lymphoma-extra-large (Bcl-XL) proteins (101). MKPs play an essential role in innate immune responses by negatively regulating MAPK. MKP-1^{-/-} mice produce hefty amounts of TNF- α , IL-10, IL-1 β , and IL-6 when challenged with LPS, making them hyper-responsive to endotoxin shock (102).

Furthermore, the NF- κ B pathway is another important route that exerts acute effects on the development and function of the immune system (102, 103). For instance, Gratacep *et al.* showed that high-level mucosal infection with *Candida* induces NF- κ B activity while low-level infection does not affect this pathway in NF- κ B activity (104). Also, this study demonstrated that only direct contact of the yeast to epithelial cells could induce NF- κ B (105, 106).

Numerous studies reported that *C. albicans* affect cancer progression and metastasis through proinflammatory pathways in a cytokines-dependent manner and expression of adhesion molecules (107). In this regard, in a previous study, we have shown that *C. albicans* infection in the tumor-bearing mice made a dysregulation in cytokine profiles and could facilitate tumor growth and skewed immune responses toward Th2 in the breast tumor microenvironment (64). Also, several experiments have shown that systemic candidiasis is accompanied by augmentation of anti-candida Th1-related responses that promote the secretion of various cytokines like IFN- γ , TNF- α , TGF- β , IL-6, IL-12, IL-15, and IL-2. In preclinical studies, proinflammatory cytokines, such as IL-2, IL-15, and TNF- α , exhibited adjuvant activity because they could up-regulate the protective anti-fungal Th1 response and block Th2 immune reaction (108).

On the other hand, an elevated Th17-induced inflammatory response may increase pathogenicity correlated with *C. albicans* survival and dissemination in the mouse model and impair protective immunity (109). Among the proinflammatory cytokines, TNF- α , IL-1 β , IL-6, IL-8, and colony-stimulating factors (CSFs) are essential cytokines involved in the host-*Candida* interactive communication. Although IL-1 β and IL-6 play an essential role in PMN infiltration, they are not as crucial as TNF- α in the anti-fungal innate response. Interestingly, previous infection with *C. albicans* can mediate cancer initiation and progression by increasing the final level of TNF- α and IL-18 (107).

Additionally, some investigations have suggested that TGF- β restricts the phagocytic capacity of activated monocytes-macrophages in *C. albicans* infection-bearing mice and leads to inhibition of IFN- γ -induced nitric oxide production, which may facilitate the progression of *C. albicans* infection (110). However, TGF- β plays dual roles in tumor environment and normal cells. Researchers argue that during the initial stages of tumor outgrowth, TGF β acts as a tumor suppressor, preventing its progression to malignancy (111). However, during late-stage human breast tumors, TGF- β expression increases, which can exert angiogenic and immunosuppressive effects in the tumor microenvironment facilitating tumor progression (111, 112). Thus, invasive *Candida* infection in the chronic stages can significantly promote the spread of cancer by stimulating the secretion of TGF- β as an immunosuppressive agent.

In addition to the previously described mechanisms, evidence shows the dominant function of CD4+ T-cell subsets called Th17 cells, in response to *C. albicans* (113). Th17 cells are a fascinating subset that play significant roles in inflammatory diseases and protection against opportunistic pathogens and cancer (114). However, the role of IL-17 in candidiasis is controversial. Some mouse studies showed a potential association of IL-17 and IL-23 with candidiasis, but others have failed to find a strong connection (115). Most probably, IL-17 plays tissue-specific roles in immunity to *C. albicans*. Anti-*Candida* activity of IL-17 was first shown in 2004. Although Th17 is a directly essential cytokine against *Candida* infection, other Th17-related cytokines such as IL-23 showed tumorigenic and metastatic properties and influenced the pathogenic potential of Th17 cells in neoplastic microenvironments (116). Also, cancer progression is indirectly promoted by IL-17 via recruiting phagocytes, particularly neutrophils (117).

Molecular mimicry

The adhesion profile is considered the first step in initial fungal infection that leads to colonization, free dissemination, and invasive infections (118). Numerous investigations indicated that some *C. albicans* surface proteins such as complement receptor 3-related protein (CR3-RP), have structural and antigenic homology with glycoprotein CD11b/CD18 on leukocytes, which are essential agents for adhesion of leukocytes to the endothelial cells (119, 120). CD11b/CD18 is found in human neutrophils, monocytes, and macrophages. Polyclonal or monoclonal antibodies that recognize a subunit of CD11b/CD18 and target CR3-RP of *C. albicans* may crosstalk with CR3 of leukocytes and impair host anti-*Candida* and anti-tumor immunity. This is called molecular mimicry, favoring invasive *Candida* infection and cancer progression (121).

Epigenetic modification

Existing data suggest that pathogenic fungi can create diversity and genome plasticity in response to stressful growth conditions by chromosomal variation and increasing copy number (122, 123). Among the *Candida* species, *C. albicans* displays extensive genomic diversity and plasticity in the *de novo* format (123, 124). Asexual mitotic genome rearrangements have been identified as the central genomic diversity in *C. albicans* variants (125). Diverse repetitive loci of DNA are commonly compressed at ribosomal DNA (rDNA) sites, centromeres, and telomeres and are assembled into heterochromatin structures and organize long repetitive sequences that contribute to genotypic phenotypic plasticity (126, 127). There are at least four major groups of long repetitive sequences in *C. albicans*: tandem telomeric repeat, long/major tandem sequences (MRS), long terminal repeats (LTRs), and ribosomal DNA repeats (rDNA) (128-130). These long repetitive sequences experience recombination in both inter- and intra-genic occurrences that immediately generate long chromosomal polymorphisms, chimeric chromosomes, and telomere-telomere chromosomal fusions (131, 132).

Heterochromatic regions exert a transcriptionally repressive environment that can disseminate over long distances (up to 50kb), occasionally silencing native genes such as reporter genes inserted at these regions independently of the essential DNA sequence (133, 134). Histone modifiers can manage the transcriptionally repressive level of

heterochromatin regions through chromatin modifications. In this regard, Sitterlé *et al.* evaluated the chromatin states correlated with DNA repeats in *C. albicans*. Their results indicated that in this species, differential heterochromatin states regulate gene expression independently of the DNA sequence, and heterochromatin remodeling is associated with adaptation in a stress situation (135). Furthermore, numerous studies have reported that some non-coding RNAs including microRNAs, have a noticeable capacity to regulate proto-oncogenes such as PIM-1 (136). Since the expression of miRNAs occurs in tissues and a tumor-specific manner, it seems that some miRNAs are subject to epigenetic regulation.

Circular RNAs (circRNAs) generated during the alternative RNA splicing process could promote breast cancer cell progression under hypoxia (137). In addition, some studies have indicated that circRNAs might also be involved in breast cancer proliferation and migration (138). On the other hand, defective matches can be established in circRNA-miRNA duplex, enabling circRNAs to serve as "miRNA sponges" and suppress miRNA-mediated degradation of mRNAs (139). Gene ontology (GO) enrichment analysis indicates that invasive *Candida* may influence the regulation of respiratory epithelial functions by interference in different miRNA expressions and alteration of many critical biological pathways (140). Also, the results of one study showed that heat-killed *C. albicans*, accompanied by other factors triggering the NF- κ B and anti-inflammatory cytokines, could induce/inhibit specific miRNAs and regulate functions of innate immune cells such as macrophages following PRR stimulation (141) (Figure 3).

Therapeutic approaches

Due to the high compatibility and flexibility of *C. albicans* to grow in different host niches, it has been identified as an important and prevalent species in cancer incidence and development (142). Now, many fungicidal agents/drugs have been developed to treat systemic candidiasis infection. The most common structural classes of anti-fungal agents include polyenes (the oldest class of anti-fungal drugs), fluoropyrimidines, echinocandins, and azoles. Each of them constitutes several subclasses that pursue different pathways to eradicate fungal-related infections (143). The innovation of new alternative compounds, alone or in combination with other anti-fungal agents can increase anti-fungal capacity (144). In this regard, Onyewu and Heitman reported that the combination of 4T1 cell lysates and heated *C. albicans* extract, by induction of both innate and adaptive immunity, amplified anti-tumor immune responses, enhanced survival rates, and reduced tumor volume in the murine model of breast cancer (143, 144).

Comprehensive and exploratory studies evaluated the anti-morphogenetic properties of thirty anti-cancer agents on the yeast to hyphal form transition of *C. albicans* and provided the possibility of repurposing and designing cancer drugs as anti-morphogenetic agents in cancer-related *C. albicans* infection (145, 146). A research study indicated estrogen antagonist tamoxifen suppressed the growth and dissemination of fluconazole (FCZ)-sensitive *C. albicans* isolated from periodontal patients (147). However, tamoxifen's exact mechanism of action as an anti-fungal agent is unknown. The drug may be involved in preventing pathogenic fungal growth and development by induction of calcium-calcineurin signaling pathway and blocking

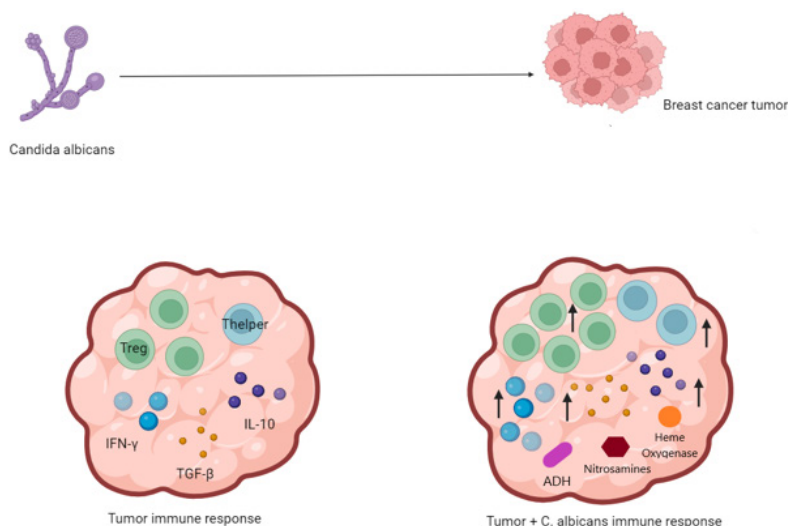


Figure 3. *Candida albicans* can modulate tumor microenvironment through secretion of different molecules
ADH: Alcohol dehydrogenases

calmodulin signaling (148, 149).

Many fungal pathogens evolved anti-oxidative factors to mediate survival during infection. But, in hypoxic and anoxic circumstances, neutrophil's natural function to efficiently generate reactive oxygen species is impaired. To this end, studies have shown that disrupting fungal redox balance could be a new therapeutic approach for producing effective and suitable drugs. Tempol, a redox-cycling nitroxide, is an anti-cancer and anti-inflammatory drug that has recently been proven to have anti-fungal effects, especially in response to systemic *C. albicans* (150). Besides, the repurposing of traditional compounds/drugs for new targets can shorten the treatment time and provide rapid therapy and novel opportunities to develop de novo anti-fungal agents (151).

Although the demand and consumption of fungicidal drugs in modern healthcare have increased significantly over the last decades, the rate of available therapeutics may not meet these demands. From another perspective, due to the primal evolutionary relationship between fungi and humans, some drugs that have a cytotoxic effect on fungi may be deleterious to humans (151, 152). Therefore, the use of promising, novel techniques and the combination of innovative screening methods with new chemical formulations can significantly advance the field of personalized drug discovery to counteract fungal and invasive *Candida* infection and subsequently hinder cancer progression.

Conclusion

Innovation in drug development is highly demanded to control and eliminate candidiasis in cancer patients and consequently prevent cancer progression. However, a simultaneous anti-tumor effect should be another property of this concern.

Authors' Contributions

M M and AA A conceived and designed the study. MM AS, P K, and FA H performed the study. P K, M B, and S A wrote the manuscript. MH Y, RSE, and M M actively revised the article. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest

None.

References

- Rohan TE, Xue Y, Li H-M, D'Alfonso TM, Ginter PS, Oktay MH, et al. Tumor microenvironment of metastasis and risk of distant metastasis of breast cancer. *J Natl Cancer Inst* 2014;106:1-11.
- Islam F, Miller RD, Siegel RL, Zheng Z, Zhao J, Han X, et al. National and state estimates of lost earnings from cancer deaths in the United States. *JAMA oncology* 2019;5:e191460.
- Tajeri M, Seyedolmohadesin M, Bayat M, Mahdavi M, Yazdi MI, Esamifar A, et al. The effect of *Candida albicans* systemic infection on matrix metalloproteinases in breast cancer bearing BALB/c mice. *Iran J Allergy Asthma Immunol* 2013;12:81-85.
- Sobel JD. Vulvovaginal candidosis. *The Lancet* 2007;369:1961-1971.
- Chung L-M, Liang J-A, Lin C-L, Sun L-M, Kao C-H. Cancer risk in patients with candidiasis: a nationwide population-based cohort study. *Oncotarget* 2017;8:63562-63573.
- Klimesova K, Jiraskova Zakostelska Z, Tlaskalova-Hogenova H. Oral bacterial and fungal microbiome impacts colorectal carcinogenesis. *Front Microbiol* 2018;9:774.
- Coker OO, Nakatsu G, Dai RZ, Wu WKK, Wong SH, Ng SC, et al. Enteric fungal microbiota dysbiosis and ecological alterations in colorectal cancer. *Gut* 2019;68:654-662.
- Jacobsen ID, Wilson D, Wächtler B, Brunke S, Naglik JR, Hube B. *Candida albicans* dimorphism as a therapeutic target. *Expert Rev Anti Infect Ther* 2012;10:85-93.
- Dühring S, Germerodt S, Skerka C, Zipfel PF, Dandekar T, Schuster S. Host-pathogen interactions between the human innate immune system and *Candida albicans*—understanding and modeling defense and evasion strategies. *Front Microbiol* 2015; 6:625.
- Qin Y, Zhang L, Xu Z, Zhang J, Jiang Y-y, Cao Y, Yan T. Innate immune cell response upon *Candida albicans* infection. *Virulence* 2016;7:512-526.
- Takeuchi O, Akira S. Pattern recognition receptors and inflammation. *Cell* 2010;140:805-820.
- Taghavi M, Khosravi A, Mortaz E, Nikaein D, Athari SS. Role of pathogen-associated molecular patterns (PAMPS) in immune responses to fungal infections. *Eur J Pharmacol* 2017;808:8-13.
- Mech F, Wilson D, Lehnert T, Hube B, Thilo Figge M. Epithelial invasion outcompetes hypha development during *Candida albicans* infection as revealed by an image-based systems biology approach. *Cytometry Part A* 2014;85:126-139.

14. Naglik JR, König A, Hube B, Gaffen SL. Candida albicans–epithelial interactions and induction of mucosal innate immunity. *Curr Opin Microbiol* 2017;40:104-112.
15. Mayer FL, Wilson D, Hube B. Candida albicans pathogenicity mechanisms. *Virulence* 2013;4:119-128.
16. Wilson D, Thewes S, Zakikhany K, Fradin C, Albrecht A, Almeida R, et al. Identifying infection-associated genes of Candida albicans in the postgenomic era. *FEMS Yeast Res* 2009;9:688-700.
17. Li M, Chen Q, Tang R, Shen Y, Da Liu W. The expression of β -defensin-2, 3 and LL-37 induced by Candida albicans phospholipomannan in human keratinocytes. *J Dermatol Sci* 2011;61:72-75.
18. Vautier S, da Glória Sousa M, Brown GD. C-type lectins, fungi and Th17 responses. *Cytokine Growth Factor Rev* 2010;21:405-412.
19. Paulone S, Ardizzoni A, Tavanti A, Piccinelli S, Rizzato C, Lupetti A, et al. The synthetic killer peptide KP impairs Candida albicans biofilm *in vitro*. *PLoS One* 2017;12:e0181278.
20. Rodríguez-Cerdeira C, Gregorio MC, Molares-Vila A, López-Barcenas A, Fabbrocini G, Bardhi B, et al. Biofilms and vulvovaginal candidiasis. *Colloids and Surfaces B: Biointerfaces* 2019;174:110-125.
21. Moyes DL, Naglik JR. Mucosal immunity and Candida albicans infection. *Clin Dev Immunol* 2011;2011:346307.
22. De Luca A, Zelante T, D'angelo C, Zagarella S, Fallarino F, Spreca A, et al. IL-22 defines a novel immune pathway of antifungal resistance. *Mucosal Immunol* 2010;3:361-373.
23. Eyerich S, Wagener J, Wenzel V, Scarponi C, Pennino D, Albanesi C, et al. IL-22 and TNF- α represent a key cytokine combination for epidermal integrity during infection with Candida albicans. *Eur J Immunol* 2011;41:1894-1901.
24. Luo S, Skerka C, Kurzai O, Zipfel PF. Complement and innate immune evasion strategies of the human pathogenic fungus Candida albicans. *Mol Immunol* 2013;56:161-169.
25. Miramón P, Dunker C, Windecker H, Bohovych IM, Brown AJ, Kurzai O, Hube B. Cellular responses of Candida albicans to phagocytosis and the extracellular activities of neutrophils are critical to counteract carbohydrate starvation, oxidative and nitrosative stress. *PLoS One* 2012;7:e52850.
26. Lefkowitz SS, Gelderman MP, Lefkowitz DL, McGuilevsky N, Bollen A. Phagocytosis and intracellular killing of Candida albicans by macrophages exposed to myeloperoxidase. *Infect Dis* 1996;173:1202-1207.
27. Urban CF, Reichard U, Brinkmann V, Zychlinsky A. Neutrophil extracellular traps capture and kill Candida albicans yeast and hyphal forms. *Cellul Microbiol* 2006;8:668-670.
28. Jimenez-Lopez C, Lorenz MC. Fungal immune evasion in a model host–pathogen interaction: Candida albicans versus macrophages. *PLoS Pathog* 2013;9:e1003741.
29. Krysan DJ, Sutterwala FS, Wellington M. Catching fire: Candida albicans, macrophages, and pyroptosis. *PLoS Pathog* 2014;10:e1004139.
30. Takao S, Smith EH, Wang D, Chan C, Bulkley GB, Klein AS. Role of reactive oxygen metabolites in murine peritoneal macrophage phagocytosis and phagocytic killing. *Am J Physiol* 1996;271(4 Pt 1):C1278-84.
31. Liu P, Wu X, Liao C, Liu X, Du J, Shi H, et al. Escherichia coli and Candida albicans induced macrophage extracellular trap-like structures with limited microbicidal activity. *PLoS One* 2014;9:e90042.
32. Cheng S-C, Joosten LA, Kullberg B-J, Netea MG. Interplay between Candida albicans and the mammalian innate host defense. *Infect Immun* 2012;80:1304-1313.
33. Ramirez-Ortiz ZG, Means TK. The role of dendritic cells in the innate recognition of pathogenic fungi (A. fumigatus, C. neoformans and C. albicans). *Virulence* 2012;3:635-646.
34. d'Ostiani CF, Del Sero G, Bacci A, Montagnoli C, Spreca A, Mencacci A, et al. Dendritic cells discriminate between yeasts and hyphae of the fungus Candida albicans: Implications for initiation of T helper cell immunity *in vitro* and *in vivo*. *J Exp Med* 2000;191:1661-1674.
35. Kashem SW, Igyártó BZ, Gerami-Nejad M, Kumamoto Y, Mohammed J, Jarrett E, et al. Candida albicans morphology and dendritic cell subsets determine T helper cell differentiation. *Immunity* 2015;42:356-366.
36. Roudbary M, Roudbar Mohammadi S, Bozorgmehr M, Moazzeni SM. The effects of Candida albicans cell wall protein fraction on dendritic cell maturation. *Iran J Immunol* 2009;6:67-74.
37. van de Veerdonk FL, Marijnissen RJ, Kullberg BJ, Koenen HJ, Cheng S-C, Joosten I, et al. The macrophage mannose receptor induces IL-17 in response to Candida albicans. *Cell Host Microbe* 2009;5:329-340.
38. Lin L, Ibrahim AS, Xu X, Farber JM, Avanesian V, Baquir B, et al. Th1-Th17 cells mediate protective adaptive immunity against Staphylococcus aureus and Candida albicans infection in mice. *PLoS pathog* 2009;5:e1000703.
39. Wood SM, Ljunggren H-G, Bryceson YT. Insights into NK cell biology from human genetics and disease associations. *Cell Mol Life Sci* 2011;68:3479-3493.
40. Voigt J, Hünninger K, Bouzani M, Jacobsen ID, Barz D, Hube B, et al. Human natural killer cells act as phagocytes against Candida albicans and mounting an inflammatory response that modulates neutrophil antifungal activity. *Infect Dis* 2014;209:616-626.
41. Martín-Antónic I, Sufre G, Perez-Amill L, Castella M, Urbano-Ispizua A. Natural killer cells: Angels and devils for immunotherapy. *Int J Mol Sci* 2017;18:1868.
42. Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of natural killer cells. *Nat Immunol* 2008;9:503-510.
43. Quentin J, Voigt J, van der Voort R, Jacobsen ID, Verschueren I, Hube B, et al. Differential role of NK cells against Candida albicans infection in immunocompetent or immunocompromised mice. *Eur J Immunol* 2014;44:2405-2414.
44. Monifazi P, Zelante T, D'angelo C, De Luca A, Moretti S, Bozza S, et al. Balancing inflammation and tolerance *in vivo* through dendritic cells by the commensal Candida albicans. *Mucosal Immunol* 2009;2:362-374.
45. Huang H, Ostroff GR, Lee CK, Wang JP, Specht CA, Levitz SM. Distinct patterns of dendritic cell cytokine release stimulated by fungal β -glucans and toll-like receptor agonists. *Infect Immun* 2009;77:1774-1781.
46. Naglik JR. Candida immunity. *New J Sci* 2014;2014:390241.
47. Romani L. Immunity to fungal infections. *Nat Rev Immunol* 2004;4:1-23.
48. LeibundGut-Landmann S, Groß O, Robinson MJ, Osorio F, Slack EC, Tsoni SV, et al. Syk-and CARD9-dependent coupling of innate immunity to the induction of T helper cells that produce interleukin 17. *Nat Immunol* 2007;8:630-638.
49. Cenci E, Mencacci A, Spaccapelo R, Tonnetti L, Mosci P, Ennsle K-H, et al. T helper cell type 1 (Th1)-and Th2-like responses are present in mice with gastric candidiasis but protective immunity is associated with Th1 development. *J Infect Dis* 1995;171:1279-1288.
50. Fidel Jr PL. History and update on host defense against vaginal candidiasis. *Am J Reprod Immunol* 2007;57:2-12.
51. Gringhuis SI, Wevers BA, Kaptein TM, Van Capel TM, Theelen B, Boekhout T, et al. Selective C-Rel activation via Malt1 controls anti-fungal TH-17 immunity by dectin-1 and dectin-2. *PLoS Pathog* 2011;7:e1001259.
52. Pietrella D, Rachini A, Pines M, Pandey N, Mosci P, Bistoni F, et al. Th17 cells and IL-17 in protective immunity to vaginal candidiasis. *PLoS One* 2011;6:e22770.
53. Xu X, Wang R, Su Q, Huang H, Zhou P, Luan J, et al. Expression of Th1-Th2-and Th17-associated cytokines in laryngeal carcinoma. *Oncol Lett* 2016;12:1941-1948.
54. Bober LA, Grace MJ, Pugliese-Sivo C, Rojas-Triana A, Waters T, Sullivan LM, et al. The effect of GM-CSF and G-CSF on human

- neutrophil function. *Immunopharmacology* 1995;29:111-119.
55. Weisbart RH, Kwan L, Golde DW, Gasson JC. Human GM-CSF primes neutrophils for enhanced oxidative metabolism in response to the major physiological chemoattractants. *Blood* 1987;69:18-21.
 56. Dale DC, Liles WC, Llewellyn C, Price TH. Effects of granulocyte-macrophage colony-stimulating factor (GM-CSF) on neutrophil kinetics and function in normal human volunteers. *Am J Hematol* 1998;57:7-15.
 57. Kasahara S, Jhingran A, Dhingra S, Salem A, Cramer RA, Hohl TM. Role of granulocyte-macrophage colony-stimulating factor signaling in regulating neutrophil antifungal activity and the oxidative burst during respiratory fungal challenge. *J Infect Dis* 2016;213:1289-1298.
 58. Pandiyan P, Conti HR, Zheng L, Peterson AC, Mathern DR, Hernández-Santos N, et al. CD4+ CD25+ Foxp3+ regulatory T cells promote Th17 cells *in vitro* and enhance host resistance in mouse *Candida albicans* Th17 cell infection model. *Immunity* 2011;34:422-434.
 59. Josefowicz SZ, Lu L-F, Rudensky AY. Regulatory T cells: mechanisms of differentiation and function. *Annu Rev Immunol* 2012;30:531-564.
 60. Kekäläinen E, Tuovinen H, Joensuu J, Gylling M, Franssila R, Pöntynen N, et al. A defect of regulatory T cells in patients with autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy. *J Immunol* 2007;178:1208-1215.
 61. Kirchner FR, Littringer K, Altmeier S, Tran VDT, Schönherr F, Lemberg C, et al. Persistence of *Candida albicans* in the oral mucosa induces a curbed inflammatory host response that is independent of immunosuppression. *Front Immunol* 2019;10:330.
 62. Whibley N, MacCallum DM, Vickers MA, Zafreen S, Waldmann H, Hori S, et al. Expansion of Foxp3+ T-cell populations by *Candida albicans* enhances both Th17-cell responses and fungal dissemination after intravenous challenge. *Eur J Immunol* 2014;44:1069-1083.
 63. Ahmadi N, Ahmadi A, Kheirali E, Yadegari MH, Bayat M, Shajiei A, et al. Systemic infection with *Candida albicans* in breast tumor bearing mice: Cytokines dysregulation and induction of regulatory T cells. *J Mycol Med* 2019;29:49-55.
 64. Conche C, Greten FR. Fungi enter the stage of colon carcinogenesis. *Immunity*. 2018;49:384-386.
 65. Yu D, Liu Z. The research progress in the interaction between *Candida albicans* and cancers. *Frontiers in Microbiology*. 2022;13:988734.
 66. Ramirez-Garcia A, Rementeria A, Aguirre-Urizar JM, Moragues MD, Antoran A, Pellon A, et al. *Candida albicans* and cancer: Can this yeast induce cancer development or progression? *Crit Rev Microbiol* 2016;42:181-193.
 67. Swann P, Magee P. Nitrosamine-induced carcinogenesis. The alkylation of nucleic acids of the rat by N-methyl-N-nitrosourea, dimethylnitrosamine, dimethyl sulphate and methyl methanesulphonate. *Biochem J* 1968;110:39-47.
 68. Archer M. Mechanisms of action of N-nitroso compounds. *Cancer Surv* 1989;8:241-250.
 69. Larsson SC, Bergkvist L, Wolk A. Processed meat consumption, dietary nitrosamines and stomach cancer risk in a cohort of Swedish women. *Int J Cancer* 2006;119:915-919.
 70. Song P, Wu L, Guan W. Dietary nitrates, nitrites, and nitrosamines intake and the risk of gastric cancer: A meta-analysis. *Nutrients* 2015;7:9872-9895.
 71. Chu F, Li G. Simultaneous occurrence of fumonisin B1 and other mycotoxins in moldy corn collected from the People's Republic of China in regions with high incidences of esophageal cancer. *Appl Environ Microbiol* 1994;60:847-852.
 72. Gankhuyag N, Lee K-H, Cho J-Y. The role of nitrosamine (NNK) in breast cancer carcinogenesis. *J Mammary Gland Biol Neoplasia* 2017;22:159-170.
 73. Krogh P. The role of yeasts in oral cancer by means of endogenous nitrosation. *Acta Odontol Scand* 1990;48:85-88.
 74. Nagy K, Sonkodi I, Szöke I, Nagy E, Newman H. The microflora associated with human oral carcinomas. *Oral oncol* 1998;34:304-308.
 75. Jayachandran AL, Katragadda R, Thyagarajan R, Vajravelu L, Manikesi S, Kaliappan S, Jayachandran B. Oral Candidiasis among cancer patients attending a tertiary Care Hospital in Chennai, South India: an evaluation of Clinicomycological association and antifungal susceptibility pattern. *Can J Infect Dis Med Microbiol* 2016;2016:8758461.
 76. Tamgadge S, Tamgadge A, Pillai A. Association of *Candida* sp. with the Degrees of Dysplasia and Oral Cancer: A study by calcofluor white under fluorescent microscopy. *Iran J Pathol* 2017;12:348-355.
 77. Gupta V, Abhishek K, Balasundari S, Devendra NK, Shadab K, Anupama M. Identification of *Candida albicans* using different culture media and its association in leukoplakia and oral squamous cell carcinoma. *J Oral Maxillofac Pathol* 2024;28:23-28.
 78. Holt JT, Thompson ME, Szabo C, Robinson-Benion C, Arteaga CL, King M-C, Jensen RA. Growth retardation and tumour inhibition by BRCA1. *Nat Genet* 1996;17:298-302.
 79. Humphrey JS, Salim A, Erdos MK, Collins FS, Brody LC, Klausner RD. Human BRCA1 inhibits growth in yeast: Potential use in diagnostic testing. *Proc Natl Acad Sci U S A* 1997;94:5820-5825.
 80. Lin NU, Claus E, Sohl J, Razzari AR, Arnaout A, Winer EP. Sites of distant recurrence and clinical outcomes in patients with metastatic triple-negative breast cancer: High incidence of central nervous system metastases. *Cancer* 2008;113:2638-2645.
 81. Gainza-Ciraqui ML, Nieminen MT, Novak Frazer L, Aguirre-Urizar JM, Moragues MD, Rautemaa R. Production of carcinogenic acetaldehyde by *Candida albicans* from patients with potentially malignant oral mucosal disorders. *J Oral Pathol Med* 2013;42:245-249.
 82. Seitz HK, Suckel F. Acetaldehyde as an underestimated risk factor for cancer development: role of genetics in ethanol metabolism. *Genes Nutr* 2010;5:121-128.
 83. Manzo-Avalos S, Saavedra-Molina A. Cellular and mitochondrial effects of alcohol consumption. *Int J Environ Res Public Health* 2010;7:4281-304.
 84. Humans IWGotEoCRt, Cancer IAfRo, Organization WH. Re-evaluation of Some Organic Chemicals, Hydrazine and Hydrogen Peroxide: Other compounds reviewed in plenary sessions. *IARC Monogr Eval Carcinog Risks Hum*. 1999;71:1-1554.
 85. McCullough M, Jaber M, Barrett A, Bain L, Speight P, Porter S. Oral yeast carriage correlates with presence of oral epithelial dysplasia. *Oral Oncol* 2002;38:391-393.
 86. Nieminen MT, Novak-Frazer L, Rautemaa V, Rajendran R, Sorsa T, Ramage G, et al. A novel antifungal is active against *Candida albicans* biofilms and inhibits mutagenic acetaldehyde production *in vitro*. *PLoS One* 2014;9:e97864.
 87. Abraham J, Balbo S, Crabb D, Brooks PJ. Alcohol metabolism in human cells causes DNA damage and activates the fanconi anemia-breast cancer susceptibility (FA-BRCA) DNA damage response network. *Alcoholism: Alcohol Clin Exp Res* 2011;35:2113-2120.
 88. Kim D, Yukl ET, Moënne-Loccoz P, Ortiz de Montellano PR. Fungal heme oxygenases: functional expression and characterization of Hmx1 from *Saccharomyces cerevisiae* and CaHmx1 from *Candida albicans*. *Biochemistry*. 2006;45:14772-14780.
 89. Santos R, Buisson N, Knight S, Dancis A, Camadro J-M, Lesuisse E. Haemin uptake and use as an iron source by *Candida albicans*: Role of CaHMX1-encoded haem oxygenase. *Microbiology* 2003;149:579-588.
 90. Overhaus M, Moore BA, Barbato JE, Behrendt FF, Doering JG, Bauer AJ. Biliverdin protects against polymicrobial sepsis by modulating inflammatory mediators. *Am J Physiol Gastrointest Liver Physiol* 2006;290:G695-703.
 91. Manns JM, Mosser DM, Buckley HR. Production of a hemolytic

- factor by *Candida albicans*. *Infect Immun* 1994;62:5154-156.
92. Weissman Z, Kornitzer D. A family of *Candida* cell surface haem-binding proteins involved in haemin and haemoglobin-iron utilization. *Mol Microbiol* 2004;53:1209-1220.
 93. Okamoto-Shibayama K, Kikuchi Y, Kokubu E, Sato Y, Ishihara K. Csa2, a member of the Rbt5 protein family, is involved in the utilization of iron from human hemoglobin during *Candida albicans* hyphal growth. *FEMS Yeast Res* 2014;14:674-767.
 94. Coburn R, Williams W, Kahn S. Endogenous carbon monoxide production in patients with hemolytic anemia. *J Clin Invest* 1966;45:460-468.
 95. Lin C-W, Shen S-C, Hou W-C, Yang L-Y, Chen Y-C. Heme oxygenase-1 inhibits breast cancer invasion via suppressing the expression of matrix metalloproteinase-9. *Mol Cancer Ther* 2008;7:1195-1206.
 96. Gandini NA, Alonso EN, Fermento ME, Mascará M, Abba MC, Coló GP, et al. Heme oxygenase-1 has an antitumor role in breast cancer. *Antioxid Redox Signal* 2019;30:2030-2049.
 97. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* 2006;124:783-801.
 98. Mogensen TH. Pathogen recognition and inflammatory signaling in innate immune defenses. *Clin Microbiol Rev* 2009;22:240-273.
 99. Böhringer M, Pohl S, Schulze S, Albrecht-Eckardt D, Piegsa J, Weber M, et al. *Candida albicans* infection leads to barrier breakdown and a MAPK/NF- κ B mediated stress response in the intestinal epithelial cell line C2BBel. *Cell Microbiol* 2016;18:889-904.
 100. D'Souza WN, Chang C-F, Fischer AM, Li M, Hedrick SM. The Erk2 MAPK regulates CD8 T cell proliferation and survival. *J Immunol* 2008;181:7617-7629.
 101. Oh H, Ghosh S. NF- κ B: Roles and regulation in different CD4+ T-cell subsets. *Immunol Rev* 2013;252:41-51.
 102. Gerondakis S, Siebenlist U. Roles of the NF- κ B pathway in lymphocyte development and function. *Cold Spring Harb Perspect Biol* 2010 May;2:a000182.
 103. Gratacap RL, Rawls JF, Wheeler RT. Mucosal candidiasis elicits NF- κ B activation, proinflammatory gene expression and localized neutrophilia in zebrafish. *Dis Model Mech* 2013;6:1260-1270.
 104. Ali S, Lazennec G. Chemokines: novel targets for breast cancer metastasis. *Cancer Metastasis Rev* 2007;26:401-420.
 105. Esquivel-Velázquez M, Ostoa-Saloma P, Palacios-Arreola MI, Nava-Castro KE, Castro JI, Morales-Montero J. The role of cytokines in breast cancer development and progression. *J Interferon Cytokine Res* 2015;35:1-16.
 106. Rodríguez-Cuesta J, Hernando FL, Mendoza L, Gallot N, de Cerio AAD, Martínez-de-Tejada C, Vidal-Vanaclocha F. *Candida albicans* enhances experimental hepatic melanoma metastasis. *Clin Exp Metastasis* 2010;27:35-42.
 107. Pikman R, Ben-Ami R. Immune modulators as adjuncts for the prevention and treatment of invasive fungal infections. *Immunotherapy* 2012;4:1869-1882.
 108. Zelante T, De Luca A, Bonifazi P, Montagnoli C, Bozza S, Moretti S, et al. IL-23 and the Th17 pathway promote inflammation and impair antifungal immune resistance. *Eur J Immunol* 2007;37:2695-2706.
 109. Vodovotz Y, Bogdan C, Paik J, Xie Q, Nathan C. Mechanisms of suppression of macrophage nitric oxide release by transforming growth factor beta. *J Exp Med* 1993;178:605-613.
 110. Derynck R, Akhurst RJ, Balmain A. TGF- β signaling in tumor suppression and cancer progression. *Nat Genet* 2001;29:117-129.
 111. Walker RA, Dearing SJ. Transforming growth factor beta1 in ductal carcinoma in situ and invasive carcinomas of the breast. *Eur J Cancer* 1992;28:641-644.
 112. Acosta-Rodríguez EV, Rivino L, Geginat J, Jarrossay D, Gattorno M, Lanzavecchia A, et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. *Nat Immunol* 2007;8:639-646.
 113. Stockinger B, Omenetti S. The dichotomous nature of T helper 17 cells. *Nat Rev Immunol* 2017;17:535-544.
 114. Yano J, Noverr MC, Fidel Jr PL. Cytokines in the host response to *Candida* vaginitis: Identifying a role for non-classical immune mediators, S100 alarmins. *Cytokine*. 2012;58:118-128.
 115. Huang W, Na L, Fidel PL, Schwarzenberger P. Requirement of interleukin-17A for systemic anti-*Candida albicans* host defense in mice. *J Infect Dis* 2004;190:624-631.
 116. Donskov F, von der Maase H. Impact of immune parameters on long-term survival in metastatic renal cell carcinoma. *J Clin Oncol* 2006;24:1997-2005.
 117. Moyes DL, Richardson JP, Naglik JR. *Candida albicans*-epithelial interactions and pathogenicity mechanisms: scratching the surface. *Virulence*. 2015;6:338-346.
 118. Gustafson K, Vercellotti G, Bendel C, Hostetter M. Molecular mimicry in *Candida albicans*. Role of an integrin analogue in adhesion of the yeast to human endothelium. *J Clin Invest* 1991;87:1896-1902.
 119. Dana N, Todd Rd, Pitt J, Springer TA, Arnaout MA. Deficiency of a surface membrane glycoprotein (Mo1) in man. *J Clin Invest* 1984;73:153-159.
 120. Gilmore BJ, Retsinas EM, Loren JS, Hostetter MK. An iC3b receptor on *Candida albicans*: structure, function, and correlates for pathogenicity. *J Infect Dis* 1988; 57:38-46.
 121. Forche A, Abbey D, Pithkul T, Weinzierl M, Ringstrom T, Bruck D, et al. Stressors rates and types of loss of heterozygosity in *Candida albicans*. *MBio* 2011;2:e00129-11.
 122. Forche A, Hunt J, Abbey D, Issi L, Guiducci C, Martinez DA, et al. The evolution of drug resistance in clinical isolates of *Candida albicans*. *Elife*. 2015;4:e00662.
 123. Hirakawa MP, Martinez DA, Sakthikumar S, Anderson MZ, Berlin A, Gujja S, et al. Genetic and phenotypic intra-species variation in *Candida albicans*. *Genome Res* 2015;25:413-425.
 124. Lephart PR, Magee PT. Effect of the major repeat sequence on mitotic recombination in *Candida albicans*. *Genetics* 2006;174:1737-1744.
 125. Braun BR, van Het Hoog M, d'Enfert C, Martchenko M, Dungan J, Kuo A, et al. A human-curated annotation of the *Candida albicans* genome. *PLoS Genet* 2005;1:e21.
 126. Jones T, Federspiel NA, Chibana H, Dungan J, Kalman S, Magee B, et al. The diploid genome sequence of *Candida albicans*. *Proc Natl Acad Sci U S A* 2004 ;101:7329-7334.
 127. Chibana H, Iwaguchi S, Homma M, Chindamporn A, Nakagawa Y, Tanaka K. Diversity of tandemly repetitive sequences due to short periodic repetitions in the chromosomes of *Candida albicans*. *J Bacteriol*1994;176:3851-3858.
 128. Goodwin TJ, Poulter RT. The CARE-2 and rel-2 repetitive elements of *Candida albicans* contain LTR fragments of a new retrotransposon. *Gene* 1998;218:85-93.
 129. Goodwin TJ, Poulter RT. Multiple LTR-retrotransposon families in the asexual yeast *Candida albicans*. *Genome Res* 2000;10:174-191.
 130. Selmecki A, Forche A, Berman J. Aneuploidy and isochromosome formation in drug-resistant *Candida albicans*. *Science*. 2006;313:367-370.
 131. Selmecki A, Forche A, Berman J. Genomic plasticity of the human fungal pathogen *Candida albicans*. *Eukaryot Cell* 2010;9:991-1008.
 132. Kadosh D. Shaping up for battle: morphological control mechanisms in human fungal pathogens. *PLoS Pathog* 2013;9:e1003795.
 133. Rusche LN, Kirchmaier AL, Rine J. The establishment, inheritance, and function of silenced chromatin in *Saccharomyces cerevisiae*. *Annu Rev Biochem* 2003;72:481-516.
 134. Sitterlé E, Maufrais C, Sertour N, Palayret M, d'Enfert C, Bougnoux M-E. Within-host genomic diversity of *Candida albicans* in healthy carriers. *Sci Rep* 2019 ;9:2563.

135. Thomas M, Lange-Grünweller K, Weirauch U, Gutsch D, Aigner A, Grünweller A, *et al.* The proto-oncogene Pim-1 is a target of miR-33a. *Oncogene* 2012;31:918-928.
136. Liang G, Liu Z, Tan L, Su A, Jiang WG, Gong C. HIF1 α -associated circDENND4C promotes proliferation of breast cancer cells in hypoxic environment. *Anticancer Res* 2017;37:4337-4343.
137. He R, Liu P, Xie X, Zhou Y, Liao Q, Xiong W, *et al.* circGFRA1 and GFRA1 act as ceRNAs in triple negative breast cancer by regulating miR-34a. *J Exp Clin Cancer Res* 2017;36:145.
138. Guil S, Esteller M. RNA-RNA interactions in gene regulation: the coding and noncoding players. *Trends Biochem Sci* 2015;40:248-256.
139. Muhammad SA, Fatima N, Syed N-i-H, Wu X, Yang XF, Chen JY. MicroRNA expression profiling of human respiratory epithelium affected by invasive *Candida* infection. *PLoS One* 2015;10:e0136454.
140. Monk CE, Hutvagner G, Arthur JSC. Regulation of miRNA transcription in macrophages in response to *Candida albicans*. *PLoS One* 2010;5:e13669.
141. Ramirez-Garcia A, Gallot N, Abad A, Mendoza L, Rementeria A, Hernando FL. Molecular fractionation and characterization of a *Candida albicans* fraction that increases tumor cell adhesion to hepatic endothelium. *Appl Microbiol Biotechnol* 2011;92:133-145.
142. Onyewu C, Heitman J. Unique applications of novel antifungal drug combinations. *Anti-Infective Agents in Medicinal Chemistry*. 2007;6:3-15.
143. Wakharde AA, Halbandge SD, Phule DB, Karuppaiyl SM. Anticancer drugs as antibiofilm agents in *Candida albicans*: Potential targets. *Assay Drug Dev Technol* 2018;16:232-246.
144. Routh MM, Chauhan NM, Karuppaiyl SM. Cancer drugs inhibit morphogenesis in the human fungal pathogen, *Candida albicans*. *Braz J Microbiol* 2013;44:855-859.
145. Routh MM, Raut JS, Karuppaiyl SM. Dual properties of anticancer agents: an exploratory study on the *in vitro* anti-*Candida* properties of thirty drugs. *Chemotherapy* 2011;57:372-380.
146. Muthular M, Bálamo F, Passero P, Jewtuchowicz V, Miozza V, Villalba MB, *et al.* Effects of tamoxifen on periodontal disease and *Candida albicans* of patients with breast cancer and other pathologies. *Future microbiol* 2019;14:129-137.
147. Liu S, Hou Y, Liu W, Lu C, Wang W, Sun S. Components of the calcium-calcineurin signaling pathway in fungal cells and their potential as antifungal targets. *Eukaryot Cell* 2015;14:324-334.
148. Dolan K, Montgomery S, Buchheit B, DiDone L, Wellington M, Krysan DJ. Antifungal activity of tamoxifen: *In vitro* and *in vivo* activities and mechanistic characterization. *Antimicrob Agents Chemother* 2009;53:3337-3346.
149. Hosseinzadeh A, Stylianou M, Lopes JP, Müller DC, Häggman A, Holmberg S, *et al.* Stable redox-cycling nitroxide tempol has antifungal and immune-modulatory properties. *Front Microbiol* 2019;10:1843.
150. Moran GP, Coleman DC, Sullivan DJ. Comparative genomics and the evolution of pathogenicity in human pathogenic fungi. *Eukaryot Cell* 2011;10:34-42.
151. Taylor JW. Evolutionary perspectives on human fungal pathogens. *Cold Spring Harb Perspect Med* 2014;5: a019588.

