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Effect of mild gestational diabetes mellitus on histological, ultrastructural, and quantitative morphometric alterations of rat fetal liver

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Objective(s): Gestational diabetes mellitus (GDM), one of the most common metabolic disorders in pregnancy, impacts maternal and fetal health. This study was designed to assess the effects of mild GDM on the histology, ultrastructure, and morphometry of fetal liver tissue.

Materials and Methods: In this experimental study, twenty pregnant rats were randomly allocated into control and streptozotocin (STZ)-induced diabetic groups. Mild hyperglycemia was induced by intraperitoneal injection of STZ (40 mg/kg/bw) on the 5th day of gestation. At day 19 of gestation, fetal livers were separated and subjected to histological, transmission electron microscopic, and quantitative morphometric examinations.

Results: In the GDM group, PAS staining was positive, revealing scattered eosinophilic inclusions in some hepatocytes. Masson trichrome staining was also positive and showed some fibrous tissue as fine fibers in the portal spaces that extended to the central vein. Reticulin staining in the GDM group was focally positive in the areas of fibrosis and the portal spaces. Ultrastructural examination showed pyknotic nuclei, karyolysis, degranulation and vesiculation of the rough endoplasmic reticulum, and degeneration of mitochondria in the GDM group. The morphometric examination demonstrated that the mean area of hepatocytes was significantly lower in the GDM group than in the control group (*P*<0.05). Moreover, the mean diameter of the central vein and the density of megakaryocytes were significantly higher in the GDM group than in the control group (*P*<0.05).

Conclusion: Uncontrolled mild GDM induced the histological, ultrastructural and morphometric alterations in the fetal liver.

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Introduction

Diabetes mellitus (DM), as one of the leading causes of death in the world, imposes a heavy global burden on public health (1).

Gestational diabetes mellitus (GDM) is the most common medical complication and metabolic disturbance of pregnancy (2). It has attracted considerable research attention due to its severe risks and adverse health effects.

Untreated GDM can lead to short and long-term complications for the mother and fetus, including gestational hypertension, cesarean section, preeclampsia, shoulder dystocia, birth trauma, macrosomia, neonatal hypoglycemia, hypocalcemia, and hyperbilirubinemia (3, 4).

The elevated transfer of glucose from the mother with DM to the fetus leads to hyperglycemia and hyperinsulinemia in the fetus, which in turn stimulates the growth of insulindependent tissues and organs such as the liver (5-9).

Despite extensive studies on the adverse effects of DM, limited data are available regarding the impact of mild GDM on the fetal liver structure. In the present study, an attempt was made to investigate the effects of mild GDM on the histology, ultrastructure, and morphometry of fetal liver tissue in streptozotocin (STZ)-induced diabetic rats as an experimental model.

Materials and Methods

This study was approved by the Ethics Committee of Golestan University of Medical Sciences, Gorgan, Iran (IR. GOUMS.AEC.1401.007). The experimental procedures were conducted in line with the Guide for the Care and Use of Laboratory Animals.

Experimental design

Adult male and female Wistar rats were used for this study. Rats were mated overnight (2 females: 1 male). Copulation was confirmed by the presence of sperm in the vaginal smear the following morning (gestational day 0) (10-12). Twenty pregnant rats were randomly allocated into control and STZ-induced diabetic groups.

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Induction of DM

Mild hyperglycemia was induced in pregnant rats by intraperitoneal administration of STZ (Sigma, USA), with a dose of 40 mg/kg body weight, freshly dissolved in citrate buffer (pH 4.5), on the 5th day of gestation (13, 14). The rats in the control group received an equivalent volume of citrate buffer.

The diabetic state was verified by measuring the fasting blood glucose level (120–300 mg/dl) (15) using an Accu-Chek Active glucometer 72 hr after STZ injection. The normal fasting blood glucose level (<120 mg/dl) was also checked in rats in the control group.

Histological studies

Light microscopy

Six pregnant rats from each group were sacrificed using ketamine (90 mg/kg body weight) and xylazine (10 mg/kg body weight) anesthesia mixture on day 19 of pregnancy. Then the fetal livers were removed and fixed in 10% neutralbuffered formalin. The specimens were then dehydrated in an ascending series of ethanol, cleared with xylene, and embedded in paraffin. Four to five micrometer-thick sections were stained with periodic acid-Schiff (PAS), Masson trichrome, and reticulin. Hematoxylin and eosin (H&E) staining was used for quantitative morphometric analysis.

Sections were observed under an OLYMPUS BX51 microscope, and images were taken with an OLYMPUS DP12 digital camera.

Electron microscopy

For the electron microscopic study, samples were fixed in 2.5% glutaraldehyde (PBS-based EM grade) and processed as per the standard protocol (16). Sections were examined under a transmission electron microscope (TEM) (Leo 912 AB, Germany).

Morphometric study and statistical analysis

H&E-stained sections at 4 μm thickness were used for quantitative morphometric analysis. Measurements were done using five non-overlapping fields from 10–20 sections.

Parameters were examined

- *Using 100X oil-immersion objective a.* Area and diameter of hepatocyte b. Area and diameter of nucleus c. Diameter of central vein
- d. Diameter of bile duct

Using 40X objective: Density of megakaryocytes in 100000 μm2 of fetal liver

Morphometric parameters were measured using Olysia Bio software (Olympus Optical Co. LTD, Tokyo-Japan).

Statistics analysis

Results were presented as mean ± standard deviation (SD). The Shapiro-Wilk test was used to evaluate the normality of the data. Comparing means of variables between groups performed by the independent two-sample t-test. The correlation between variables was determined by Pearson's correlation coefficient test. All statistical analyses were carried out using Stata Version 16.0. A *P*-value<0.05 was considered statistically significant.

Results

Light microscopic and TEM observations of fetal liver

Examination of sections obtained from the livers of the control group revealed normal histological structure.

PAS staining was positive, revealing scattered eosinophilic inclusions in some hepatocytes in the GDM group (Figure 1, white arrows), while it was negative in the control group.

Masson trichrome staining was also positive and showed some fibrous tissue as fine fibers in the portal spaces that extended to the central vein (porto-central fibrosis) in the GDM group (Figure 2), while it was negative in the control group.

Reticulin staining was positive around the central vein and portal space vessels in the control group. In the GDM group, it was focally positive in the areas of fibrosis and the portal spaces (Figure 3).

In the ultrastructural study, pyknotic nuclei followed by karyolysis, as well as degranulation and vesiculation of the rough endoplasmic reticulum (RER), along with distortion and degeneration of mitochondria were found in the GDM

Figure 1. Photomicrograph of the rat fetal liver section in GDM group (PAS staining, \times 100, scale bar = 20 µm) GDM: Gestational diabetes mellitus

Figure 2. Photomicrographs of rat fetal liver sections in GDM group (Masson trichrome staining) GDM: Gestational diabetes mellitus

Figure 3. Photomicrographs of rat fetal liver sections (A) Control group, (B–D) GDM group (Reticulin staining) GDM: Gestational diabetes mellitus

Figure 4. Transmission electron micrographs of rat fetal liver sections in GDM group (A) Pyknotic nuclei (yellow arrow) and karyolysis (red arrow), (B) degranulation and vesiculation of the RER, (C) distortion and degeneration of mitochondria. GDM: Gestational diabetes mellitus

group compared to the control group (Figure 4).

Morphometric results

Morphometric parameters of fetal liver in GDM and control groups are given in Table 1 and Figure 5. The mean hepatocyte area and diameter were lower in the GDM group in comparison with the control group. These differences were statistically significant (mean difference area 14.03 ± 5.65 µm2 and diameter 1.02 ± 0.39 µm; *P*<0.05). However, there was no significant difference between groups in the nuclear area and diameter (mean difference area 0.78 ± 1.17 μ m² and diameter 0.09 \pm 0.15 μ m; *P*>0.05). A significant

Table 1. Morphometric evaluation of rat fetal liver in GDM and control groups

*SD. Standard deviation; **. Significant level in *P*<0.05.

Hepatocyte diameter (long axis); Nucleus, central vein, and bile duct diameter (X, Y axes); Density of megakaryocytes: number of megakaryocytes in 100000 µm² GDM: Gestational diabetes mellitus

Figure 5. Photomicrographs of rat fetal liver sections

(A) Control group (H&E, \times 40, scale bar = 50 µm), (B) Control group (H&E, \times 100, scale bar = 20 µm), (C) GDM group (H&E, \times 100, scale bar = 20 µm). Morphometric parameters: area and diameter of hepatocyte and nucleus, diameter of central vein and bile duct, and the density of megakaryocytes. Hepatocyte (H), central vein (CV), bile duct (BD), megakaryocyte (M), blood sinusoid (S), and portal vein (PV).

**. Correlation is significant at the 0.01 level (2-tailed)

*. Correlation is significant at the 0.05 level (2-tailed)

GDM: Gestational diabetes mellitus

difference in the mean density of megakaryocytes between the groups was observed (mean difference between groups = -2.60 ± 0.54 No/100000 µm2 ; *P*<0.001). Furthermore, the mean diameter of the central vein in the GDM group was higher than that in the control group. This difference was statistically significant (mean difference between groups = -2.33 ± 0.67 µm; $P = 0.001$). No significant difference was found in bile duct diameter between the groups (mean difference between groups = $-0.29 \pm 0.33 \, \mu m$; *P*=0.382).

Furthermore, the correlation of hepatocyte area with the nucleus area and diameter in GDM and control groups is shown in Table 2. The parameters did not correlate in the control group, as shown by Pearson's correlation coefficient test, while there was a strong positive correlation between hepatocyte area and nucleus area $(r = 0.61, P = 0.002)$ in the GDM group.

Discussion

In the present study, numerous histological, ultrastructural, and morphometric alterations were detected in the fetal liver following uncontrolled mild GDM.

These alterations included pyknotic nuclei, karyolysis, degranulation and vesiculation of theRER, and degeneration of mitochondria in the GDM group. Moreover, some fibrous tissue, as fine fibers in the portal spaces that extended to the central vein, was observed in the GDM group.

Similar to our results, in a study by El-Sayyad *et al*., in which experimental DM was induced by two successive intraperitoneal injections of STZ (60 mg/kg b.w) on days 5 and 6 of gestation and pregnant rats with blood glucose levels > 350 mg/dl were included in the diabetic group, abnormal mitochondria and RER were detected in the liver of 19-day-old fetuses of diabetic dams. Also, moderate fibrotic change was found in the liver of diabetic dams (17).

In addition, increased fibrous tissue was observed after six weeks in the liver of diabetic male rats (60 mg/kg b.w., intraperitoneal injection of STZ) compared to control in the study of Alshathly *et al*. (18).

Despite the morphometric studies of the liver of adult diabetic rats, the quantitative aspects of fetal liver structure have been less investigated in this regard. In our study, the mean hepatocyte area and diameter were significantly lower in the GDM group in comparison with the control group. Moreover, the mean diameter of the central vein was significantly higher in the GDM group than in the control group.

Previous experimental studies have revealed morphometric changes in the liver of diabetic rats (19- 22). The results of the study by Salahshoor *et al*. (19) demonstrated a significant increase in the size of hepatocytes and central vein in diabetic male Wistar rats (60 mg/kg b.w., intraperitoneal injection of STZ) compared to the normal control group.

In another study, the mean area of hepatocytes, nuclei, and nucleolus reduced in the periportal zone and increased in the perivenous zone in the diabetic male Wistar rats (STZinduced DM, 80 mg/kg b.w., intraperitoneal injection). The increase of hepatocyte area in the perivenous zone and the decrease of nucleus area in the periportal zone were significant in the diabetic group compared to the control group (20).

In another study (21) the morphometric parameters of the liver (Hepatocyte diameter and Central hepatic vein) in diabetic male Wistar rats (STZ-induced DM, 60 mg/kg b.w., single intraperitoneal injection) were increased.

In addition, DM (60 mg/kg b.w., intraperitoneal injection of STZ) caused an increase in the size of hepatocytes and their nuclei with a decrease in the nucleus-to-plasma ratio in rat liver in another study (22).These differences may be due to the type of DM (mild GDM) and the survey of the liver in the fetal stage in our study.

Furthermore, in our study, the density of megakaryocytes was significantly higher in the GDM group than in the control group. Researchers reported abundant lymphocytes and megakaryocytosis in the liver of fetuses of diabetic rats (STZ-induced DM before pregnancy, 35 mg/kg b.w., intraperitoneal injection) on gestational day 20 (23).

Liver injury in GDM animal models can occur due to hyperglycemic conditions, which cause hyperinsulinemia and insulin resistance. Hyperglycemic conditions cause inflammatory conditions and oxidative stress and thus worsen the liver injury process by triggering NF-B activation, which stimulates the pro-apoptotic genes activity in liver cells and enhances the production of reactive oxygen species (24, 25).

Limitation

There was no possibility of using ultrasound to determine the time of gestation in rats.

Conclusion

This study highlighted the histological, ultrastructural, and morphometric alterations in the fetal liver due to uncontrolled mild GDM.

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Authors' Contributions

MJ G, M G, and S AK conceived and designed the study; S AK processed and collected data, and performed experiments; S G contributed to the data collection; A M analyzed and interpreted the results; A R performed the statistical analysis. All authors contributed to writing the manuscript and approved the final version.

Conflicts of Interest

All authors declare that they have no conflicts of interest.

References

1. World Health Organization (WHO). The top 10 causes of death. https://www.who.int/news-room/fact-sheets/detail/the-top-10 causes-of-death.

2. Carr DB, Gabbe S. Gestational diabetes: Detection, management, and implications. Clinical Diabetes 1998; 16: 4-12.

3. Group HSCR, Metzger BE, Lowe LP, Dyer AR, Trimble ER, Chaovarindr U, *et al*. Hyperglycemia and adverse pregnancy outcomes. N Engl J Med 2008; 358: 1991-2002.

4. Perkins JM, Dunn JP, Jagasia SM. Perspectives in gestational diabetes mellitus: A review of screening, diagnosis, and treatment. Clinical Diabetes 2007; 25: 57-62.

5. Pedersen J. Weight and length at birth of infants of diabetic mothers. Eur J Endocrinol 1954; 16: 330-342.

6. Langer O. Fetal macrosomia: Etiologic factors. Clin Obstet Gynecol 2000; 43: 283-297.

7. Larciprete G, Valensise H, Vasapollo B, Novelli GP, Parretti E, Altomare F, *et al.* Fetal subcutaneous tissue thickness (SCTT) in healthy and gestational diabetic pregnancies. Ultrasound Obstet Gynecol 2003; 22: 591-597.

8. Khoury JC, Dolan LM, VanDyke R, Rosenn B, Feghali M, Miodovnik M. Fetal development in women with diabetes: imprinting for a life-time? J Matern Fetal Neonatal Med 2012; 25: 11-14.

9. Gojnic M, Stefanovic T, Perovic M, Arsic B, Garalejic E, Micic J, *et al*. Prediction of fetal macrosomia with ultrasound parameters and maternal glycemic controls in gestational diabetes mellitus. Clin Exp Obstet Gynecol 2012; 39: 512-515.

10. Canavan JP, Holt J, Goldspink DF. Growth of the rat foetal liver in gestational diabetes. Comp Biochem Physiol A Physiol 1991; 99: 473-476.

11. Nazari Z, Nabiuni M, Saeidi M, Golalipour MJ. Gestational diabetes leads to down-regulation of CDK4-pRB-E2F1 pathway genes in pancreatic islets of rat offspring. Iran J Basic Med Sci 2017; 20: 150-154.

12. Cederberg J, Picard JJ, Eriksson UJ. Maternal diabetes in the rat impairs the formation of neural-crest derived cranial nerve ganglia in the offspring. Diabetologia 2003; 46: 1245-1251.

13. Merzouk H, Madani S, Chabane Sari D, Prost J, Bouchenak M, Belleville J. Time course of changes in serum glucose, insulin, lipids and tissue lipase activities in macrosomic offspring of rats with streptozotocin-induced diabetes. Clin Sci (Lond) 2000; 98: 21-30.

14. Damasceno DC, Sinzato YK, Bueno A, Netto AO, Dallaqua B, Gallego FQ, *et al*. Mild diabetes models and their maternal-fetal repercussions. J Diabetes Res 2013; 2013: 473575.

15. Saito FH, Damasceno DC, Kempinas WG, Morceli G, Sinzato YK, Taylor KN, *et al*. Repercussions of mild diabetes on pregnancy in Wistar rats and on the fetal development. Diabetol Metab Syndr 2010; 2:26.

16. Bozzala JJ, Russel LD. Electron Microscopy Principles Techniques for Biologists, Jones and Barlett Publishers, Sudbury, Massachusetts. 2 nd Edition. 1998, 19-45, 72-144.

17. El-Sayyad HIH, Al-Haggar MMS, El-Ghawet HA, Bakr IHM. Effect of maternal diabetes and hypercholesterolemia on fetal liver of albino Wistar rats. Nutrition 2014; 30: 326-336.

18. Alshathly MR. Efficacy of Ginger (Zingiber officinale) in ameliorating streptozotocin-induced diabetic liver injury in rats: Histological and biochemical studies. J Microsc Ultrastruct 2019; $7: 91-101$.

19. Salahshoor MR, Mohammadi MM, Roshankhah S, Najari N, Jalili C. Effect of *Falcaria vulgaris* on oxidative damage of liver in diabetic rats. J Diabetes Metab Disord 2019;18:15-23.

20. Golalipour MJ, Ghafari S, Farsi MM. Effect of *Urtica dioica* L. Extract on quantitative morphometric alterations of liver parenchymal cells in STZ diabetic rats. Int J Morphol 2009; 27: 1339-1344.

21. Roshankhah S, Shabanizadeh A, Abdolmaleki A, Gholami MR, Salahshoor MR. Evaluation of biomarkers in liver following *Solanum melongena* green calyx administration in diabetic rats. J Diabetes Metab Disord 2020; 19: 1115-1127.

22. Welt K, Weiss J, Martin R, Dettmer D, Hermsdorf T, Asayama K, *et al.* Ultrastructural, immunohistochemical and biochemical investigations of the rat liver exposed to experimental diabetes und acute hypoxia with and without application of Ginkgo extract. Exp Toxicol Pathol 2004; 55: 331-345.

23. Mohamed S, Salem N, Hassan H, Abdelrahman G. Role of vitamin E in modulating the effect of Streptozotocin-induced maternal diabetes on fetal liver development in albino rat. Medicine Updates 2021; 7: 25-47.

24. Mahata LE, Ali H, Murni AW. Effect of streptozotocin on liver histology damage in rats model of gestational diabetes mellitus. Int J Res Rev 2021; 8: 18-22.

25. Ugwu MN, Umar IA, Utu-Baku AB, Dasofunjo K, Ukpanukpong RU, Yakubu OE, *et al*. Anti-oxidant status and organ function in streptozotocin-induced diabetic rats treated with aqueous, methanolic and petroleum ether extracts of Ocimum basilicum leaf. J Appl Pharm Sci 2013; 3: S75- S79.