

Potential of polymeric particles as future vaccine delivery systems/adjuvants for parenteral and non-parenteral immunization against tuberculosis: A systematic review

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ABSTRACT

Objective(s): Production of effective tuberculosis (TB) vaccine is necessity. However, the development of new subunit vaccines is faced with concerns about their weak immunogenicity. To overcome such problems, polymers-based vaccine delivery systems have been proposed to be used via various routes. The purpose of this study was to determine the potential of polymeric particles as future vaccine delivery systems/adjuvants for parenteral and non-parenteral immunization against TB.

Materials and Methods: PubMed, Scopus, Science-Direct, and the ISI web of knowledge databases were searched for related keywords. A total of 420 articles, written up to June 25, 2016, were collected on the potential of polymeric particles as TB vaccine delivery systems after parenteral and non-parenteral immunization. Thirty-one relevant articles were selected by applying inclusion and exclusion criteria.

Results: It was shown that the immunogenicity of TB vaccines had been improved by using biodegradable and non-biodegradable synthetic polymers as well as natural polymers and they are better able to enhance the humoral and cellular immune responses, compared to TB vaccines alone. The present study revealed that various polymeric particles, after *M. tuberculosis* challenge in animal models, provide long-lasting protection against TB. PLGA (poly (lactide-co-glycolide)) and chitosan polymers were widely used as TB vaccine delivery systems/adjuvants.

Conclusion: It seems that PLGA and chitosan polymers are well-suited particles for the parenteral and non-parenteral administration of TB vaccines, respectively. Non-biodegradable synthetic polymers in comparison with biodegradable synthetic and natural polymers have been used less frequently. Therefore, further study on this category of polymers is required.

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Introduction

Among main intracellular pathogens, *Mycobacterium tuberculosis* (*M. tuberculosis*) is known as the cause of a terrible infectious disease with a high mortality rate all over the world (1, 2). Tuberculosis (TB) vaccination is the most important approach to long-term control of the disease (3). The BCG vaccine, which is a live attenuated strain of *Mycobacterium bovis*, bacillus Calmette-Guérin, is the only licensed and cost-effective vaccine but with limited and variable effects on specific individuals in prevention of active disease (0-85%) (3, 4). Therefore, the development of new vaccines is an urgent need. Currently, at least 15 TB vaccine candidates are located in various phases of clinical trials, which are classified into three main groups: 1) prime group, includes vaccines that are designed as a replacement for the current BCG vaccine, 2) prime-boost group, includes vaccines that act as boosters for the BCG vaccine and 3)

immunotherapy group, includes therapeutic vaccines for effective treatment of various forms of TB disease as well as drug-resistant strains of *M. tuberculosis* (5). However, most of these vaccines are weak immunogens, and polymer-based particles as vaccine delivery systems/adjuvants could help to attain the following goals: 1) for optimal stimulation of innate and adaptive immune systems, 2) protection of antigens from *in vivo* enzymatic degradation, 3) effective antigen targeting to antigen-presenting cells (APCs) followed by its processing by both MHC-I and II routes and then induction of CD4⁺ and CD8⁺ T cells, 4) control of inducing cell-mediated responses (Th1 or Th2) by controlling the particle size, and 5) usage as an adjuvant (6-8). Among various biomaterials used in the development of protein, peptide, and DNA vaccines, polymers are of great importance due to some advantages including biocompatibility with synthetic polymers, lower toxicity,

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biocompatibility, easy availability, and biodegradability of natural polymers (8-12). Polymers could be categorized as 1) biodegradable synthetic polymers like PLGA (poly (lactide-co-glycolide)), PLA (poly (lactide)) (both are FDA approved) (8-11), poly (alkyl cyanoacrylates), poly (ϵ -caprolactone), and polyphosphates, 2) non-biodegradable synthetic polymers like polyvinylpyrrolidone, poloxamers, and poly (methyl methacrylates), and 3) natural polymers like proteins (albumin, collagen, and gelatin) and polysaccharides (chitosan/chitin and alginate) (13). In spite of the advantages of polymeric nanoparticles (NPs), they also have some disadvantages including toxicity, non-degradability, complexity, and costly synthesis process for synthetic polymers, and more complicated structure, complexity, and expensive extraction process for natural polymers (12).

There is two main delivery routes for vaccine administration, non-parenteral (mucosal) vaccination via intranasal, oral, vaginal, and rectal routes, and parenteral via subcutaneous and intramuscular (14). To date, parenteral routes are the common delivery routes for TB vaccine candidates, and there is much less research on mucosal vaccination (14). Since the respiratory mucosa is the site of TB entry, mucosal vaccination, especially the nasal route could provide good protection against TB (14). This could be performed by the production of neutralizing antibodies like secretory IgA (sIgA), systemic IgG, and also activation of various CD4⁺ T cells such as Th1, Th2, and Th17, and also CD8⁺ T cells (CTLs). These could strongly stimulate both mucosal and systemic immune responses (15, 16). Th1, Th17, and CTLs mediated immunity are essential for the host defense against intracellular pathogens that enter through various mucosal surfaces, whereas CD4⁺ Th2 cells are effective against extracellular organisms (16-18). Antibody responses via organism opsonization and consequently the more efficient processing by dendritic cells (DCs) have synergistic effects on the immune system (19). Additionally, the nasal cavity possesses other advantages such as patient compliance, better epithelium permeability, and needle-free and self-administration (14, 20). In the present review, the potential of various polymeric particles as future vaccine delivery systems/adjuvants for parenteral and mucosal immunization against TB has been investigated.

Materials and Methods

Literature search

PubMed, Scopus, Science-Direct, and the ISI web of knowledge databases were searched to identify relevant studies up to June 25, 2016. Data from published English language articles were obtained by using the medical terms including “*M. tuberculosis* vaccine” or “TB vaccine” and “polymeric NPs” or “polymeric microparticles” and

“PLGA” and “PLA” and “poly (alkyl cyanoacrylates)” and “poly (ϵ -caprolactone)” and “polyphosphates” particles and “polyvinylpyrrolidone” and “poloxamers” and “poly (methyl methacrylates)” particles and “protein (albumin, collagen and gelatin)” particles and “polysaccharide (chitosan/chitin and alginate)” particles and “immunization” or “vaccine delivery”. Finally, a supplementary search in the Google Scholar search engine and hand searching of reference lists to find the excluded articles was conducted.

Evaluation criteria

In this systematic review, the main inclusion criteria for selection of relevant articles included: 1) published original articles written in English, 2) evaluation of various TB vaccines only based on different polymeric particles, 3) parenteral and mucosal administration of TB vaccines, and 4) assessment of different polymeric particles as adjuvants. Exclusion criteria included: 1) abstracts of articles, 2) review articles and books, 3) use of other particles as vaccine delivery systems/adjuvants, 4) assessment of vaccine delivery in other organisms, 5) duplicate studies, and 6) assessment of drug delivery in *M. tuberculosis*.

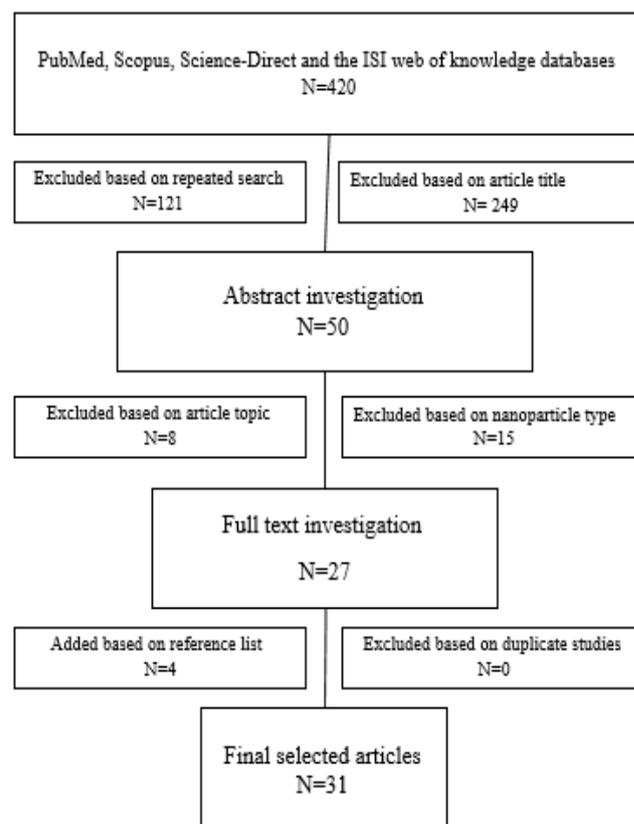


Figure 1: Flowchart of literature search and inclusion and exclusion criteria

Table 1. Polymeric particles as tuberculosis vaccine delivery systems

First author (Ref)	Year	Polymer	Antigen (s)	Adjuvant (s)	Preparation method	Size	Zeta-Potential (mV)	Route of administration	Animal	Profile of immune responses		
										Antibody	Cytokine	Challenge
Rose et al (21)	2015	PLGA (75:25)	MOMP	DDA:TDB (CAF01)	O/W	<250 nm	>5	Subcutaneous	Female B6C3F1 mice	IgG1 IgG2a IgA	IFN- γ IL-17a IL-5	NA
Carletti et al (22)	2013	PLGA (50:50)	Apa	TDM	W/O/W	NA	NA	Subcutaneous Intramuscular	Female BALB/c mice	NA	NA	<i>M. tuberculosis</i> H37Rv
Shi et al (23)	2010	PLGA (75:25)	TB10.4- Ag85B	MDP-BSA	Emulsion /spray-drying	3.0 μ m	-25	Pulmonary	NA	NA	IL-2	NA
Bivas-Benitaa et al (24)	2009	PLGA (53:47)	Rv1733c	PEI	W/O/W	235 - 275 nm	+38.8 - +64.3	Intramuscular Intranasal	Female BALB/c mice	NA	IFN- γ IL-12 TNF- α	NA
Kirby et al (25)	2008	PLGA (75:25)	Ag85B-ESAT-6	DDA:TDB (CAF01) DDA TDB	W/O/W	1.50 \pm 0.13 μ m	0-15	Subcutaneous	Female C57BL/6j mice	IgG1 IgG2b	IFN- γ	NA
de Paula et al (26)	2007	PLGA (50:50)	HSP65	TDM	W/O/W	<10 μ m	NA	Intratracheal	Female Hartley guinea pigs and Female BALB/c mice	IgG2a	IFN- γ	<i>M. tuberculosis</i> H37Rv
Lu et al (27)	2007	PLGA (75:25)	rAg85B	MDP TDB	Emulsion /spray-drying	3.4-4.3 μ m	NA	NA	NA	NA	IL-2	NA
Ha et al (28)	2006	PLGA (NA)	rAg85A ESAT-6	IL-12EM AS01B alum	W/O/W	NA	NA	Subcutaneous	Female C57BL/6 mice	Total IgG IgG1 IgG2a	IFN- γ	<i>M. tuberculosis</i> H37Rv
Cai et al (29)	2005	PLGA (50:50)	Ag85B MPT-64 MPT-83	DDA	W/O/W	<5 μ m	NA	Intramuscular	Female C57BL/6 mice	IgG	IFN- γ	<i>M. tuberculosis</i> H37Rv
Evans et al (30)	2004	PLGA (50:50)	Mtb8.4	MPL RC-529	Hydrophobic ion-pairing technique	2 μ m	NA	Intramuscular Subcutaneous	Female C57BL/6 mice	IgG IgG2a	IFN- γ	NA
Lima et al (31)	2003	PLGA (50:50)	HSP65	TDM	W/O/W	<5 μ m	NA	Intramuscular	Female BALB/c mice	IgG1 IgG2a	IFN- γ IL-10 IL-4	<i>M. tuberculosis</i> H37Rv
Lima et al (32)	2001	PLGA (50:50)	NA	TDM	W/O/W	NA	NA	Intraperitoneal Intratracheal	Female BALB/c mice	NA	IL-6 TNF- α IL-10 IFN- γ IL-12 IL-4	<i>M. tuberculosis</i> H37Rv
Dhiman et al (33)	1998	PLGA (50:50)	71-kDa cell wall protein	FIA	W/O/W	<0.65 μ m	NA	Intramuscular Subcutaneous	NA	NA	NA	<i>M. tuberculosis</i> H37Rv
Carpenter et al (34)	2005	PLA	ESAT-6	Alum	W/O/W	1.179 μ m \pm 0.07	NA	Pulmonary Intranasal Intramuscular	Female BALB/c mice	IgG	IFN- γ IL-4	NA
Venkataprasad et al (35)	1999	PLA PLGA (75:25, 50:50)	38 kDa protein	NA	W/O/W	PLA (3-5 μ m) and PLG (0.3-0.8 μ m)	NA	Subcutaneous	Female C57BL/10 mice	NA	IFN- γ IL-4	NA
Todoroff et al (36)	2013	Poloxamer 407	Ag85A	CpG oligonucleotide	NA	27 nm	NA	Pulmonary (Intratracheal)	Female BALB/c mice	IgG, IgG1, and IgG2a	IFN- γ TNF- α IL-2 IL-17a	NA

Continued Table 1.

Orr et al (37)	2014	NA	ID93	GLA-SE	NA	< 120 nm	-13	Intramuscular	Female C57Bl/6 mice	IgG1 IgG2c	IFN-γ TNF-α IL-2	<i>M. tuberculosis</i> H37Rv
Yeboah et al (38)	2009	Albumin	Dead whole cells and whole cell lysate	NA	Spray-drying method	3.52±0.13 μm 6.61±0.70 μm	-13.05 ±19.18 -40.28 ±7.86	Oral	Rat	IgG IgA	NA	NA
Meerak et al (39)	2013	Chitosan	Ag85B	NA	Coacervation method	100–200 nm	NA	Subcutaneous Intranasal	Female BALB/c inbred	total IgG IgG2a	IFN-γ IL-2 IL-4	NA
Feng et al (40)	2013	Chitosan	Esat-6/3e-FL	NA	Ionic crosslink and coacervation	311.2±34 nm	30.6 mV	Intramuscular Intranasal	Female C57Bl/6 mice	NA	IFN-γ IL-12 IL-10 IL-4	<i>M. tuberculosis</i> H37Rv
Ai et al (41)	2013	Chitosan	pHSP65pep	NA	Coacervation method	350–400 nm	NA	Intranasal Intradermal	Female BALB/c mice	IgA IgG IgG1 IgG2a	IFN-γ	<i>M. bovis</i>
Verma et al (42)	2013	Chitosan-TPP	CFP-10 CFP-21		Ionotropic gelation	250–300 nm	41 ±5.29	NA	NA	NA	IFN-γ IL-4	NA
Caetano et al (43)	2013	Chitosan/Alginate/TPP	BCG	NA	Ionotropic gelation	33.865 ±5.347 μm	+10.91 ±3.55	Intranasal Subcutaneous	Female BALB/c mice	IgG IgG1 IgG2a IgA	NA	NA
Zhu et al (44)	2007	Chitosan	AMM	IFA	Precipitation/coacervation method	5.78 ±0.65 μm	32.77 ±1.51	Subcutaneous	Female C57Bl/6 mice	IgG1 IgG2a	IFN-γ IL-4	NA
Bivas-Benita et al (45)	2004	Chitosan	DNA vaccine encoding eight HLA-A*0201-restricted T-cell epitopes	NA	Complexation-coacervation method	376±59 nm	21±4	Pulmonary Intramuscular	Female HLA-A2 transgenic mice	NA	IFN-γ	NA
Dobakhti et al (46)	2009	NA	BCG	Sodium alginate	NA	NA	NA	Subcutaneous	Female BALB/c mice	Total IgG IgG2a IgG1	IFN-γ	<i>M. bovis</i>
Ajdary et al (47)	2007	Sodium alginate	BCG	NA	NA	11.5 μm	NA	Subcutaneous Oral	Female BALB/c mice	Total IgG IgG2a IgG1	IFN-γ IL-4	<i>M. bovis</i>
Dobakhti et al (48)	2006	Calcium alginate	BCG	NA	Internal emulsification method	11 μm	NA	Subcutaneous Oral	Female BALB/c mice	Total IgG IgG2a IgG1	NA	NA
Wilkinson et al (49)	2000	Polystyrene	Ag85A, B and C	NA	NA	2 μm	NA	NA	NA	NA	IFN-γ	NA
Yu et al (50)	2012	Fe ₃ O ₄ -Glu-polyethyleneimine	Ag85A-ESAT-6-IL-21	NA	NA	NA	+36 mV	Intramuscular Subcutaneous	Male C57BL/6 mice	NA	IFN-γ	<i>M. tuberculosis</i> H37Rv
Ballester et al (51)	2011	Pluronic-stabilized polypropylene sulfide	Ag85B	CpG	Emulsion polymerization and surface functionalized	30 nm	NA	Intradermal Pulmonary	Female C57BL/6 mice	NA	IFN-γ TNF-α IL-6 IL-1β IL-17a IL-2	<i>M. tuberculosis</i> Erdman strain

Abbreviations: O/W: Oil-in-water single emulsion, W/O/W: Double emulsion/solvent evaporation, MOMP: Major outer-membrane protein, Apa: Alanine-proline antigen, MDP: Muramyl dipeptide, BSA: Bovine serum albumin, AS01B: Composed of MPL and QS21 (saponin molecule), RC-529 adjuvant: A synthetic ω-amin-oalkyl-2-amino-2-deoxy-4-phosphono-β-d-glucopyranoside (AGP) which is structurally related to the major hexaacyl component of MPL adjuvant, FIA: Freund's incomplete adjuvant, ID93: Fusion protein containing *M. tuberculosis* genes Rv3619, Rv1813, Rv3620, and Rv2608, GLA-SE: A mixture of squalene, DMPC (1,2-dimyristoyl-sn-glycero-3-phosphocholine), poloxamer 188, glycerol, and ammonium phosphate buffer, Esat-6/3e-FL: Esat-6 three T cell epitopes (Esat-6/3e) and fms-like tyrosine kinase 3 ligand (FL) genes, pHSP65pep: pECANS plasmid with four epitopes cast in the gene backbone of HSP65, namely ESAT-6₆₄₋₇₆, Ag85A₁₂₄₋₁₃₅, CFP-10₅₅₋₆₉, and Ag85B₁₄₁₋₁₅₃, TPP: Tripolyphosphate, AMM: Ag85B-MPT64₁₉₀₋₁₉₈-Mtb8.4, Fe₃O₄-Glu-PEI: Fe₃O₄-Glutamic acid-Polyethyleneimine, NA: Not available

Results

A total of 420 original articles were collected from different studies using systematic search in databases (Figure 1). Searching was limited to polymeric particles and their roles as vaccine delivery systems/adjuvants for parenteral and mucosal immunization against TB written up to June 25, 2016. All obtained studies in the present paper were reviewed to extract the following data: 1) year of study, 2) type of polymer and adjuvant, 3) antigen(s), 4) size, zeta-potential, and preparation method of particles, 5) route of administration, 6) animal, and 7) profile of immune responses. Thirty-one selected articles were classified into three groups based on the inclusion and exclusion criteria.

Biodegradable synthetic polymers

Among biodegradable synthetic polymers (PLGA, PLA, poly (alkyl cyanoacrylates), poly (ϵ -caprolactone), and polyphosphates), PLGA polymers are the most widely used as TB vaccine delivery systems/adjuvants. This systematic review includes 14 studies on PLGA polymers. It is evident that PLGA (50:50) is the most frequently used polymer. All studies have used the double emulsion/solvent evaporation method (W/O/W) to prepare PLGA polymers. After administration through various routes, PLGA polymers induced high levels of antibody and Th1, Th2, and Th17 immune responses. The impact of different adjuvants such as cationic lipid, DDA (dimethyldioctadecylammonium bromide), TDB (trehalose 6, 6'-dibehenate), PEI (polyethyleneimine), TDM (trehalose dimycolate), and lipid adjuvants, such as MPL (monophosphoryl lipid A), either separately or in combination, on the particle size, zeta-potential, and profile of immune responses were reviewed (Table 1).

So far, just two studies have investigated the application of PLA polymers as vaccine delivery systems for TB subunit vaccines. Data shows that the microencapsulated antigen was highly immunogenic and nasal delivery of the microencapsulated TB antigen generated specific cellular immune responses (Table 1).

However, no study on the application of other biodegradable synthetic polymers, poly (alkyl cyanoacrylates), poly (ϵ -caprolactone), or polyphosphates, as TB vaccine delivery systems/adjuvants was found for the present systematic review.

Non-biodegradable synthetic polymers

Our study shows the higher capacity of poloxamer polymers amongst other non-biodegradable synthetic polymers, like polyvinylpyrrolidone and poly (methyl methacrylates), in the induction of protective immunity against *M. tuberculosis*. It has been shown that after pulmonary delivery of the TB antigen with poloxamer polymers, as single adjuvants or combined

with other adjuvants; these polymers have good potentials in stimulation of humoral and cell-mediated immune responses. However, no study was found on polyvinylpyrrolidone or poly (methyl methacrylates) to better compare the non-biodegradable synthetic polymers as adjuvants, nano-, or micro-carriers for TB vaccines.

Natural polymers

Studies on albumin, collagen, and gelatin as natural polymers were not found except one on the albumin polymer. It has been reported that albumin polymers can induce antigen-specific mucosal and systemic immune responses. To date, there are no studies on collagen and gelatin as TB vaccine carriers. Ten studies on chitosan/chitin and alginate (natural polysaccharide polymers) were found. Chitosan polymers are widely used as TB vaccine delivery systems/adjuvants. It was shown that chitosan can induce increased levels of antibody and IFN- γ secretion against TB antigens.

Discussion

Main prevention strategies against TB are prophylactic and post-exposure vaccination. Prophylactic vaccines may be used either for replacing BCG, like live mycobacterial vaccines, or as a booster for BCG, like subunit vaccines. Post-exposure vaccines are used for elimination of the disease in latently-infected individuals and to prevent disease reactivation (52). Ag85A, Ag85B, Ag85C, TB10.4, MPT-64, MPT-83, ESAT-6, CFP-10, Mtb8.4, and hsp65 are among main TB antigens that are used as DNA or subunit vaccines, either alone or as a fusion protein. These antigens could induce specific antibodies and cell-mediated immune responses (Table 1). These are mostly expressed in the early phase of infection by the replicating bacteria, however, to induce a broad spectrum of immune responses, antigens expressed in latency phase of non-replicating bacteria are also important, because they are often hidden from the immune system (53). This review suggests that ideal vaccination approach against both acute and latent TB infections is using both early and late stage antigens of *M. tuberculosis* (multistage booster vaccines).

TB DNA and subunit vaccines have high safety profiles and have shown promising results in animal models, however, in human clinical trials, they were poor immunogens (8, 24). To overcome this problem, polymer-based particles have been used as adjuvant/delivery systems to potentiate the immune responses. Among them, PLGA particles were mostly used for delivery of prophylactic TB vaccines (Table 1). The PLGA (50:50) copolymer as compared with the various polymer compositions (like 75:25 and 53:47), was the most frequently used PLGA polymer. The PLGA (75:25) as an intermediate copolymer has

more efficient protein release (due to faster degradation) than the PLA homopolymer and more prolonged release profile in comparison with the PLGA (50:50) copolymer (25). As presented in Table 1, using cationic lipids as adjuvants in combination with PLGA polymers could affect the size, zeta-potential, polydispersity index (PDI), entrapment efficiency, *in vitro* release profile, and also elicit simultaneously humoral and cell-mediated immune responses (8, 25). The cationic lipids skew the immune response towards the Th1 branch of immunity and IgG2a production, which is important for immunization against TB. In the presence of cationic lipids, reduction in particle size and PDI, change of zeta potential from negative to positive, and the decrease in antigen entrapment were observed. Similar results have been shown by Wedlock *et al* and Jensen *et al* (54, 55). As shown in Table 1, encouraging results have been observed after administration of PLGA-based TB vaccines via various routes. Several studies have shown that the PLGA polymer is well-tolerated in the most common injection sites, i.e., subcutaneous and intramuscular spaces (56). Due to biocompatibility, adjuvanticity, and prolonged release profile of PLGA particles, mucosal (nasal, oral, and pulmonary) administration of PLGA-based vaccines have shown promising results (25).

PLA particles can be built with the same method as PLGA particles and induced robust Th1-type responses (IFN- γ) against TB (Table 1). However, the main disadvantages of PLA polymers are the possibility of denaturation of encapsulated protein and also change in immunogenicity of loaded antigens during particle formulation and in the release period (35). To preserve the antigens' integrity, antigens could be adsorbed onto the PLA or PLGA particles.

Other biodegradable synthetic polymers, such as poly (ϵ -caprolactone), are also suitable for preparation of vaccine delivery systems (56). However, these polymers have not been tried for delivery of TB vaccines.

Non-biodegradable synthetic polymers compared to biodegradable synthetic and natural polymers have been less frequently used as carrier/adjuvant for TB vaccines. However, as discussed in the Todoroff *et al* study, Poloxamer 407 (a non-biodegradable synthetic polymer) combined with a CpG oligonucleotide, induced immune responses against the *M. tuberculosis* antigen (Ag85A) (Table 1) (36).

It is suggested that natural polymers due to milder preparation process, lower prices, and their adjuvant potential are attractive materials for replacement with biodegradable synthetic polymers (56). The protective and immunogenic efficacy of the vaccines based on the natural polymers has been shown in many studies. Yeboah *et al* and Dobakhti *et al* have shown that oral delivery of albumin and alginate microspheres loaded with TB vaccines could induce

mucosal and systemic immune responses (38, 46). As Table 1 represents, alginate-based TB vaccines have been administered subcutaneously and orally. According to these studies, alginates may be appropriate for utilization in oral vaccination and will protect encapsulated BCG against degradation in the stomach and induce strong Th1 immune responses against TB (46-48, 56). Several studies have indicated that chitosan-based vaccines can significantly promote Th1 and CTL immune responses and prevent TB infection (39-45). According to the different studies shown in Table 1, due to antigen depot, efficient uptake by membrane epithelium, and mucoadhesive and adjuvant properties of these biodegradable particulate delivery systems, they are ideal intranasal vaccine delivery systems/adjuvants against TB. As mentioned in Table 1, polymer-based TB vaccines have shown promising results in pre-clinical studies and were able to protect animal models against challenge with the virulent *M. tuberculosis* H37Rv strain, however, none of these vaccines entered clinical trials. Due to the results obtained in this study, we are optimistic about introduction of polymer-based TB vaccines to the clinical trial pipeline as potential antigen delivery systems/adjuvants for parenteral and non-parenteral immunization.

Conclusion

In summary, the present systematic review reveals that to develop a new TB vaccine, more attention to antigens expressed in early and latency phase of *M. tuberculosis* infection is needed. On the other hand, for improving the weak immunogenicity of these antigens, co-delivery of TB vaccines with polymeric particles could be beneficial in reduction of obstacles against development of new and more efficient TB vaccines. The present study revealed that various polymeric particles could be used as carriers for parenteral and mucosal administration of TB vaccine candidates. Among the polymers reviewed, PLGA and chitosan polymers are the most outstanding ones that show promising results, after subcutaneous and mucosal administration, respectively. Non-biodegradable synthetic polymers in comparison with biodegradable synthetic and natural polymers have been used less frequently. Therefore, further study on this category of polymers is required.

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