Iranian Journal of Basic Medical Sciences

ijbms.mums.ac.ir

Cinnamaldehyde improves methamphetamine-induced spatial learning and memory deficits and restores ERK signaling in the rat prefrontal cortex

Mohammad Saeed ¹, Ameneh Ghadiri ², Farzin Hadizadeh ^{3, 4}, Armin Attaranzadeh ⁵, Mohaddeseh Sadat Alavi ⁶, Leila Etemad ^{7*}

¹ School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

² Department of Internal Medicine and Medical Specialties, Sapienza University of Rome, Rome, Italy

³ Biotechnology Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran

⁴ Department of Medicinal Chemistry, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

⁵ Milad Infertility Center, Imam Reza Hospital, Mashhad University of Medical Sciences, Mashhad, Iran

⁶ Division of Neurocognitive Sciences, Psychiatry and Behavioral Sciences Research Center, Mashhad University of Medical Sciences, Mashhad, Iran ⁷ Pharmaceutical Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran

ARTICLE INFO ABSTRACT Article type: **Objective(s)**: Methamphetamine is a stimulant compound that penetrates readily into the central Original article nervous system. Repeated exposure to methamphetamine leads to damage in the dopaminergic and serotonergic axons of selected brain regions. Previous studies showed that cinnamaldehyde *Article history:* Received: Sep 2, 2018 improved memory impairment in animals. In the present study, we aimed to elucidate the effects of cinnamaldehyde on methamphetamine-induced memory impairment in rats. Accepted: Oct 18, 2018 Materials and Methods: Male Wistar rats received methamphetamine (10 mg/kg, intraperitoneally) Keywords: for 7 days. Thirty minutes before each injection, animals were given cinnamaldehyde (20, 40, or 80 Cinnamaldehyde mg/kg) or rivastigmine (1 mg/kg). The spatial learning and memory were examined using the Morris ERK1/2 water maze test. The expression of extracellular signal-regulated kinase (ERK) phosphorylation in the Learning deficit frontal cortex and hippocampus was also detected by immunohistochemical method. Memory deficit Results: Administration of methamphetamine increased the latency to find the platform in the learning Methamphetamine phase, while administration of cinnamaldehyde (40 mg/kg) or rivastigmine before methamphetamine reversed the increased latency. Administration of cinnamaldehyde, at the dose of 40 mg/kg with methamphetamine, increased the time and distance traveled in the target quadrant in comparison with the amphetamine group. Moreover, the methamphetamine and cinnamaldehyde-treated group had higher expression of phosphorylated ERK1/2 in the prefrontal cortex in comparison with the methamphetamine-treated animals. Conclusion: The present data demonstrated that repeated METH administration impaired cognitive performance through the ERK pathway and decreased the phosphorylation of ERK1/2 in the prefrontal cortex while administration of cinnamaldehyde restored both effects. Accordingly, cinnamaldehyde may be a valuable therapeutic tool for the treatment of cognitive deficits associated with methamphetamine consumption.

Please cite this article as:

Saeed M, Ghadiri A, Hadizadeh F, Attaranzadeh A, Alavi MS, Etemad L. Cinnamaldehyde improves methamphetamine-induced learning and spatial memory deficits and restores ERK signaling in the rat prefrontal cortex. Iran J Basic Med Sci 2018; 21:1316-1321. doi: 10.22038/IJBMS.2018.35368.8427

Introduction

Methamphetamine (METH) or crystal METH is a powerful stimulant drug that belongs to the amphetamine family. According to the published data, METH consumption is ranked as the second drug abused all over the world (1). Probably, low cost of synthesis and relative ease of access are the main reasons for the rise in METH abuse. METH has also been approved by the U.S. Food and Drug Administration (FDA) for treatment of obesity and the attention deficit hyperactivity disorder (ADHD) in children(2). Acute and chronic administration of METH produces a range of adverse effects including mental, behavioral, and cardiovascular disorders. METH can easily cross the blood-brain barrier and penetrate into the central nervous system (CNS) (3, 4). METH enhances the release of norepinephrine, serotonin, and dopamine from the cytosol into the synapses. In fact, it acts as an indirect agonist of monoamine receptors

(5). Acute and especially long-term exposure to METH leads to neurological deficits through chemical and molecular changes including excitotoxicity, oxidative stress, and apoptosis (6). Repeated exposure to METH impairs spatial learning and memory. METH exposure, even after many years of abstinence, produces deficits in selective attention, working memory, impulse control and information processing. It is well documented that extracellular signal-regulated kinase (ERK) pathway is involved in METH- induced behavioral and cognitive responses. ERK, a member of the mitogen-activated protein kinase (MAPK) superfamily, is expressed widely in the PFC, hippocampus and striatum. The activated ERK, phosphorylated ERK (pERK), activates downstream of cAMP-response element-binding protein (CREB) and exerts broad effects on biological and behavioral responses (7, 8).

IJ MS

Cinnamomum cassia (C. cassia) is a traditional medicinal

*Corresponding author: Leila Etemad. Pharmaceutical Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran. Tel: +98-5137112611; Fax: +98-38823251; Email: Etemadl@mums.ac.ir

herb that has been used worldwide especially in Asia to treat diabetes, dyspepsia, anxiety, ischemia, cancers, and inflammatory diseases (9-12). Cinnamaldehyde, also known as cinnamic aldehyde (CA), is a natural product from cinnamon tree, which is a yellow viscous liquid (13). It is the main component of cinnamon oil produced from the stem bark of *C.cassia* (14). CA has a broad range of biological effects including anti-tumor (15), antioxidant , and anti-inflammatory activities (16). In addition, CA plays neuroprotective roles in Alzheimer's and Parkinson's diseases while its mechanism(s) of action is as yet unclear (17, 18).

Considering the potential neuroprotective properties of CA, this study was designed to elucidate the effects of CA on learning and spatial memory deficits induced by METH in rats and to explore the underlying mechanism.

Materials and Method

Animals and drugs

Seven groups of six male Wistar rats, weighing 200–250 g, were obtained from the Animal Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences. The animal house temperature was maintained at 23 ± 2 °C with a 12 hr light/dark cycle. Rats had free access to food and tap water. Experiments were approved by the Animal Care Committee of Mashhad University of Medical Sciences, Mashhad, Iran.

CA and rivastigmine were purchased from Sigma (Germany). METH was donated by the Department of Medicinal Chemistry, School of Pharmacy, Mashhad University of Medical Sciences (Mashhad, Iran). Cinnamaldehyde was dissolved in 1% Tween 80 plus sterile normal saline as vehicle (19). All other chemicals were dissolved in normal saline. The anti-phospho-P44/42 Map (Erk1/2) and rabbit polyclonal antibody were purchased from Cell Signaling Technology (#9101 and #7074).

Groups and treatments

Five groups of rats were treated by METH (10 mg/kg) for seven days. Thirty min before METH injection, they received vehicle (control group), CA (20, 40 and 80 mg/kg) or rivastigmine (RIV) (1 mg/kg) (positive control group). Controls received normal saline and one group received normal saline plus CA at the dose of 80 mg/kg. All injections were given intraperitoneally (IP). At the end of the seventh day, spatial learning and memory were examined using the Morris water maze (MWM) test. One day after completion of the MWM test, all animals were sacrificed and their brains were dissected and stored in formalin (Figure 1).

Morris water maze

The Morris water maze consisted of a black circular tank (160 cm in diameter and 60 cm high) filled with water (depth 25 cm, 20–22 °C) and a square platform (10 cm diameter) was submerged 2 cm below the surface of the water in the center of the west quadrant. The pool was divided into four equal size quadrants, as four different starting points, labeled North (N), South (S), East (E), and West (W). The MWM test was performed in three distinct phases. In brief, at the pre-adaptation



Figure 1. Diagrammatic sketch of the behavioral and histopathological experiments. MWM: Morris water maze; SAC: sacrificed for histopathological experiments

phase, each rat was habituated to MWM without the platform for 30 sec. During the acquisition phase, the rats were allowed a maximal time of 60 sec for swimming to find the hidden platform (escape latency) with a resting time of 15 sec on the platform. For each animal, four trials were carried out daily for 5 consecutive days. In the probe test as the third phase, the platform was removed and the animals were allowed to swim for 60 sec. The swim speed, time spent in the target quadrant, and distance traveled in the target quadrant, where the platform was previously found, were recorded using a camera above the pool and analyzed using a software package (Radiab, Ver 2.1, Iran) (20).

Immunohistochemical evaluation

After completion of the MWM task, rats were sacrificed and the brains were removed and placed into 10% formalin and processed for paraffin embedding. The blocks were cut at 5 μ m thickness. For the histopathological examination, tissue sections were placed on slides and dried overnight at 45 °C, dewaxed in xylene, and rehydrated in graded alcohols. Endogenous peroxidase activity was blocked by incubation in 3% hydrogen peroxide (H₂O₂) and then incubated in a blocking solution. The primary antibody, phosphorylated ERK1/2, was placed on the slides at appropriate dilution. Sections were then incubated with rabbit anti-IgG conjugated to horseradish peroxides (Dako) and DAB substrate.

The phosphorylated ERK1/2 immunoreactivity level of each sample was evaluated semi-quantitatively under a light microscope by evaluating the signal intensity (0, 1+, 2+, or 3+) and the percentage of cells showing positive nuclear staining $(0, \text{ none}; 0.1, \text{ less than one-tenth}; 0.5, \text{ less than one half}; and 1, greater than one half}). The intensity and proportion scores were then multiplied using a semi-quantitative scoring method to assign a histochemical score (H-score)(21).$

Statistical analysis

All data were expressed as mean±SEM. In the Morris water maze test, the swimming speed and time spent to find the platform during training sessions were analyzed using repeated measures ANOVA. The time spent, distance traveled, and swimming speed in the probe trial were analyzed using one-way ANOVA followed by Tukey's test. In order to compare the mean H-Score, Kruskal–Wallis test followed by unpaired Mann-Whitney post hoc test was performed. P< 0.05 was considered as the level of statistical significance.

Results

Effect of METH and CA on escape latency and swim speed during the learning phase

The results showed that during the acquisition phase,



Figure 2. Escape latency (A) and swim speed (B) in METH-treated rats (10 mg/kg) that received CV (20, 40, or 80 mg/kg) or RIV (1 mg/kg) in the training days using the Morris water maze test. METH: Methamphetamine, CA: Cinnamaldehyde, RIV: Rivastigmine. The results are reported as mean+SEM, ### P<0.001 compared with the control group; * P<0.05, ** P<0.01 compared with the METH group

the performance of all groups was improved as indicated by decreased escape latency. However, administration of METH increased latency to find the hidden platform during five days compared with the control group (P< 0.001). RIV and CA (40 mg/kg) when administrated before METH, decreased latency time, which was significantly different from the METH-treated group at days 4 and 5 (P< 0.01 and P< 0.05, respectively) (Figure 2A). There were no differences between the swimming speeds of any group during the learning phase (Figure 2B).

Effect of METH and CA on time spent, distance traveled, and swim speed in the probe trial

The results showed that METH-treated rats failed to remember the platform location such that the time spent and distance traveled in the target quadrant were decreased (P<0.05 and P<0.001, respectively). Treatment with CA (40 mg/kg) and RIV significantly increased the time spent in the target quadrant, as compared with the METH group (Figure 3A, P<0.05). In the same way, as shown in figure 3B, the distance traveled in the target quadrant was significantly increased following co-administration of CA (40 mg/kg) and RIV with METH compared with the METH group, implying improvement



Figure 3. Time spent (A), distance traveled (B), and swim speed (C) in METH-treated rats (10 mg/kg) that received CV (20, 40, or 80 mg/kg) or RIV (1 mg/kg) in the target quadrant using the Morris water maze test. METH: Methamphetamine, CA: Cinnamaldehyde, RIV: Rivastigmine. The results are reported as mean+SEM, # P < 0.05 and ### P < 0.001 compared with the control group; * P < 0.05, *** P < 0.001 compared with the METH group



Figure 4. Effects of methamphetamine, cinnamaldehyde, and rivastigmine on phospho ERK levels using H-Score. METH: Methamphetamine, CA: Cinnamaldehyde, RIV: Rivastigmine. ### *P* < 0.001 compared with the control group and ** *P* < 0.01 compared with the METH group

of memory recall. Furthermore, there were no significant differences between the swimming speeds of any group on the 15th day of the experiment (Figure 3C).

Effect of METH and CA on PFC and hippocampal phospho-ERK1/2 levels

The results of PFC immunohistochemistry staining in different treatment groups are shown in Figure 4. The METH group exhibited a lower phospho-ERK1/2 level than the control group (P<0.001). Co-administration of CA (40 mg/kg) and METH increased the phospho-ERK1/2 level compared with the METH group (Figure 4, P<0.01). The riv-treated group had also an increased phospho-ERK1/2 level compared with the METH





Figure 5. Phospho-ERK 1/2 expression in the prefrontal cortex of rats that received normal saline plus tween (A), METH (B), CA (80 mg/kg) (C), METH plus CA (20, 40, or 80 mg/kg) (D, E, and F, respectively) or METH plus RIV(G). Phospho-ERK 1/2-positive cells were decreased after METH exposure and reversed after RIV and CA (40 mg/kg) administrations. METH: Methamphetamine, CA: Cinnamaldehyde, RIV: Rivastigmine. Magnification = 400X

group (Figure 5, P<0.01). There were no changes in the phospho-ERK1/2 levels in the hippocampus of any group (data not shown).

Discussion

METH is a powerful mental stimulant that is widely abused. Scientists believe that long-term METH use leads to cognitive defect in humans. Clinically, consumption of this substance has been associated with various disorders such as verbal and nonverbal memory impairment, attention deficit disorder, and motor impairment (22, 23). Morris water maze has been introduced as a valid animal model to investigate spatial learning and memory in rodents (24). The results of this study showed that CA at the dose of 40 mg/kg and RIV when co-administrated with METH significantly reversed the METH-induced memory impairment by increasing the time and distanced travel in the target quadrant in probe trial. To evaluate the influence of swimming ability on learning performance, mean speeds of all animals were measured and no significant difference was found between groups indicating that motor function and motivational and visual abilities were not affected during experiments. It was reported that pretreatment with Cinnamon extract (200 and 400 mg/kg, orally, 21 days) significantly reversed the scopolamine deteriorative effect on escape latency in the MWM test (25).

Cognitive impairment during chronic administration of METH has been attributed to changes in the ERK signaling pathways (8). ERK belongs to MAPKs family and plays a critical regulatory role in cellular functions such as differentiation, proliferation, development, as well as, memory and learning processing. Accumulating evidence indicates that ERK cascade is implicated in learning and formation of memory and behavioral responses to drugs of abuse such as METH and cocaine (8). The present data showed that repeated METH

administration impaired cognitive performance through the ERK pathway and decreased the phosphorylation of ERK1/2 in the PFC, while co-administration of CA and RIV restored it. However, ERK expression in the hippocampus was not affected by METH administration. In accordance with our study, it has been reported that repeated METH treatment increased escape latency in the training phase and decreased the number of crossings of the target quadrant in the probe test and modulated the MAPK-ERK signaling pathway by reduction of the ERK1/2 level in the PFC but not in the hippocampus and led to memory and learning abnormalities (7). The results of other studies also revealed that METH administration in single dose can induce apoptosis and gliosis in striatum and cortex regions and reduction in phospho ERK in PFC at repeated dose in mice (26, 27). The PFC has an essential role in short-term/working memory and decision-making, while the hippocampus is involved in the formation of long-term memories. The PFC mediates decision-making functions and is, therefore, a key neuroanatomical region in addictive behaviors (7).

Evidence suggests that cinnamon has a potential role in the prevention of neurodegenerative diseases (28). The protective effect of cinnamon and its active components against neuroinflammation was evaluated using lipopolysaccharide (LPS)-induced inflammation both in vitro and in vivo (28). In a mouse model of LPS-induced memory impairment and synaptic plasticity inhibition, CA pretreatment decreased nitric oxide production and interleukin-1β release in primary microglia. CA was also able to reduce the levels of phosphorylated ERK1/2 in the hippocampus of mice treated with LPS and interfered with the MEK1/2-ERK1/2 signaling pathway and destabilized inducible nitric oxide synthase mRNA expression. CA significantly diminished memory deficit and improved synaptic plasticity in LPS-treated mice (29). Cinnamon and its metabolite, sodium benzoate,

can activate the cAMP response element binding protein (CREB) via protein kinase A (PKA) and up-regulation of molecules associated with hippocampal plasticity. In agreement, oral administration of cinnamon and sodium benzoate improved spatial memory consolidation and transformed poor learning mice to good learners (30).

Based on our findings, METH treatment induced neuronal abnormalities in the PFC, but not in the hippocampus, and CA at medium dose significantly reversed it. The results of our study showed that CA at the dose of 80 mg/kg did not improve memory impairment or the expression level of phosphorylated ERK1/2. This finding is in accordance with the results of Iersel *et al.* They investigated the effect of CA on glutathione S-transferase activity in skin cancer cells. CA increased the level of GSH against the carbonyl derivatives toxicity in human melanoma cells. However, this effect was reversed at concentrations higher than 40 μ M (31). This finding implies that the protective effects of CA may be vanished at certain doses and this should be considered in future studies.

Conclusion

The study data showed that cinnamaldehyde had neuroprotective effects and improved cognition and learning function in METH-treated animals through activation of the ERK pathway in the PFC. Our findings also showed that the effects of CA were not dosedependent and this may limit the clinical applications of this natural compound.

Acknowledgment

The results shown here are from a doctoral thesis. This study was financially supported by the Research Council of Mashhad University of Medical Sciences and National Institute for Medical Research and Development (951155), Iran.

Conflicts of Interest

All authors declare no conflicts of interest.

References

1. Courtney KE, Ray LA. Methamphetamine: an update on epidemiology, pharmacology, clinical phenomenology, and treatment literature. Drug Alcohol Depend 2014;143:11-21.

2. Onakpoya IJ, Heneghan CJ, Aronson JK. Post-marketing withdrawal of anti-obesity medicinal products because of adverse drug reactions: a systematic review. BMC Medicine 2016;14:1-11.

3. Cruickshank CC, Dyer KR. A review of the clinical pharmacology of methamphetamine. Addiction 2009;104:1085-1099.

4. Ghadiri A, Etemad L, Moshiri M, Moallem SA, Jafarian AH, Hadizadeh F, *et al.* Exploring the effect of intravenous lipid emulsion in acute methamphetamine toxicity. Iran J Basic Med Sci 2017;20:138-144.

5. Halpin LE, Collins SA, Yamamoto BK. Neurotoxicity of methamphetamineand3,4-methylenedioxymethamphetamine. Life Sci 2014;97:37-44.

6. Yu S, Zhu L, Shen Q, Bai X, Di X. Recent advances in methamphetamine neurotoxicity mechanisms and its molecular pathophysiology. Behav Neurol 2015;2015:1-11.

7. Chen YJ, Liu YL, Zhong Q, Yu YF, Su HL, Toque HA, *et al.* Tetrahydropalmatine protects against methamphetamineinduced spatial learning and memory impairment in mice. Neurosci Bull 2012;28:222-232. 8. Sun WL, Quizon PM, Zhu J. Molecular mechanism: ERK signaling, drug addiction, and behavioral effects. Prog Mol Biol Transl Sci 2016;137:1-40.

9. Liao JC, Deng JS, Chiu CS, Hou WC, Huang SS, Shie PH, *et al.* Anti-inflammatory activities of cinnamomum cassia constituents in vitro and in vivo. Evid Based Complement Alternat Med 2012;2012:1-12.

10. Zaidi SF, Aziz M, Muhammad JS, Kadowaki M. Review: Diverse pharmacological properties of Cinnamomum cassia: A review. Pak J Pharm Sci 2015;28:1433-1438.

11. Medagama AB. The glycaemic outcomes of Cinnamon, a review of the experimental evidence and clinical trials. Nutr J 2015;14:1-12.

12. Sohrabi R, Pazgoohan N, Seresht HR, Amin B. Repeated systemic administration of the cinnamon essential oil possesses anti-anxiety and anti-depressant activities in mice. Iran J Basic Med Sci 2017;20:708-714.

13. Bickers D, Calow P, Greim H, Hanifin JM, Rogers AE, Saurat JH, *et al.* A toxicologic and dermatologic assessment of cinnamyl alcohol, cinnamaldehyde and cinnamic acid when used as fragrance ingredients. Food Chem Toxicol 2005;43:799-836.

14. Lv C, Yuan X, Zeng HW, Liu RH, Zhang WD. Protective effect of cinnamaldehyde against glutamate-induced oxidative stress and apoptosis in PC12 cells. Eur J Pharmacol 2017;815:487-494.

15. Herdwiani W, Someradji AA, Elfahmi., TAN MI. A review of cinnamon as a potent anticancer drug. Asian J Pharm Clin Res 2016;9:8-13.

16. Rao PV, Gan SH. Cinnamon: a multifaceted medicinal plant. Evid Based Complement Alternat Med 2014;2014:642942.

17. Bae WY, Choi JS, Jeong JW. The neuroprotective effects of cinnamic aldehyde in an MPTP mouse model of parkinson's disease. Int J Mol Sci 2018;19:1-10.

18. Momtaz S, Hassani S, Khan F, Ziaee M, Abdollahi M. Cinnamon, a promising prospect towards Alzheimer's disease. Pharmacol Res 2018;130:241-258.

19. Montrucchio DP, Cordova MM, Santos AR. Plant derived aporphinic alkaloid S-(+)-dicentrine induces antinociceptive effect in both acute and chronic inflammatory pain models: evidence for a role of TRPA1 channels. PLoS One 2013;8:e67730.

20. Moghimi M, Parvardeh S, Zanjani TM, Ghafghazi S. Protective effect of alpha-terpineol against impairment of hippocampal synaptic plasticity and spatial memory following transient cerebral ischemia in rats. Iran J Basic Med Sci 2016;19:960-969.

21. Handra-Luca A, Bilal H, Bertrand J-C, Fouret P. Extracellular signal-regulated ERK-1/ERK-2 pathway activation in human salivary gland mucoepidermoid carcinoma: association to aggressive tumor behavior and tumor cell proliferation. Am J Pathol 2003;163:957-967.

22. Gonzalez R, Rippeth JD, Carey CL, Heaton RK, Moore DJ, Schweinsburg BC, *et al.* Neurocognitive performance of methamphetamine users discordant for history of marijuana exposure. Drug Alcohol Depend 2004;76:181-190.

23. Thompson PM, Hayashi KM, Simon SL, Geaga JA, Hong MS, Sui Y, *et al.* Structural abnormalities in the brains of human subjects who use methamphetamine. J Neurosci 2004;24:6028-6036.

24. Vorhees CV, Williams MT. Assessing spatial learning and memory in rodents. ILAR J 2014;55:310-332.

25. Jain S, Sangma T, Shukla SK, Mediratta PK. Effect of Cinnamomum zeylanicum extract on scopolamine-induced cognitive impairment and oxidative stress in rats. Nutr Neurosci 2015;18:210-216.

26. González B, Raineri M, Cadet JL, García-Rill E, Urbano FJ, Bisagno V. Modafinil improves methamphetamine-induced object recognition deficits and restores prefrontal cortex ERK signaling in mice. Neuropharmacology 2014;87:188-197.

27. Moshiri M, Hosseiniyan SM, Moallem SA, Hadizadeh F,

Jafarian AH, Ghadiri A, *et al.* The effects of vitamin B12 on the brain damages caused by methamphetamine in mice. Iran J Basic Med Sci 2018;21:434-438.

28. Stavinoha RC, Vattem DA. Potential neuroprotective effects of cinnamon. ICMJE 2015;8:24-46.

29. Zhang L, Zhang Z, Fu Y, Yang P, Qin Z, Chen Y, *et al.* Trans-cinnamaldehyde improves memory impairment by blocking microglial activation through the destabilization of iNOS mRNA in mice challenged with lipopolysaccharide. Neuropharmacology 2016;110:503-518.

30. Modi KK, Rangasamy SB, Dasarathi S, Roy A, Pahan K. Cinnamon Converts Poor Learning Mice to Good Learners: Implications for Memory Improvement. J Neuroimmune Pharmacol 2016;11:693-707.

31. Iersel ML, Ploemen JP, Struik I, van Amersfoort C, Keyzer AE, Schefferlie JG, *et al.* Inhibition of glutathione S-transferase activity in human melanoma cells by alpha,beta-unsaturated carbonyl derivatives. Effects of acrolein, cinnamaldehyde, citral, crotonaldehyde, curcumin, ethacrynic acid, and trans-2-hexenal. Chem Biol Interact 1996;102:117-132.