

A study on the immune response induced by a DNA vaccine encoding Mtb32C-HBHA antigen of *Mycobacterium tuberculosis*

Roghayeh Teimourpour ¹, Hadi peeridogaheh ¹, Amir Teimourpour ², Mohsen Arzanlou ¹, Zahra Meshkat ^{3*}

¹ Department of Microbiology, School of Medicine, Ardabil University of Medical Sciences, Ardabil, Iran

² Department of Epidemiology and Biostatistics, School of Public Health, Tehran University of Medical science, Tehran, Iran

³ Antimicrobial Resistance Research Center, Mashhad University of Medical Sciences, Mashhad, Iran

ARTICLE INFO

Article type:

Original article

Article history:

Received: Apr 4, 2017

Accepted: Aug 10, 2017

Keywords:

BCG

DNA

Mycobacterium - tuberculosis

PCR

Plasmid

ABSTRACT

Objective(s): Tuberculosis (TB) has still remained a global health issue. One third of the world's population is infected with tuberculosis and the current BCG vaccine has low efficiency; hence, it is necessary to develop a new vaccine against TB. The aim of the current study was to evaluate the efficiency of a novel DNA vaccine encoding Mtb32C-HBHA antigen in inducing specific immune responses against *Mycobacterium tuberculosis*.

Materials and Methods: A DNA plasmid vaccine expressing Mtb32C-HBHA fusion protein was constructed and its ability in protein expression was examined by RT-PCR and Western blot methods. Female BALB/c mice were vaccinated with 100 µg of purified recombinant vector in an attempt to assess its immunogenicity and protective efficacy. Further, the cytokines, IFN-γ, IL-12, IL-4, IL-10, and TGF-β were assessed.

Results: The levels of all the studied cytokines were significantly increased ($P < 0.05$) compared with the control group. IFN-γ production in the group receiving DNA vaccine plus BCG was increased compared with those receiving only DNA vaccine or BCG ($P < 0.001$).

Conclusion: The immunogenicity of the new chimeric DNA vaccine was confirmed alone and in combination with BCG. Based on the results of the current study, the constructed DNA vaccine induced the expression of Mtb32C-HBHA fusion protein efficiently *in vitro*. Furthermore, high levels of the specific cytokines were induced in mice. By using this DNA vaccine as a booster after BCG, higher amounts of IFN-γ will be produced.

► Please cite this article as:

Teimourpour R, peeridogaheh H, Teimourpour A, Arzanlou M, Meshkat Z. A study on the immune response induced by a DNA vaccine encoding Mtb32C-HBHA antigen of *Mycobacterium tuberculosis*. Iran J Basic Med Sci 2017; 20:1119-1124. doi: 10.22038/IJBMS.2017.9445

Introduction

Tuberculosis (TB) is a life threatening disease caused by *Mycobacterium tuberculosis* and has remained a major global public health problem (1). About 9 million new cases of infection with TB and 2 million deaths due to TB are reported each year (2). Despite the relatively high efficiency of BCG vaccine, its effectiveness in the adult pulmonary system and population of endemic regions remains controversial. This necessitates the development of a new vaccine against TB with higher efficiency than the currently available BCG (3).

Recently, different vaccines such as *M. tuberculosis* mutants, recombinant BCG expressing various immunogenic antigens, subunit vaccines based on the highly immunogenic *M. tuberculosis* antigens, and DNA vaccine have been introduced (4-6). DNA vaccine is a simple way to introduce different antigens to the immune system, which is capable of inducing both cellular and humoral immune responses (7). Previous studies have shown that Mtb32C and HBHA are highly immunogenic antigens. HBHA is a mycobacterial

adhesin that binds to proteoglycan molecules on the surface of the lung epithelial cells (8, 9). HBHA participates in extrapulmonary dissemination of *M. tuberculosis* from lung to spleen. Previous studies have indicated that immunization with HBHA can induce a protection equal to that of BCG vaccination and can be used as a booster (10-12). Mtb32C is conserved in BCG and *M. tuberculosis*, and has specific epitopes (GAPINSATAM). Among MHC-1 restricted mycobacterial epitopes, Mtb32C can enhance CD8⁺ T-cell-dependent protective immunity and induce higher IFN-γ production (13, 14). In fact, IFN-γ has a key role in triggering cellular responses against intracellular pathogens such as *M. tuberculosis*. Consequently, it can be used as a specific antigen in IFN-γ production.

Prime-boost strategy is generally considered to be a promising way to enhance the efficacy of BCG (15). Using this strategy, the designated vaccine containing highly immunogenic mycobacterial antigens is administered to BCG pre-treated animals. This vaccination regimen induces higher levels of protection against TB compared

*Corresponding author: Zahra Meshkat. Antimicrobial Resistance Research Center, Mashhad University of Medical Sciences, Mashhad, Iran. Tel: +98-51-38012453; Fax: +98-51-38002287; email: meshkatz@mums.ac.ir

with BCG alone. In the current study, a DNA vaccine based on *mtb32C* and *hbha* genes was constructed, and its ability to stimulate the immune system alone and as a booster was evaluated in an animal model.

Materials and Methods

Pathogen-free female BALB/c mice between 6 and 8 weeks of age were obtained from Razi Vaccine and Serum Research Institute (Mashhad, Iran). All mice were kept under SPF conditions and maintained in accordance with the Ethical Policies of Mashhad University of Medical Sciences.

Plasmid construction

Gene isolation, cloning, fusion, and plasmid construction were performed previously (16, 17). Briefly, *Mtb32C* and *HBHA* genes were isolated from *M. tuberculosis* H37Rv genome and were cloned into pCDNA3.1 (+). Restriction enzyme analysis and DNA sequencing confirmed the accuracy of the cloning procedure.

Transfection, RT-PCR, and Western blot

To confirm the *in vitro* expression of *Mtb32C-HBHA* fusion genes, the plasmid was transfected into HeLa cells (American Type Culture Collection Manassas, VA, USA) using lipofectamine transfection reagent according to the manufacturer's instructions (Invitrogen, USA). Then, 72 hr after transfection, the cells were treated with 0.5 ml trypsin (Invitrogen, USA) and were incubated for 10 min. Cell suspension was harvested and total RNA was extracted using RNX-Plus (SinaClon, Iran), as described previously (16, 17). Purified RNA was used for cDNA synthesis using cDNA synthesis kit (Pars Tous, Iran) and was amplified by PCR. To detect the presence of His-Tag marker in chimeric *Mtb32C-HBHA* protein (a marker of constructed rather than natural protein), Western blot method was performed using mouse anti-His Tag antibody as the primary antibody and peroxidase conjugated rabbit anti-mouse IgG as the secondary antibody (AbD SeroTec, USA).

Vaccination

A total of 40 female BALB/c mice were randomly divided into these groups (10 mice in each group): control (pCDNA3.1+ vector), vaccine (pCDNA3.1+-*Mtb32C-HBHA*), BCG and BCG prime plus booster vaccine (pCDNA3.1+-*Mtb32C-HBHA*). The BCG and BCG prime plus booster vaccine groups were firstly immunized subcutaneously with BCG (5×10^5 CFU/PBS) (Pasteur institute, Iran) and then boosted three times intramuscularly with 100 μ g designated vaccine at two-week intervals. The control group and vaccine groups were immunized three times intramuscularly at two-week intervals with 100 μ g pCDNA3.1 (+) and recombinant vectors, respectively. Four weeks after the last immunization, all animals were sacrificed and their spleens were excised for further analysis (18).

Cytokine assay

Four weeks after the third vaccination, the spleen lymphocyte was extracted from all vaccinated and negative control mice. The 3×10^5 viable cells were cultured in 96-well microplates with RPMI 1640 medium (Invitrogen, USA) supplemented with 10% fetal calf serum (FCS), penicillin, and streptomycin in a total volume of 300 μ l.

Splenocytes extracted from BCG vaccinated mice were cultured with different concentrations (0.5-5 mg/ml) of heat-killed *M. tuberculosis* H37Rv (MTB antigen) and 10^4 - 10^5 CFU of live BCG bacterium (BCG antigen). The plates were incubated at 37 °C in a humidified incubator with 5% CO₂ for 72 hr. After the stimulation, the supernatants were harvested and IFN- γ release assay was performed using ELISA kits (eBioscience, SanDiego, CA) according to the manufacturer's instructions. The maximum production of IFN- γ was seen in 0.5 mg/ml of MTB antigen and 2×10^4 CFU of BCG antigen in PBS. Based on these results, all splenocytes extracted from all groups were stimulated with these antigen concentrations (19, 20).

The freshly isolated splenocytes were incubated with MTB and BCG antigens for three days. At the end of day three, cell supernatants (after centrifugation) were used for subsequent analysis.

The concentrations of IFN- γ , interleukin-4 (IL-4), IL-12, TGF- β , and IL-10 in the culture supernatants were evaluated by an ELISA kit (eBioscience, SanDiego, CA) following the manufacturer's recommendations. Assay sensitivity levels for IFN- γ , IL-12p70, IL-4, and IL-10 were 15 pg/ml, 15 pg/ml, 4 pg/ml, 30 pg/ml, and 8 pg/ml, respectively. The data were expressed as mean \pm standard deviation for each group of mice (eBioscience, SanDiego, CA).

Statistical analysis

The normality of the acquired data was assessed by Kolmogorov-Smirnov test. All data were analyzed by SPSS19.0 software using two-way ANOVA, and $P < 0.05$ was considered statistically significant.

Results

RT-PCR and Western blot to assess the expression of DNA construct

Expression of the constructed DNA vaccine was first confirmed by RT-PCR to detect mRNA production, and was followed by Western blot to detect the recombinant *Mtb32C-HBHA* protein. The constructed *Mtb32C-HBHA* fusion protein was successfully expressed in the eukaryotic system. The amplified fragment using *Mtb32C* and *HBHA* specific primers demonstrated that recombinant vector could be transcribed in HeLa cell line. To assess the ability of vector in producing recombinant protein in cell culture system, Western blot technique and staining with specific antibodies and ECL detection reagent were used (abCam, UK) (Figures 1 and 2).

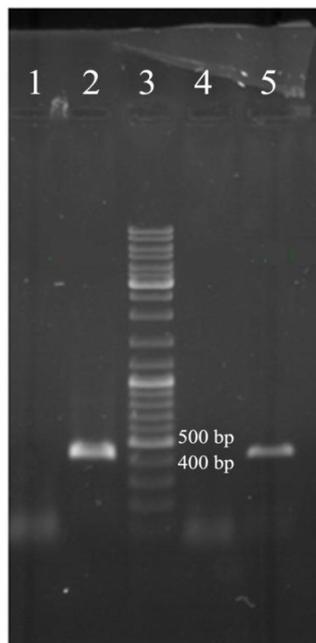


Figure 1. RT-PCR results on cDNA from transfected HeLa cells (lane numbers 2 and 5) and non-transfected cells (lane numbers 1 and 4); lane 3: 1 kb DNA size marker (Fermentas, Germany)

Cytokine assay

Four weeks after the last immunization, the mice were sacrificed, and specific cytokine production (IFN γ , IL-10, TGF- β , IL-12 and IL-4) was evaluated by ELISA in the supernatant of the splenocytes cultures after *in vitro* stimulation with MTB (0.5 mg/ml) and BCG antigens (2×10^4 CFU) for 72 hr. All mice vaccinated with Mtb32C-HBHA as a DNA vaccine, BCG and BCG-prime boosted with Mtb32C-HBHA (in the absence of adjuvant) produced high levels of IFN γ , IL-10, TGF- β , IL-12, and IL-4 compared to the control group of mice vaccinated only with pCDNA3.1 vector ($P < 0.05$).

Significant differences (at significance level of $P < 0.001$) were observed in all vaccinated groups (BCG, DNA vaccine, and BCG-vaccine) in the four examined cytokines in comparison with those in the control group.

As shown in Figure 3 and Table 1, significant differences were found between BCG-vaccine and vaccine and control groups at the significance level $P < 0.001$, while the difference between BCG and control group was at $P < 0.05$. The BCG-vaccinated mice boosted with Mtb32C-HBHA had a higher level of IFN γ than other groups. Splenocytes from BCG-primed mice boosted with Mtb32C-HBHA produced substantially higher IFN γ (265.73 \pm 119 pg/ml) than BCG-vaccinated mice (85.24 \pm 54 pg/ml), Mtb32C-HBHA vaccinated mice (144.78 \pm 37 pg/ml), and control mice (25.57 \pm 13 pg/ml). Significant differences

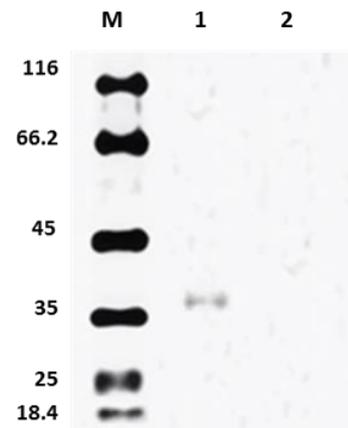


Figure 2. Western blot analysis of cell lysate from HeLa cells transfected with pCDNA3.1-Mtb32C-HBHA construct (lane 1) and untransfected cells (lane 2); lane M: protein size marker (Fermentas Company, Germany)

between BCG-vaccine group and BCG and vaccine groups were at $P < 0.01$, while the difference between BCG and vaccine group was at $P < 0.05$. Also, IFN γ cytokine was increased in all groups except the control group. The mice vaccinated with BCG plus Mtb32C-HBHA induced the highest levels of IFN γ responses in the spleen. The combination of Mtb32C-HBHA and BCG significantly increased the expression of IFN γ compared with BCG or Mtb32C-HBHA alone.

The amount of IL-12 in the group receiving Mtb32C-HBHA vaccine (31.98 \pm 7.45) was higher than that of BCG (24.78 \pm 6.79), BCG-vaccine (28.84 \pm 7.54), and control (6.54 \pm 2.18) groups. Significant differences were observed only between BCG and vaccine group at $P < 0.01$.

In the case of IL-4 and TGF- β , no statistically significant differences were seen between BCG, BCG-vaccine and vaccine groups ($P > 0.05$), but there were significant differences between control group and other groups. IL-4 production levels of BCG (8.53 \pm 4.2), BCG-vaccine (8.58 \pm 2.8) and vaccine (9.65 \pm 2.8) groups were higher than that of the control (3.64 \pm .93) group.

As shown in Figure 3 and Table 1, splenocytes from the mice receiving BCG (8.53 \pm 1.88), BCG-vaccine (8.22 \pm 1.41) and vaccine (9.93 \pm 4.18) showed higher TGF- β production than control (3.06 \pm 1.18) group and the group receiving just pCDNA3.1(+) vector.

The amount of IL-10 was significantly higher in BCG-vaccine group (1142.43 \pm 577) than in BCG (825.64 \pm .454), vaccine (783.84 \pm 394), and control (289.88 \pm 67.54) groups. Furthermore, there were no significant differences between BCG and vaccine groups ($P > 0.05$), but significant differences were observed between BCG-vaccine group and other groups, including BCG and vaccine groups at $P < 0.01$.

Table 1. Spleen cytokine profile in different groups of BALB/c mice

Cytokine	IL-12p70	IL-4	IL-10	IFN- γ	TGF- β
Vaccination groups					
Control	6.54 \pm 2.18	3.64 \pm .93	289.88 \pm 76.54	25.57 \pm 13	3.06 \pm 1.18
Vaccine	31.98 \pm 7.45	9.65 \pm 2.8	783.84 \pm 394	144.78 \pm 37	9.93 \pm 4.18
BCG	24.78 \pm 6.79	8.53 \pm 4.2	825.64 \pm .454	85.24 \pm 54	8.53 \pm 1.88
BCG-vaccine	28.84 \pm 7.5	8.58 \pm 2.8	1142.43 \pm .577	265.73 \pm 119	8.22 \pm 1.41

Four weeks after the third Mtb32C-HBHA injection, ten mice from each group were sacrificed and their spleen cells were cultured in the presence of MTB and BCG antigens. Levels of cytokines were measured in a 72 hr culture supernatant. Values are expressed in pg/ml and represented as mean \pm standard deviation. The mice were primed with BCG and then boosted with 100 μ g Mtb32C- HBHA (BCG-HBHA) subcutaneously

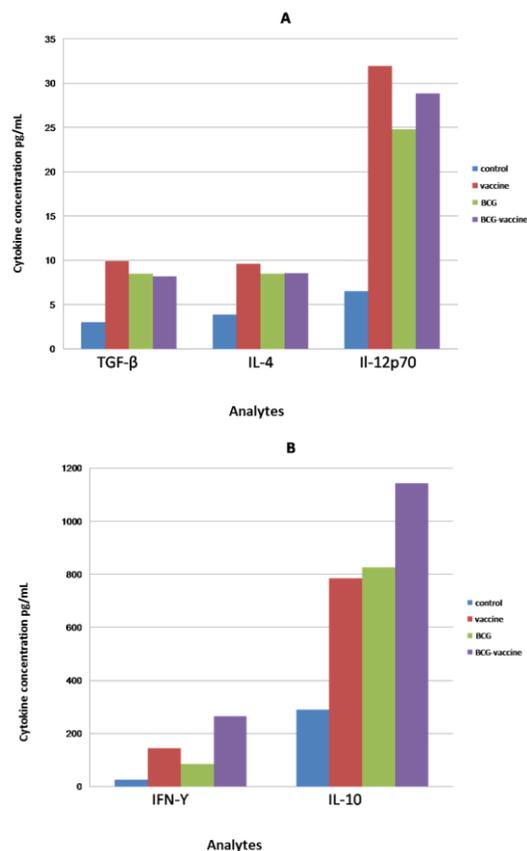


Figure 3. The immune response to Mtb32C-HBHA after the last vaccination. The mice were divided into four groups and were immunized with BCG alone (BCG group), BCG/Mtb32C-HBHA (vaccine +BCG group), Mtb32C-HBHA alone (vaccine group), and empty vector (control group)

The immune responses for IFN- γ (B), IL-10(B), IL-12(A), IL-4 (A), and TGF- β (A) after stimulation were quantified by ELISA. The results are expressed as mean \pm standard deviation for the mice per group, tested in triplicates. The two-way ANOVA test was used to analyze the results, and significant difference was set as $P < 0.05$

Discussion

The goals of WHO regarding TB infection are 50% reduction in TB mortality by 2015 and reduction of the incidence of new cases to less than one in a million people by 2050. These objectives are expected to be achieved to provide at least partially new drugs and more effective vaccines (21, 22). Producing more

effective vaccines than BCG is a critical step in global TB control. Nevertheless, replacing BCG vaccine with other vaccines is not rational because BCG is more effective against severe forms of childhood TB. DNA vaccine has been considered an alternative candidate for inducing an immune response against TB. In animal models, DNA vaccine has provided satisfactory results, whereas DNA vaccines in human have comparatively lower immunogenicity. This defect can be resolved in the prime-boost strategy (4, 23). In this study, the constructed DNA vaccine was used alone and in combination with BCG in prime-boost strategy. Rouanet *et al.* showed that HBHA antibody titers were low during BCG vaccination. This indicates that despite the presence of *hbha* gene in BCG strain, it cannot be expressed. Therefore, the efficacy of BCG vaccine can be elevated by adding HBHA protein (24).

Administration of pCDNA-Mtb32C-HBHA vaccine showed that it was safe and well tolerated. Other studies have provided evidence that HBHA can be used as a booster vaccine, and methylation in the C-terminal domain of mycobacterial HBHA plays an important role in T-cell stimulation (11, 25). This study showed that Mtb32C-HBHA without post-translational modifications is able to induce high levels of IFN- γ as a cellular immune response marker (13). Indeed, Mtb32C could compensate this deficiency and could enhance HBHA immunogenicity (14). Previous studies have shown that HBHA in the presence of a specific Th1 adjuvant is able to induce strong cellular immune responses against *M. tuberculosis*, identical to that of BCG (32-34). In the present study, the ability of pCDNA3.1-Mtb32C-HBHA vector to stimulate immune responses was evaluated in the absence of adjuvant. Mtb32C-HBHA fusion protein without C-terminal methylation and in the absence of an adjuvant could stimulate strong cellular immune responses in BALB/c mice. Mtb32C-HBHA induced a robust Th1 response that is characterized by elevated levels of IL-12, IFN- γ , and TGF- β in contrast to low levels of IL-4. The findings of this research indicated that pCDNA-Mtb32C-HBHA construct could be used in heterologous prime boost regimen because in this strategy other cytokines which are necessary

for development, magnitude and maintenance of T cells such as IL-12 and TGF- β were produced. This result is also consistent with the results of the study performed by Guerrero *et al.* in which high levels of IL-10 were observed (32, 33). Another study concluded that the absence of methylation pattern in HBHA affects T-reg cells function, but others suggested that after induction, Th1 cells are the main source of IL-10 production. Cellular immune response has a critical role against intracellular pathogens, including *M. tuberculosis*. Protection against TB is dependent on cellular immunity, so IFN- γ and IL-12 as intracellular pathogens have a pivotal role in protection against *M. tuberculosis* (26-28). Consistent with the results of previous studies, boosting Mtb32C-HBHA vaccination following BCG priming exhibited significant increase in IFN- γ and IL-12 levels compared to other groups. Previous studies have reported that elevation of TGF- β in the presence of IL-10 and IL-12 stimulates T-reg and memory T cells, respectively. Also in our study, all these cytokines were elevated compared to control group (29). Spleen lymphocytes from all groups, which were stimulated by both antigens, produced significant amounts of IL12p70. High production of IL12p70 was observed in the group receiving only the constructed vaccine and in the group immunized by BCG- Mtb32C-HBHA vaccine in the prime-boost strategy. The highest rate of IL-12 was observed in the vaccine group, which indicated the designated vaccine can efficiently be expressed in the host cells and has the ability to promote antigen-presenting cells (APCs). Elevation of IL-12 along with TGF- β can trigger responses characterizing the effectors-memory T cells (30, 31). This study showed that administration of BCG in combination with DNA vaccine induced stronger Th1 immune response because IFN- γ was produced at greater levels than IL-4 by spleen lymphocyte upon *in vitro* stimulation. The Th1-type immune response has a key role in host response to *M. tuberculosis* infection. The obtained results strongly support this idea that vaccination with pCDNA3.1-Mtb32C-HBHA plasmid and use of prime-boost regimen enhance the immune response, which is turned toward Th1 type. In Prime-boost strategy, employing DNA vaccines has been shown to elicit stronger and more diverse cellular immune responses than BCG vaccine alone.

Conclusion

It was shown that IFN- γ production in the mice primed with BCG and then boosted with DNA vaccine was more remarkable than those vaccinated only with BCG or vaccine. Indeed, this construct can induce not only high levels of IFN- γ but also other cytokines such as IL-12 and TGF- β . The profile of cytokine production by spleen cells showed that the vaccine is so strong that can stimulate the immune system. These findings

indicate that prime-boost vaccination scheme is an efficient vaccination model against TB.

Acknowledgment

The results described in this paper were part of a PhD student thesis (Thesis No.454-A). This study was financially supported by Mashhad University of Medical Sciences, Mashhad, Iran (Grant No. 910016).

References

1. Brewer TF. Preventing tuberculosis with bacillus Calmette-Guerin vaccine: a meta-analysis of the literature. *Clin Infect Dis* 2000; 3:S64-67.
2. Dara M, Dadu A, Kremer K, Zaleskis R, Kluge HH. Epidemiology of tuberculosis in WHO European Region and public health response. *Eur Spine J* 2013; 22:549-555.
3. Colditz GA, Brewer TF, Berkey CS, Wilson ME, Burdick E, Fineberg HV, *et al.* Efficacy of BCG vaccine in the prevention of tuberculosis. Meta-analysis of the published literature. *JAMA* 1994; 271:698-702.
4. Delogu G, Fadda G. The quest for a new vaccine against tuberculosis. *J Infect Dev Ctries* 2009; 3:5-15.
5. Dietrich G, Viret JF, Hess J. Mycobacterium bovis BCG-based vaccines against tuberculosis: novel developments. *Vaccine* 2003; 21:667-670.
6. Parida SK, Kaufmann SH. Novel tuberculosis vaccines on the horizon. *Curr Opin Immunol* 2010; 22:374-384.
7. Ghanem A, Healey R, Adly FG. Current trends in separation of plasmid DNA vaccines: a review. *Anal Chim Acta* 2013; 760:1-15.
8. Pethe K, Aumercier M, Fort E, Gatot C, Loch C, Menozzi FD. Characterization of the heparin-binding site of the mycobacterial heparin-binding hemagglutinin adhesin. *J Biol Chem* 2000; 275:14273-14280.
9. Vidal Pessolani MC, Marques MA, Reddy VM, Loch C, Menozzi FD. Systemic dissemination in tuberculosis and leprosy: do mycobacterial adhesins play a role? *Microbes Infect* 2003; 5:677-684.
10. Krishnan N, Robertson BD, Thwaites G. The mechanisms and consequences of the extra-pulmonary dissemination of Mycobacterium tuberculosis. *Tuberculosis (Edinb)* 2010; 90:361-366.
11. Guerrero GG, Debie AS, Loch C. Boosting with mycobacterial heparin-binding haemagglutinin enhances protection of Mycobacterium bovis BCG-vaccinated newborn mice against M. tuberculosis. *Vaccine* 2010; 28:4340-4347.
12. Loch C, Hougardy JM, Rouanet C, Place S, Mascart F. Heparin-binding hemagglutinin, from an extrapulmonary dissemination factor to a powerful diagnostic and protective antigen against tuberculosis. *Tuberculosis (Edinb)* 2006; 86:303-309.
13. Irwin SM, Izzo AA, Dow SW, Skeiky YA, Reed SG, Alderson MR, *et al.* Tracking antigen-specific CD8 T lymphocytes in the lungs of mice vaccinated with the Mtb72F polyprotein. *Infect Immun* 2005; 73:5809-5816.
14. Skeiky YA, Alderson MR, Owendale PJ, Guderian JA, Brandt L, Dillon DC, *et al.* Differential immune responses and protective efficacy induced by components of a tuberculosis polyprotein vaccine, Mtb72F, delivered as naked DNA or recombinant protein. *J Immunol* 2004; 172:7618-7628.

15. Romano M, D'Souza S, Adnet PY, Laali R, Jurion F, Palfliet K, *et al.* Priming but not boosting with plasmid DNA encoding mycolyl-transferase Ag85A from *Mycobacterium tuberculosis* increases the survival time of *Mycobacterium bovis* BCG vaccinated mice against low dose intravenous challenge with *M. tuberculosis* H37Rv. *Vaccine* 2006; 24:3353-3364.
16. Teimourpour R, Sadeghian A, Meshkat Z, Esmaelizad M, Sankian M, Jabbari AR. Construction of a DNA vaccine encoding *Mtb32C* and *HBHA* genes of *mycobacterium tuberculosis*. *Jundishapur J Microbiol* 2015; 8:e21556.
17. Teimourpour R, Zare H, Rajabnia R, Yahyapoor Y, Meshkat Z. Evaluation of the eukaryotic expression of *mtb32C-hbha* fusion gene of *Mycobacterium tuberculosis* in Hepatocarcinoma cell line. *Iran J Microbiol* 2016; 8:132-138.
18. Meshkat Z, Teimourpour A, Rashidian S, Arzanlou M, Teimourpour R. Immunogenicity of a DNA Vaccine Encoding Ag85a-Tb10. 4 antigens from *mycobacterium tuberculosis*. *Iran J Immunol* 2016; 13:289.
19. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 1976; 72:248-254.
20. Meshkat Z, Mirshahabi H, Meshkat M, Kheirandish M, Hassan ZM. Strong immune responses induced by a DNA vaccine containing HPV16 truncated E7 C-terminal linked to HSP70 gene. *Iran J Immunol* 2011; 8:65.
21. Ottenhoff TH, Kaufmann SH. Vaccines against tuberculosis: where are we and where do we need to go? *PLoS Pathogens* 2012; 8:e1002607.
22. Martin C. Tuberculosis vaccines: past, present and future. *Curr Opin Pulm Med* 2006; 12:186-191.
23. Feng CG PU, Demangel C, Spratt JM, Malin AS, Britton WJ. Priming by DNA immunization augments protective efficacy of *Mycobacterium bovis* Bacille Calmette-Guerin against tuberculosis. *Infect Immun* 2001; 69:4174-4176.
24. Rouanet C, Lecher S, Loch C. Subcutaneous boosting with heparin binding haemagglutinin increases BCG-induced protection against tuberculosis. *Microbes Infect* 2009; 11:995-1001.
25. Guerrero GG, Loch C. Recombinant HBHA boosting effect on BCG-induced immunity against *Mycobacterium tuberculosis* infection. *Clin Dev Immunol* 2011; 2011:730702.
26. Barker LF, Brennan MJ, Rosenstein PK, Sadoff JC. Tuberculosis vaccine research: the impact of immunology. *Curr Opin Immunol* 2009; 21:331-338.
27. Cavalcanti YV, Brelaz MC, Neves JK, Ferraz JC, Pereira VR. Role of TNF-Alpha, IFN-Gamma, and IL-10 in the Development of Pulmonary Tuberculosis. *Pulm Med* 2012; 2012:10.
28. Redford P, Murray P, O'Garra A. The role of IL-10 in immune regulation during *M. tuberculosis* infection. *Mucosal Immunol* 2011; 4:261-270.
29. Jung YJ, Ryan L, LaCourse R, North RJ. Increased interleukin-10 expression is not responsible for failure of T helper 1 immunity to resolve airborne *Mycobacterium tuberculosis* infection in mice. *Immunology* 2003; 109:295-299.
30. Wyndham-Thomas C, Corbiere V, Dirix V, Smits K, Domont F, Libin M, *et al.* Key role of effector memory CD4+ T lymphocytes in a short-incubation heparin-binding hemagglutinin gamma interferon release assay for the detection of latent tuberculosis. *Clin Vaccine Immunol* 2014; 21:321-328.
31. Shafiani S DC, Ertelt JM, Moguche AO, Siddiqui I, Smigiel KS, *et al.* Pathogen-specific Treg cells expand early during *mycobacterium tuberculosis* infection but are later eliminated in response to Interleukin-12. *J Exp Med* 2013; 206:1261-1270.
32. Guerrero GG, Loch C. Recombinant HBHA boosting effect on BCG-induced immunity against *Mycobacterium tuberculosis* infection. *Clin Dev Immunol* 2011; 2011:730702.
33. Guerrero GG, Feunou FP, Loch C. The coiled-coil N-terminal domain of the heparin-binding haemagglutinin is required for the humoral and cellular immune responses in mice. *Mol Immunol* 2008; 46:116-24.
34. Lebrun P, Raze D, Fritzinger B, Wieruszski J-M, Biet F, Dose A, *et al.* Differential contribution of the repeats to heparin binding of HBHA, a major adhesin of *Mycobacterium tuberculosis*. *PLoS One* 2012; 7:e32421.